

(12) INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(19) World Intellectual Property Organization
International Bureau



(43) International Publication Date
25 May 2001 (25.05.2001)

PCT

(10) International Publication Number
WO 01/36684 A2

(51) International Patent Classification⁷: **C12Q 1/68**

(21) International Application Number: PCT/US00/31743

(22) International Filing Date:
16 November 2000 (16.11.2000)

(25) Filing Language: English

(26) Publication Language: English

(30) Priority Data:
09/443,184 19 November 1999 (19.11.1999) US

(63) Related by continuation (CON) or continuation-in-part (CIP) to earlier application:
US 09/443,184 (CIP)
Filed on 19 November 1999 (19.11.1999)

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(81) Designated States (national): AE, AG, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, BZ, CA, CH, CN, CR, CU, CZ, DE, DK, DM, DZ, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MA, MD, MG, MK, MN, MW, MX, MZ, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, TZ, UA, UG, US, UZ, VN, YU, ZA, ZW.

(84) Designated States (regional): ARIPO patent (GH, GM, KE, LS, MW, MZ, SD, SL, SZ, TZ, UG, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE, TR), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, GW, ML, MR, NE, SN, TD, TG).

Published:

— Without international search report and to be republished upon receipt of that report.

For two-letter codes and other abbreviations, refer to the "Guidance Notes on Codes and Abbreviations" appearing at the beginning of each regular issue of the PCT Gazette.

(54) Title: MAMMALIAN TOXICOLOGICAL RESPONSE MARKERS

(57) Abstract: The present invention relates to mammalian nucleic acid and protein molecules comprising a plurality of nucleic acid and protein molecules. The mammalian nucleic acid molecules can be used as hybridizable array elements in a microarray in diagnostic and therapeutic applications including detecting metabolic and toxicological responses, and in monitoring drug mechanism of action. The protein molecules can be used in a pharmaceutical composition. The present invention also relates to methods for screening compounds and therapeutics for metabolic responses indicative of a toxic compound or molecule.

WO 01/36684 A2

MAMMALIAN TOXICOLOGICAL RESPONSE MARKERS

This application is filed under the Patent Cooperation Treaty and claims the benefit of U.S. Nonprovisional Application No. 09/443,184, our Docket No. PC-0007 US, filed 19th November, 1999.

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TECHNICAL FIELD

The present invention relates to mammalian nucleic acid and protein molecules, and methods for their use in diagnostic and therapeutic applications including detecting metabolic and toxicological responses, and in monitoring drug mechanism of action.

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BACKGROUND ART

Toxicity testing is a mandatory and time-consuming part of drug development programs in the pharmaceutical industry. A more rapid screen to determine the effects upon metabolism and to detect toxicity of lead drug candidates may be the use of gene expression microarrays. For example, microarrays of various kinds may be produced using full length genes or gene fragments. These arrays can then be used to test samples treated with the drug candidates to elucidate the gene expression pattern associated with drug treatment. This gene pattern can be compared with gene expression patterns associated with compounds which produce known metabolic and toxicological responses.

Benzo(a)pyrene is a known rodent and likely human carcinogen and is the prototype of a class of compounds, the polycyclic aromatic hydrocarbons (PAH). It is metabolized by several forms of cytochrome P450 (P450 isozymes) and associated enzymes to form both activated and detoxified metabolites. The ultimate metabolites are the bay-region diol epoxide, benzo(a)pyrene-7,8-diol-9,10-epoxide (BPDE) and the K-region diol epoxide, 9-hydroxy benzo(a)pyrene-4,5-oxide, both of which induce formation of DNA adducts. DNA adducts have been shown to persist in rat liver up to 56 days following treatment with benzo(a)pyrene at a dose of 10 mg/kg body weight three times per week for two weeks (Qu and Stacey (1996) Carcinogenesis 17:53-59).

Acetaminophen is a widely-used analgesic. It is metabolized by specific cytochrome P450 isozymes with the majority of the drug undergoing detoxification by glucuronic acid, sulfate and glutathione conjugation pathways. However, at supratherapeutic doses, acetaminophen is metabolized to an active intermediate, *N*-acetyl-*p*-benzoquinone imine (NAPQI) which can cause hepatic and renal failure. NAPQI then binds to sulfhydryl groups of proteins causing their inactivation and leading to subsequent cell death (Kroger *et al.* (1997) Gen. Pharmacol. 28:257-263).

Clofibrate is an hypolipidemic drug which lowers elevated levels of serum triglycerides. In rodents, chronic treatment produces hepatomegaly and an increase in hepatic peroxisomes (peroxisome

proliferation). Peroxisome proliferators (PPs) are a class of drugs which activate the PP-activated receptor in rodent liver, leading to enzyme induction, stimulation of S-phase, and a suppression of apoptosis (Hasmall and Roberts (1999) Pharmacol. Ther. 82:63-70). PPs include the fibrate class of hypolipidemic drugs, phenobarbitone, thiazolidinediones, certain non-steroidal anti-inflammatory drugs, and naturally-occurring fatty acid-derived molecules (Gelman *et al.* (1999) Cell. Mol. Life Sci. 55:932-943). Clofibrate has been shown to increase levels of cytochrome P450 4A. It is also involved in transcription of β -oxidation genes as well as induction of PP-activated receptors (Kawashima *et al.* (1997) Arch. Biochem. Biophys. 347:148-154). Peroxisome proliferation that is induced by both clofibrate and the chemically-related compound fenofibrate is mediated by a common inhibitory effect on mitochondrial membrane depolarization (Zhou and Wallace (1999) Toxicol. Sci. 48:82-89).

Toxicological effects in the liver are also induced by other compounds. These can include carbon tetrachloride (a necrotic agent), hydrazine (a steatotic agent), α -naphthylisothiocyanate (a cholestatic agent), 4-acetylaminofluorene (a liver mitogen), and their corresponding metabolites, which are used in experimental protocols to measure toxicological responses (Waterfield *et al.* (1993) Arch. Toxicol. 67:244-254).

The present invention provides mammalian nucleic acid and protein molecules, their use in diagnostic and therapeutic applications including detecting metabolic and toxicological responses, and in monitoring drug mechanism of action.

DISCLOSURE OF INVENTION

The invention provides a method for detecting or diagnosing the effect of a test compound or molecule associated with increased or decreased levels of nucleic acid molecules in a mammalian subject. The method comprises treating a mammalian subject with a known toxic compound or molecule which elicits a toxicological response, measuring levels of a plurality of nucleic acid molecules, selecting from the plurality of nucleic acid molecules those nucleic acid molecules that have levels modulated in samples treated with known toxic compounds or molecules when compared with untreated samples. Some of the levels may be upregulated by a toxic compound or molecule, others may be downregulated by a toxic compound or molecule, and still others may be upregulated with one known toxic compound or molecule and be downregulated with another known toxic compound or molecule. The selected nucleic acid molecules which are upregulated and downregulated by a known toxic compound or molecule are arrayed upon a substrate. The method further comprises measuring levels of nucleic acid molecules in the sample after the sample is treated with the toxic compound or molecule. Levels of nucleic acid molecules in a sample so treated are then compared with the plurality of the arrayed nucleic acid molecules to identify which sample nucleic acid molecules are upregulated and downregulated by the test

compound or molecule. In one embodiment, the nucleic acid molecules are hybridizable array elements of a microarray.

Preferably, the comparing comprises contacting the arrayed nucleic acid molecules with the sample nucleic acid molecules under conditions effective to form hybridization complexes between the arrayed nucleic acid molecules and the sample nucleic acid molecules; and detecting the presence or absence of the hybridization complexes. In this context, similarity may mean that at least 1, preferably at least 5, more preferably at least 10, of the upregulated arrayed nucleic acid molecules form hybridization complexes with the sample nucleic acid molecules at least once during a time course to a greater extent than would the probes derived from a sample not treated with the test compound or molecule or a known toxic compound or molecule. Similarity may also mean that at least 1, preferably at least 5, more preferably at least 10, of the downregulated arrayed nucleic acid molecules form hybridization complexes with the sample nucleic acid molecules at least once during a time course to a lesser extent than would the sample nucleic acid molecules of a sample not treated with the test compound or a known toxic compound. In one aspect, the arrayed nucleic acid molecules comprise SEQ ID NOs:1-47 or fragments thereof.

Preferred toxic compounds are selected from the group consisting of hypolipidemic drugs, n-alkylcarboxylic acids, n-alkylcarboxylic acid precursors, azole antifungal compounds, leukotriene D4 antagonists, herbicides, pesticides, phthalate esters, phenyl acetate, dehydroepiandrosterone (DHEA), oleic acid, methanol and their corresponding metabolites, acetaminophen and its corresponding metabolites, benzo(a)pyrene, 3-methylcholanthrene, benz(a)anthracene, 7,12-dimethylbenz(a)anthracene, their corresponding metabolites, and the like, carbon tetrachloride, hydrazine, α -naphthylisothiocyanate, 4-acetylaminofluorene, and their corresponding metabolites. Preferred tissues are selected from the group consisting of liver, kidney, brain, spleen, pancreas and lung.

The arrayed nucleic acid molecules comprise fragments of messenger RNA transcripts of genes that are upregulated-or-downregulated at least 2-fold, preferably at least 2.5-fold, more preferably at least 3-fold, in tissues treated with known toxic compounds when compared with untreated tissues. Preferred arrayed nucleic acid molecules are selected from the group consisting of SEQ ID NOs:1-47 or fragments thereof, some of whose expression is upregulated following treatment with a toxic compound or molecule and others of whose expression is downregulated following treatment with a toxic compound or molecule. More preferable are SEQ ID NOs:2, 4, 6, 8, 9, and 11 which are upregulated following treatment with a toxic compound or molecule, and SEQ ID NOs:1, 4, and 7 which are downregulated following treatment with a toxic compound or molecule.

The invention also provides a method comprising measuring levels of nucleic acid molecules in a sample after the sample is treated with a test compound or molecule. Levels of nucleic acid molecules in

a sample so treated are then compared with the plurality of the arrayed nucleic acid molecules to identify which sample nucleic acid molecules are upregulated and downregulated by the test compound or molecule. In one embodiment, the nucleic acid molecules are hybridizable array elements of a microarray.

5 Alternatively, the invention provides methods for screening a sample for a metabolic response to a test compound or molecule.

 Alternatively, the invention provides methods for screening a test compound or molecule for a previously unknown metabolic response.

 In another aspect, the invention provides methods for preventing a toxicological response by
10 administering complementary nucleotide molecules against one or more selected upregulated nucleic acid molecules or a ribozyme that specifically cleaves such molecules. Alternatively, a toxicological response may be prevented by administering sense nucleotide molecules for one or more selected downregulated nucleic acid molecules.

 In yet another aspect, the invention provides methods for preventing a toxicological response by
15 administering an agonist which initiates transcription of a gene comprising a downregulated nucleic acid molecule of the invention. Alternatively, a toxicological response may be prevented by administering an antagonist which prevents transcription of a gene comprising an upregulated nucleic acid molecule of the invention.

 In another aspect, the invention provides nucleic acid molecules whose transcript levels are
20 modulated in a sample during a metabolic response to a toxic compound or molecule. The invention also provides nucleic acid molecules whose transcript levels are upregulated in a sample during a metabolic response to a toxic compound or molecule. The invention also provides nucleic acid molecules whose transcript levels are downregulated in a sample during a metabolic response to a toxic compound or molecule. Upregulation or downregulation is at least 2-fold, more preferably at least 2.5-fold, even more
25 preferably at least 3-fold. The metabolic response to a toxic compound or molecule may be a toxicological response. The invention also provides mammalian nucleic acid molecules which are homologous to the upregulated and downregulated nucleic acid molecules. In one aspect, preferred arrayed nucleic acid molecules are selected from the group consisting of SEQ ID NOs:1-47, or fragments thereof.

30 The invention also provides a method for using a molecule selected from SEQ ID NOs:1-59 or a portion thereof to screen a library of molecules to identify at least one ligand which specifically binds the selected molecule, the method comprising combining the selected molecule with the library of molecules under conditions allowing specific binding, and detecting specific binding, thereby identifying a ligand which specifically binds the selected molecule.

Such libraries include DNA and RNA molecules, peptides, peptide nucleic acids, agonists, antagonists, antibodies, immunoglobulins, drug compounds, pharmaceutical agents, and other ligands. In one aspect, the ligand identified using the method modulates the activity of the selected molecule. In an analogous method, the selected molecule or a portion thereof is used to purify a ligand. The method involves combining the selected molecule or a portion thereof with a sample under conditions to allow specific binding, detecting specific binding between the selected molecule and ligand, recovering the bound selected molecule, and separating the selected molecule from the ligand to obtain purified ligand. The invention further provides a method for using at least a portion of the proteins encoded by SEQ ID NOs:1-47 and the proteins of SEQ ID NOs:48-59 to produce antibodies.

The invention further provides a method for inserting a marker gene into the genomic DNA of an animal to disrupt the expression of the natural nucleic acid molecule. The invention also provides a method for using the nucleic acid molecule to produce an animal model system, the method comprising constructing a vector containing the nucleic acid molecule; introducing the vector into a totipotent embryonic stem cell; selecting an embryonic stem cell with the vector integrated into genomic DNA; microinjecting the selected cell into a blastocyst, thereby forming a chimeric blastocyst; transferring the chimeric blastocyst into a pseudopregnant dam, wherein the dam gives birth to a chimeric animal containing at least one additional copy of nucleic acid molecule in its germ line; and breeding the chimeric animal to generate a homozygous animal model system.

The invention also provides a substantially purified mammalian protein or a portion thereof. The invention further provides isolated and purified proteins encoded by the nucleic acid molecules of SEQ ID NOs:1-11, 17-33, 36, 39, and 41. The invention further provides isolated and purified protein molecule of SEQ ID NOs:50 and 53. Additionally, the invention provides a pharmaceutical composition comprising a substantially purified mammalian protein or a portion thereof in conjunction with a pharmaceutical carrier.

The invention further provides an isolated and purified mammalian nucleic acid molecule variant having at least 70% nucleic acid sequence identity to the mammalian nucleic acid molecule selected from SEQ ID NO:1-47 and fragments thereof. The invention also provides an isolated and purified nucleic acid molecule having a sequence which is complementary to the mammalian nucleic acid molecule comprising a nucleic acid molecule selected from SEQ ID NO:1-47 and fragments thereof.

The invention further provides an expression vector containing at least a fragment of the mammalian nucleic acid molecule selected from the group consisting of SEQ ID NOs:1-47. In another aspect, the expression vector is contained within a host cell.

The invention also provides a method for producing a mammalian protein, the method comprising the steps of: (a) culturing the host cell containing an expression vector containing a

mammalian nucleic acid molecule of the invention under conditions suitable for the expression of the polypeptide; and (b) recovering the polypeptide from the host cell culture.

The invention also provides a pharmaceutical composition comprising a substantially purified mammalian protein encoded by SEQ ID NOs:1-11, 17-33, 36, 39, and 41 and the amino acid sequence of SEQ ID NOs:50 and 53 and fragments thereof, in conjunction with a suitable pharmaceutical carrier.

The invention further includes an isolated and purified antibody which binds to a mammalian protein encoded by SEQ ID NOs:1-11, 17-33, 36, 39, and 41 and mammalian protein of SEQ ID NOs:50 and 53 or fragments thereof. The invention also provides a purified agonist and a purified antagonist.

BRIEF DESCRIPTION OF THE SEQUENCE LISTING

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The Sequence Listing contains the nucleic acid sequence of exemplary mammalian nucleic acid molecules of the invention, SEQ ID NOs:1-47, 60-135, 137, and 138; the protein sequence of exemplary mammalian protein molecules of the invention, SEQ ID NOs:48-59 and 136.

MODES FOR CARRYING OUT THE INVENTION

Definitions

"Sample" is used in its broadest sense. A sample containing nucleic acid molecules may comprise a bodily fluid; a cell; an extract from a cell, chromosome, organelle, or membrane isolated from a cell; genomic DNA, RNA, or cDNA in solution or bound to a substrate; a biological tissue or biopsy thereof; a fingerprint or tissue print; natural or synthetic fibres; in a solution; in a liquid suspension; in a gaseous suspension; in an aerosol; and the like.

"Plurality" refers preferably to a group of one or more members, preferably to a group of at least about 10, and more preferably to a group of at least about 100 members, and even more preferably a group of 10,000 members.

"Substrate" refers to a rigid or semi-rigid support to which nucleic acid molecules or proteins are bound and includes membranes, filters, chips, slides, wafers, fibers, magnetic or nonmagnetic beads, gels, capillaries or other tubing, plates, polymers, and microparticles with a variety of surface forms including wells, trenches, pins, channels and pores.

"Modulates" refers to a change in activity (biological, chemical, or immunological) or lifespan resulting from specific binding between a molecule and either a nucleic acid molecule or a protein.

"Microarray" refers to an ordered arrangement of hybridizable array elements on a substrate. The array elements are arranged so that there are preferably at least ten or more different array elements, more preferably at least 100 array elements, even more preferably at least 1000 array elements, and most preferably 10,000. Furthermore, the hybridization signal from each of the array elements is individually distinguishable. In a preferred embodiment, the array elements comprise nucleic acid molecules.

"Nucleic acid molecule" refers to a nucleic acid, oligonucleotide, nucleotide, polynucleotide or any fragment thereof. It may be DNA or RNA of genomic or synthetic origin, double-stranded or single-stranded, and combined with carbohydrate, lipids, protein, or other materials to perform a particular activity such as transformation or form a useful composition such as a peptide nucleic acid (PNA). "Oligonucleotide" is substantially equivalent to the terms amplimer, primer, oligomer, element, target, and probe and is preferably single stranded.

"Protein" refers to an amino acid sequence, oligopeptide, peptide, polypeptide, or portions thereof whether naturally occurring or synthetic. Exemplary portions are the first twenty consecutive amino acids of a mammalian protein encoded by SEQ ID NOs:1-11, 17-33, 36, 39, and 41 and mammalian protein of SEQ ID NOs:50 and 53.

"Up-regulated" refers to a nucleic acid molecule whose levels increased in a treated sample compared with the nucleic acid molecule in an untreated sample.

"Down-regulated" refers to nucleic acid molecule whose levels decreased in a treated sample compared with the nucleic acid molecule in an untreated sample.

"Toxic compound" or "toxic agent" is any compound, molecule, or agent that elicits a biochemical, metabolic, and physiological response in an individual or animal, such as i) DNA damage, ii) cell damage, iii) organ damage or cell death, or iv) clinical morbidity or mortality.

"Toxicological response" refers to a biochemical, metabolic, and physiological response in an individual or animal which has been exposed to a toxic compound or agent.

"Fragment" refers to an Incyte clone or any part of a molecule which retains a usable, functional characteristic. Useful fragments include oligonucleotides and polynucleotides which may be used in hybridization or amplification technologies or in regulation of replication, transcription or translation. Exemplary fragments are the first sixty consecutive nucleotides of SEQ ID NOs:1-47. Useful fragments also include polypeptides and protein molecules which have antigenic potential and which may be used with a suitable pharmaceutical carrier in a pharmaceutical composition. Exemplary fragments are the first twenty consecutive amino acids of a mammalian protein encoded by SEQ ID NOs:1-11, 17-33, 36, 39, and 41 and mammalian protein of SEQ ID NOs:50 and 53.

"Hybridization complex" refers to a complex between two nucleic acid molecules by virtue of the formation of hydrogen bonds between purines and pyrimidines.

"Ligand" refers to any compound, molecule, or agent which will bind specifically to a complementary site on a nucleic acid molecule or protein. Such ligands stabilize or modulate the activity of nucleic acid molecules or proteins of the invention and may be composed of at least one of the following: inorganic and organic substances including nucleic acids, proteins, carbohydrates, fats, and lipids.

"Percent identity" or "% identity" refers to the percentage of sequence similarity found in a comparison of two or more amino acid or nucleic acid sequences. Percent identity can be determined electronically, e.g., by using the MEGALIGN program (DNASTAR, Madison WI) which creates alignments between two or more sequences according to methods selected by the user, e.g., the clustal method. (See, e.g., Higgins, D.G. and P.M. Sharp (1988) *Gene* 73:237-244.) The clustal algorithm groups sequences into clusters by examining the distances between all pairs. The clusters are aligned pairwise and then in groups. The percentage similarity between two amino acid sequences, e.g., sequence A and sequence B, is calculated by dividing the length of sequence A, minus the number of gap residues in sequence A, minus the number of gap residues in sequence B, into the sum of the residue matches between sequence A and sequence B, times one hundred. Gaps of low or of no similarity between the two amino acid sequences are not included in determining percentage similarity. Percent identity between nucleic acid sequences can also be counted or calculated by other methods known in the art, e.g., the Jotun Hein method. (See, e.g., Hein, J. (1990) *Methods Enzymol.* 183:626-645.) Identity between sequences can also be determined by other methods known in the art, e.g., by varying hybridization conditions.

"Substantially purified" refers to nucleic acid molecules or proteins that are removed from their natural environment and are isolated or separated, and are at least about 60% free, preferably about 75% free, and most preferably about 90% free, from other components with which they are naturally associated.

The Invention

The present invention provides mammalian nucleic acid and protein molecules and method of using the nucleic acid molecules for screening test compounds and molecules for toxicological responses. Additionally the invention provides methods for characterizing the toxicological responses of a sample to a test compound or molecule. In particular, the present invention provides a composition comprising a plurality of nucleic acid molecules derived from human cDNA libraries, monkey cDNA libraries, mouse cDNA libraries, normal rat liver cDNA libraries, normalized rat liver cDNA libraries, prehybridized rat liver cDNA libraries, subtracted rat liver cDNA libraries, and rat kidney cDNA libraries. The nucleic acid molecules have been further selected for exhibiting upregulated or downregulated gene expression

in rat livers when the rats have been exposed to a known hepatotoxin, including a peroxisomal proliferator (PP), acetaminophen or one of its corresponding metabolites, a polycyclic aromatic hydrocarbon (PAH), carbon tetrachloride, hydrazine, α -naphthylisothiocyanate, 4-acetylaminofluorene, and their corresponding metabolites.

5 PPs include hypolipidemic drugs, such as clofibrate, fenofibrate, clofenic acid, nafenopin, gemfibrozil, ciprofibrate, bezafibrate, halofenate, simfibrate, benzofibrate, etofibrate, WY-14,643, and the like; n-alkylcarboxylic acids, such as trichloroacetic acid, valproic acid, hexanoic acid, and the like; n-alkylcarboxylic acid precursors, such as trichloroethylene, tetrachloroethylene, and the like; azole antifungal compounds, such as bifonazole, and the like; leukotriene D4 antagonists; herbicides;
10 pesticides; phthalate esters, such as di-[2-ethylhexyl] phthalate, mono-[2-ethylhexyl] phthalate, and the like; and natural chemicals, such as phenyl acetate, dehydroepiandrosterone (DHEA), oleic acid, methanol, and the like. In a preferred embodiment the toxin is clofibrate, or one of its corresponding metabolites. In another preferred embodiment the toxin is fenofibrate, or one of its corresponding metabolites.

15 PAHs include compounds such as benzo(a)pyrene, 3-methylcholanthrene, benz(a)anthracene, 7,12-dimethylbenz(a)anthracene, their corresponding metabolites, and the like. In a preferred embodiment the toxin is benzo(a)pyrene, or one of its corresponding metabolites.

SEQ ID NOs:1-16 were identified by their pattern of at least two-fold upregulation or downregulation following hybridization with sample nucleic acid molecules from rat liver tissue treated
20 with a known toxic compound. SEQ ID NOs:17-47 were identified by their homology to the sample nucleic acid molecules from rat liver tissue treated with a known toxic compound. These and other nucleic acid molecules can be immobilized on a substrate as hybridizable array elements in a microarray format. The microarray may be used to characterize gene expression patterns associated with novel compounds to elucidate any toxicological responses or to monitor the effects of treatments during clinical
25 trials or therapy where metabolic responses to toxic compounds may be expected.

When the nucleic acid molecules are employed as hybridizable array elements in a microarray, the array elements are organized in an ordered fashion so that each element is present at a specified location on the substrate. Because the array elements are at specified locations on the substrate, the hybridization patterns and intensities (which together create a unique expression profile) can be
30 interpreted in terms of expression levels of particular genes and can be correlated with a toxicological response associated with a test compound or molecule.

The invention also provides a substantially purified and isolated mammalian protein comprising the protein molecule of SEQ ID NOs:50 and 53 or portion thereof. The invention further provides isolated and purified proteins encoded by the nucleic acid molecules of SEQ ID NOs:1-11, 17-33, 36, 39,

and 41, or portion thereof.

Furthermore, the present invention provides methods for screening test compounds or therapeutics for potential toxicological responses and for screening a sample's toxicological response to a particular test compound or molecule. Briefly, these methods entail treating a sample with the test compound or molecule to elicit a change in gene expression patterns comprising the expression of a plurality of sample nucleic acid molecules. Nucleic acid molecules are selected by identifying those genes in rat liver or kidney that are upregulated-or-downregulated at least 2-fold, more preferably at least 2.5-fold, most preferably at least 3-fold, when treated with a known toxic compound or molecule. The nucleic acid molecules are arrayed on a substrate. Then, the arrayed nucleic acid molecules and sample nucleic acid molecules are combined under conditions effective to form hybridization complexes which may be detected by methods well known in the art. Detection of higher or lower levels of such hybridization complexes compared with hybridization complexes derived from untreated samples and samples treated with a compound that is known not to induce a toxicological response correlates with a toxicological response of a test compound or a toxicological response to a molecule.

Complementary DNA libraries

Molecules are identified that reflect all or most of the genes that are expressed in rat liver or kidney. Molecules may be identified by isolating clones derived from several types of rat cDNA libraries, including normal rat cDNA libraries, normalized rat cDNA libraries, prehybridized rat cDNA libraries, and subtracted cDNA libraries. Clone inserts derived from these clones may be partially sequenced to generate expressed sequence tags (ESTs). Molecules are also identified by comparing the clones from rat cDNA libraries with clones from human, monkey, and mouse cDNA libraries using computer software nucleic acid comparison programs such as BLAST (see, e.g., Altschul, S.F. (1993) J. Mol. Evol. 3:290-300; Altschul, *et al.* (1990) J. Mol. Biol. 215:403-410).

In one embodiment, two collections of ESTs are identified and sequenced. A first collection of ESTs (the originator molecules) are derived from rat liver and kidney and are derived from the cDNA libraries presented in the Examples. A second collection includes ESTs derived from other rat cDNA libraries available in the ZOOSEQ database (Incyte Pharmaceuticals, Inc. Palo Alto CA).

The two collections of ESTs are clustered electronically to form master clusters of ESTs. Master clusters are formed by identifying overlapping EST molecules and assembling these ESTs. A nucleic acid fragment assembly tool, such as the Phrap tool (Phil Green, University of Washington) and the GELVIEW fragment assembly system (GCG, Madison WI), can be used for this purpose. The minimum number of clones which constitute a cluster is two. In another embodiment, a collection of human genes known to be expressed in response to toxic agents are used to select representative ESTs from the 113 rat cDNA libraries. The master cluster process is repeated for these molecules.

After assembling the clustered consensus nucleic acid sequences, a representative 5' clone is nominated from each master cluster. The most 5' clone is preferred because it is most likely to contain the complete gene. The nomination process is described in greater detail in "Relational Database and System for Storing Information Relating to Biomolecular Sequences and Reagents", USSN 09/034,807, filed March 4, 1998, herein incorporated in its entirety by reference. The EST molecules are used as array elements on a microarray.

Selection of arrayed nucleic acid molecules

Samples are treated, preferably at subchronic doses, with one or more known toxic compounds over a defined time course. Preferably, the agents are peroxisomal proliferators (PPs), acetaminophen or one of its corresponding metabolites, polycyclic aromatic hydrocarbons (PAHs), carbon tetrachloride, hydrazine, α -naphthylisothiocyanate, 4-acetylaminofluorene, or their corresponding metabolites.

The gene expression patterns derived from such treated biological samples can be compared with the gene expression patterns derived from untreated biological samples to identify and select nucleic acid molecules whose expression is either upregulated or downregulated due to the response to the toxic compounds. These selected molecules may then be employed as array elements alone or in combination with other array element molecules. Such a microarray is particularly useful to detect and characterize gene expression patterns associated with known toxic compounds. Such gene expression patterns can then be used for comparison to identify other compounds which also elicit a toxicological response.

The arrayed nucleic acid molecules can be manipulated to optimize their performance in hybridization. To optimize hybridization, the arrayed nucleic acid molecules are examined using a computer algorithm to identify portions of genes without potential secondary structure. Such computer algorithms are well known in the art and are part of OLIGO 4.06 primer analysis software (National Biosciences, Plymouth MN) or LASERGENE software (DNASTAR, Madison WI). These programs can search within nucleic acid sequences to identify stem loop structures and tandem repeats and to analyze G + C content of the sequence (those molecules with a G + C content greater than 60% are excluded). Alternatively, the arrayed nucleic acid molecules can be optimized by trial and error. Experiments can be performed to determine whether sample nucleic acid molecules and complementary arrayed nucleic acid molecules hybridize optimally under experimental conditions.

The arrayed nucleic acid molecules can be any RNA-like or DNA-like material, such as mRNAs, cDNAs, genomic DNA, peptide nucleic acids, branched DNAs and the like. The arrayed nucleic acid molecules can be in sense or antisense orientations.

In one embodiment, the arrayed nucleic acid molecules are cDNAs. The size of the DNA sequence of interest may vary, and is preferably from 50 to 10,000 nucleotides, more preferably from 150 to 3,500 nucleotides. In a second embodiment, the nucleic acid molecules are vector DNAs. In this case

the size of the DNA sequence of interest, i.e., the insert sequence, may vary from about 50 to 10,000 nucleotides, more preferably from about 150 to 3,500 nucleotides.

The nucleic acid molecule sequences of the Sequence Listing have been prepared by current, state-of-the-art, automated methods and, as such, may contain occasional sequencing errors and unidentified nucleotides. Nucleotide analogues can be incorporated into the nucleic acid molecules by methods well known in the art. The only requirement is that the incorporated nucleotide analogues must serve to base pair with sample nucleic acid molecules. For example, certain guanine nucleotides can be substituted with hypoxanthine which base pairs with cytosine residues. However, these base pairs are less stable than those between guanine and cytosine. Alternatively, adenine nucleotides can be substituted with 2,6-diaminopurine which can form stronger base pairs than those between adenine and thymidine. Additionally, the nucleic acid molecules can include nucleotides that have been derivatized chemically or enzymatically. Typical modifications include derivatization with acyl, alkyl, aryl or amino groups.

The nucleic acid molecules can be immobilized on a substrate via chemical bonding. Furthermore, the molecules do not have to be directly bound to the substrate, but rather can be bound to the substrate through a linker group. The linker groups are typically about 6 to 50 atoms long to provide exposure to the bound nucleic acid molecule. Preferred linker groups include ethylene glycol oligomers, diamines, diacids and the like. Reactive groups on the substrate surface react with one of the terminal portions of the linker to bind the linker to the substrate. The other terminal portion of the linker is then functionalized for binding the nucleic acid molecule. Preferred substrates are any suitable rigid or semirigid support, including membranes, filters, chips, slides, wafers, fibers, magnetic or nonmagnetic beads, gels, tubing, plates, polymers, microparticles and capillaries. The substrate can have a variety of surface forms, such as wells, trenches, pins, channels and pores, to which the arrayed nucleic acid molecules are bound.

The samples can be any sample comprising sample nucleic acid molecules and obtained from any bodily fluid (blood, urine, saliva, phlegm, gastric juices, etc.), cultured cells, biopsies, or other tissue preparations. The samples can be derived from any species, but preferably from eukaryotic species, and more preferably from mammalian species such as rat and human.

DNA or RNA can be isolated from the sample according to any of a number of methods well known to those of skill in the art. For example, methods of purification of nucleic acids are described in Tijssen, P. (1993) Laboratory Techniques in Biochemistry and Molecular Biology: Hybridization With Nucleic Acid Probes, Part I. Theory and Nucleic Acid Preparation, Elsevier, New York, NY. In one preferred embodiment, total RNA is isolated using the TRIZOL total RNA isolation reagent (Life Technologies, Inc., Gaithersburg MD) and mRNA is isolated using oligo d(T) column chromatography or

glass beads. When sample nucleic acid molecules are amplified it is desirable to amplify the sample nucleic acid molecules and maintain the relative abundances of the original sample, including low abundance transcripts. RNA can be amplified in vitro, in situ, or in vivo (See Eberwine US Patent No. 5,514,545).

5 It is also advantageous to include controls within the sample to assure that amplification and labeling procedures do not change the true distribution of nucleic acid molecules in a sample. For this purpose, a sample is spiked with an amount of a control nucleic acid molecule predetermined to be detectable upon hybridization to its complementary arrayed nucleic acid molecule and the composition of nucleic acid molecules includes reference nucleic acid molecules which specifically hybridize with the control arrayed nucleic acid molecules. After hybridization and processing, the hybridization signals
10 obtained should reflect accurately the amounts of control arrayed nucleic acid molecules added to the sample.

Prior to hybridization, it may be desirable to fragment the sample nucleic acid molecules. Fragmentation improves hybridization by minimizing secondary structure and cross-hybridization to other sample nucleic acid molecules in the sample or noncomplementary nucleic acid molecules.
15 Fragmentation can be performed by mechanical or chemical means.

Labeling

The sample nucleic acid molecules may be labeled with one or more labeling moieties to allow for detection of hybridized arrayed/sample nucleic acid molecule complexes. The labeling moieties can include compositions that can be detected by spectroscopic, photochemical, biochemical, bioelectronic,
20 immunochemical, electrical, optical or chemical means. The labeling moieties include radioisotopes, such as ³²P, ³³P or ³⁵S, chemiluminescent compounds, labeled binding proteins, heavy metal atoms, spectroscopic markers, such as fluorescent markers and dyes, magnetic labels, linked enzymes, mass spectrometry tags, spin labels, electron transfer donors and acceptors, and the like. Preferred fluorescent markers include Cy3 and Cy5 fluorophores (Amersham Pharmacia Biotech, Piscataway NJ).
25

Hybridization

The nucleic acid molecule sequence of SEQ ID NOs:1-47 and fragments thereof can be used in various hybridization technologies for various purposes. Hybridization probes may be designed or derived from SEQ ID NOs:1-47. Such probes may be made from a highly specific region such as the 5' regulatory region or from a conserved motif, and used in protocols to identify naturally occurring
30 sequences encoding the mammalian protein, allelic variants, or related sequences, and should preferably have at least 50% sequence identity to any of the protein sequences. The hybridization probes of the subject invention may be DNA or RNA and may be derived from the sequence of SEQ ID NOs:1-47 or from genomic sequences including promoters, enhancers, and introns of the mammalian gene.

Hybridization or PCR probes may be produced using oligolabeling, nick translation, end-labeling, or PCR amplification in the presence of the labeled nucleotide. A vector containing the nucleic acid sequence may be used to produce an mRNA probe in vitro by addition of an RNA polymerase and labeled nucleic acid molecules. These procedures may be conducted using commercially available kits such as those
5 provided by Amersham Pharmacia Biotech.

The stringency of hybridization is determined by G+C content of the probe, salt concentration, and temperature. In particular, stringency can be increased by reducing the concentration of salt or raising the hybridization temperature. In solutions used for some membrane based hybridizations, additions of an organic solvent such as formamide allows the reaction to occur at a lower temperature.

10 Hybridization can be performed at low stringency with buffers, such as 5 x SSC with 1% sodium dodecyl sulfate (SDS) at 60°C, which permits the formation of a hybridization complex between nucleotide sequences that contain some mismatches. Subsequent washes are performed at higher stringency with buffers such as 0.2 x SSC with 0.1% SDS at either 45°C (medium stringency) or 68°C (high stringency). At high stringency, hybridization complexes will remain stable only where the nucleic acid sequences are
15 completely complementary. In some membrane-based hybridizations, preferably 35% or most preferably 50%, formamide can be added to the hybridization solution to reduce the temperature at which hybridization is performed, and background signals can be reduced by the use of other detergents such as Sarkosyl or Triton X-100 and a blocking agent such as salmon sperm DNA. Selection of components and conditions for hybridization are well known to those skilled in the art and are reviewed in Ausubel
20 (supra) and Sambrook et al. (1989) Molecular Cloning, A Laboratory Manual, Cold Spring Harbor Press, Plainview NY.

Hybridization specificity can be evaluated by comparing the hybridization of specificity-control nucleic acid molecules to specificity-control sample nucleic acid molecules that are added to a sample in a known amount. The specificity-control arrayed nucleic acid molecules may have one or more sequence
25 mismatches compared with the corresponding arrayed nucleic acid molecules. In this manner, whether only complementary arrayed nucleic acid molecules are hybridizing to the sample nucleic acid molecules or whether mismatched hybrid duplexes are forming is determined.

Hybridization reactions can be performed in absolute or differential hybridization formats. In the absolute hybridization format, nucleic acid molecules from one sample are hybridized to the molecules in
30 a microarray format and signals detected after hybridization complex formation correlate to nucleic acid molecule levels in a sample. In the differential hybridization format, the differential expression of a set of genes in two biological samples is analyzed. For differential hybridization, nucleic acid molecules from both biological samples are prepared and labeled with different labeling moieties. A mixture of the two labeled nucleic acid molecules is added to a microarray. The microarray is then examined under

conditions in which the emissions from the two different labels are individually detectable. Molecules in the microarray that are hybridized to substantially equal numbers of nucleic acid molecules derived from both biological samples give a distinct combined fluorescence (Shalon et al. PCT publication WO95/35505). In a preferred embodiment, the labels are fluorescent markers with distinguishable
5 emission spectra, such as Cy3 and Cy5 fluorophores.

After hybridization, the microarray is washed to remove nonhybridized nucleic acid molecules and complex formation between the hybridizable array elements and the nucleic acid molecules is detected. Methods for detecting complex formation are well known to those skilled in the art. In a preferred embodiment, the nucleic acid molecules are labeled with a fluorescent label and measurement
10 of levels and patterns of fluorescence indicative of complex formation is accomplished by fluorescence microscopy, preferably confocal fluorescence microscopy.

In a differential hybridization experiment, nucleic acid molecules from two or more different biological samples are labeled with two or more different fluorescent labels with different emission wavelengths. Fluorescent signals are detected separately with different photomultipliers set to detect
15 specific wavelengths. The relative abundances/expression levels of the nucleic acid molecules in two or more samples is obtained.

Typically, microarray fluorescence intensities can be normalized to take into account variations in hybridization intensities when more than one microarray is used under similar test conditions. In a preferred embodiment, individual arrayed-sample nucleic acid molecule complex hybridization intensities
20 are normalized using the intensities derived from internal normalization controls contained on each microarray.

The labeled sample emits specific wavelengths which are detected using a plurality of photomultipliers. The nucleic acid molecules whose relative abundance/expression levels are modulated by treatment of a sample with a known toxic compound can be used as hybridizable elements in a
25 microarray. Such a microarray can be employed to identify expression profiles associated with particular toxicological responses. Then, a particular subset of these photomultipliers set to detect specific wavelengths. The relative expression levels of the arrayed nucleic acid molecules can be identified as to which arrayed nucleic acid molecule expression is modulated in response to a particular toxicological agent. These photomultipliers are set to detect specific wavelengths. The relative expression levels of
30 the nucleic acid molecules can be employed to identify other compounds with a similar toxicological response.

Alternatively, for some treatments with known side effects, the microarray, and expression patterns derived therefrom, is employed to prospectively define the treatment regimen. A dosage is established that minimizes expression patterns associated with undesirable side effects. This approach

may be more sensitive and rapid than waiting for the patient to show toxicological side effects before altering the course of treatment.

Generally, the method for screening a library of test compounds or molecules to identify those with a toxicological response entails selecting a plurality of arrayed genes whose expression levels are modulated in tissues treated with known toxic compounds when compared with untreated tissues. Then a sample is treated with the test compound or molecule to induce a pattern of gene expression comprising the expression of a plurality of sample nucleic acid molecules. Tissues from a mammalian subject treated at various dosages of the test compound may be screened to determine which doses may be toxic.

Then, the expression levels of the arrayed genes and the sample nucleic acid molecules are compared to identify those compounds that induce expression levels of the sample nucleic acid molecules that are similar to those of the arrayed genes. In one preferred embodiment, gene expression levels are compared by contacting the arrayed genes with the sample nucleic acid molecules under conditions effective to form hybridization complexes between arrayed genes and sample nucleic acid molecules; and detecting the presence or absence of the hybridization complexes.

Similarity may mean that at least 1, preferably at least 5, more preferably at least 10, of the upregulated arrayed genes form hybridization complexes with the sample nucleic acid molecules at least once during a time course to a greater extent than would the nucleic acid molecules of a sample not treated with the test compound. Similarity may also mean that at least 1, preferably at least 5, more preferably at least 10, of the downregulated nucleic acid molecules form hybridization complexes with the arrayed genes at least once during a time course to a lesser extent than would the nucleic acid molecules of a sample not treated with the test compound.

Such a similarity of expression patterns means that a toxicological response is associated with the compound or therapeutic tested. Preferably, the toxic compounds belong to the class of peroxisomal proliferators (PPs), including hypolipidemic drugs, such as clofibrate, fenofibrate, clofenic acid, nafenopin, gemfibrozil, ciprofibrate, bezafibrate, halofenate, simfibrate, benzofibrate, etofibrate, WY-14,643, and the like; n-alkylcarboxylic acids, such as trichloroacetic acid, valproic acid, hexanoic acid, and the like; n-alkylcarboxylic acid precursors, such as trichloroethylene, etrachloroethylene, and the like; azole antifungal compounds, such as bifonazole, and the like; leukotriene D4 antagonists; herbicides; pesticides; phthalate esters, such as di-[2-ethylhexyl] phthalate, mono-[2-ethylhexyl] phthalate, and the like; and natural chemicals, such as phenyl acetate, dehydroepiandrosterone (DHEA), oleic acid, methanol, and the like. In another embodiment, the toxic compound is acetaminophen or one of its corresponding metabolites. In yet another embodiment, the toxic compounds are polycyclic aromatic hydrocarbons (PAHs), including compounds such as benzo(a)pyrene, 3-methylcholanthrene, benz(a)anthracene, 7,12-dimethylbenz(a)anthracene, their corresponding metabolites, and the like. Of

particular interest is the study of the toxicological responses of these compounds on the liver, kidney, brain, spleen, pancreas, and lung.

Modification of Gene Expression Using Nucleic Acids

Gene expression may be modified by designing complementary or antisense molecules (DNA, RNA, or PNA) to the control, 5', 3', or other regulatory regions of the mammalian gene. Oligonucleotides designed with reference to the transcription initiation site are preferred. Similarly, inhibition can be achieved using triple helix base-pairing which inhibits the binding of polymerases, transcription factors, or regulatory molecules (Gee *et al.* In: Huber and Carr (1994) Molecular and Immunologic Approaches, Futura Publishing, Mt. Kisco NY, pp. 163-177). A complementary molecule may also be designed to block translation by preventing binding between ribosomes and mRNA. In one alternative, a library of nucleic acid molecules or fragments thereof may be screened to identify those which specifically bind a regulatory, nontranslated sequence.

Ribozymes, enzymatic RNA molecules, may also be used to catalyze the specific cleavage of RNA. The mechanism of ribozyme action involves sequence-specific hybridization of the ribozyme molecule to complementary target RNA followed by endonucleolytic cleavage at sites such as GUA, GUU, and GUC. Once such sites are identified, an oligonucleotide with the same sequence may be evaluated for secondary structural features which would render the oligonucleotide inoperable. The suitability of candidate targets may also be evaluated by testing their hybridization with complementary oligonucleotides using ribonuclease protection assays.

Complementary nucleic acids and ribozymes of the invention may be prepared via recombinant expression, *in vitro* or *in vivo*, or using solid phase phosphoramidite chemical synthesis. In addition, RNA molecules may be modified to increase intracellular stability and half-life by addition of flanking sequences at the 5' and/or 3' ends of the molecule or by the use of phosphorothioate or 2' O-methyl rather than phosphodiesterase linkages within the backbone of the molecule. Modification is inherent in the production of PNAs and can be extended to other nucleic acid molecules. Either the inclusion of nontraditional bases such as inosine, queosine, and wybutosine, and or the modification of adenine, cytidine, guanine, thymine, and uridine with acetyl-, methyl-, thio- groups renders the molecule less available to endogenous endonucleases.

Screening Assays

The nucleic acid molecule encoding the mammalian protein may be used to screen a library of molecules for specific binding affinity. The libraries may be DNA molecules, RNA molecules, PNAs, peptides, proteins such as transcription factors, enhancers, repressors, and other ligands which regulate the activity, replication, transcription, or translation of the nucleic acid molecule in the biological system. The assay involves combining the mammalian nucleic acid molecule or a fragment thereof with the

library of molecules under conditions allowing specific binding, and detecting specific binding to identify at least one molecule which specifically binds the nucleic acid molecule.

Similarly the mammalian protein or a portion thereof may be used to screen libraries of molecules in any of a variety of screening assays. The portion of the protein employed in such screening may be free in solution, affixed to an abiotic or biotic substrate (e.g. borne on a cell surface), or located intracellularly. Specific binding between the protein and molecule may be measured. Depending on the kind of library being screened, the assay may be used to identify DNA, RNA, or PNA molecules, agonists, antagonists, antibodies, immunoglobulins, inhibitors, peptides, proteins, drugs, or any other ligand, which specifically binds the protein. One method for high throughput screening using very small assay volumes and very small amounts of test compound is described in USPN 5,876,946, incorporated herein by reference, which screens large numbers of molecules for enzyme inhibition or receptor binding.

Purification of Ligand

The nucleic acid molecule or a fragment thereof may be used to purify a ligand from a sample. A method for using a mammalian nucleic acid molecule or a fragment thereof to purify a ligand would involve combining the nucleic acid molecule or a fragment thereof with a sample under conditions to allow specific binding, detecting specific binding, recovering the bound protein, and using an appropriate agent to separate the nucleic acid molecule from the purified ligand.

Similarly, the protein or a portion thereof may be used to purify a ligand from a sample. A method for using a mammalian protein or a portion thereof to purify a ligand would involve combining the protein or a portion thereof with a sample under conditions to allow specific binding, detecting specific binding between the protein and ligand, recovering the bound ligand, and using an appropriate chaotropic agent to separate the protein from the purified ligand.

Pharmacology

Pharmaceutical compositions are those substances wherein the active ingredients are contained in an effective amount to achieve a desired and intended purpose. The determination of an effective dose is well within the capability of those skilled in the art. For any compound, the therapeutically effective dose may be estimated initially either in cell culture assays or in animal models. The animal model is also used to achieve a desirable concentration range and route of administration. Such information may then be used to determine useful doses and routes for administration in humans.

A therapeutically effective dose refers to that amount of protein or inhibitor which ameliorates the symptoms or condition. Therapeutic efficacy and toxicity of such agents may be determined by standard pharmaceutical procedures in cell cultures or experimental animals, e.g., ED₅₀ (the dose therapeutically effective in 50% of the population) and LD₅₀ (the dose lethal to 50% of the population). The dose ratio between toxic and therapeutic effects is the therapeutic index, and it may be expressed as

the ratio, LD_{50}/ED_{50} . Pharmaceutical compositions which exhibit large therapeutic indexes are preferred. The data obtained from cell culture assays and animal studies are used in formulating a range of dosage for human use.

5 MODEL SYSTEMS

Animal models may be used as bioassays where they exhibit a toxic response similar to that of humans and where exposure conditions are relevant to human exposures. Mammals are the most common models, and most toxicity studies are performed on rodents such as rats or mice because of low cost, availability, and abundant reference toxicology. Inbred or outbred rodent strains provide a
10 convenient model for investigation of the physiological consequences of under- or over-expression of genes of interest and for the development of methods for diagnosis and treatment of diseases. A mammal inbred to over-express a particular gene, so that the protein is secreted in milk, may also serve as a convenient source of the protein expressed by that gene.

Toxicology

15 Toxicology is the study of the effects of test compounds, molecules, or toxic agents on living systems to identify adverse effects. The majority of toxicity studies are performed on rats or mice to help predict whether adverse effects of agents will occur in humans. Observation of qualitative and quantitative changes in physiology, behavior, homeostatic, developmental, and reproductive processes, and lethality are used to generate profiles of safe or toxic responses and to assess the consequences on
20 human health following exposure to the agent.

Toxicological tests measure the effects of a single, repeated, or long-term exposure of a subject to a substance. Substances may be tested for specific endpoints such as cytotoxicity, mutagenicity, carcinogenicity and teratogenicity. Degree of response varies according to the route of exposure (contact, ingestion, injection, or inhalation), age, sex, genetic makeup, and health status of the subject. Other tests
25 establish the toxicokinetic and toxicodynamic properties of substances. Toxicokinetic studies trace the absorption, distribution in subject tissues, metabolism, storage, and excretion of substances. Toxicodynamic studies chart biological responses that are consequences of the presence of the substance in the subject tissues.

Genetic toxicology identifies and analyzes the ability of an agent to produce damage at a cellular
30 or subcellular level. Such genotoxic agents usually have common chemical or physical properties that facilitate interaction with nucleic acids and are most harmful when mutated chromosomes are passed along to progeny. Toxicological studies may identify agents that increase the frequency of structural or functional abnormalities in progeny if administered to either parent before conception, to the mother during pregnancy, or to the developing organism. Mice and rats are most frequently used in these tests

because of their short reproductive cycle which allows investigators to breed sufficient quantities of individual animals to satisfy statistical requirements.

All types of toxicology studies on experimental animals involve preparation of a suitable form of the compound for administration, selection of the route of administration, and selection of a species which resembles the species of pharmacological interest. Dose concentrations of the compound are varied to identify, measure, and investigate a range of dose-related effects related to exposure.

Acute toxicity tests are based on a single administration of the agent to the subject to determine the symptomology or lethality of the agent. Three experiments are conducted; an experiment to define the initial dose range; an experiment to narrow the range of effective doses; and a final experiment to establish the dose-response curve.

Prolonged and subchronic toxicity tests are based on the repeated administration of the agent. Rat and dog are commonly used in these studies to provide data from species in different taxonomic orders. With the exception of carcinogenesis, there is considerable evidence that daily administration of an agent at high-dose concentrations for periods of three to four months will reveal most forms of toxicity in adult animals.

Chronic toxicity tests, with a duration of a year or more, are used to demonstrate either the absence of toxicity or the carcinogenic potential of an agent. When studies are conducted on rats, a minimum of at least one test group plus one control group are used. Animals are quarantined, examined for health, and monitored at the outset and at intervals throughout the experiment.

Transgenic Animal Models

Transgenic rodents which over-express or under-express a gene of interest may be inbred and used to model human diseases or to test therapeutic or toxic agents. (See USPN 4,736,866; USPN 5,175,383; and USPN 5,767,337; incorporated herein by reference). In some cases, the introduced gene may be activated at a specific time in a specific tissue type during fetal development or postnatally.

Expression of the transgene is monitored by analysis of phenotype or tissue-specific mRNA expression, in transgenic animals before, during, and after being challenged with experimental drug therapies.

Embryonic Stem Cells

Embryonic stem cells (ES) isolated from rodent embryos retain the potential to form an embryo. When ES cells are placed inside a carrier embryo, they resume normal development and contribute to all tissues of the live-born animal. ES cells are the preferred cells used in the creation of experimental knockout and knockin rodent strains. Mouse ES cells, such as the mouse 129/SvJ cell line, are derived from the early mouse embryo and are grown under culture conditions well known in the art. Vectors for knockout strains contain a disease gene candidate modified to include a marker gene which disrupts transcription and/or translation of the endogenous disease candidate gene *in vivo*. The vector is

introduced into ES cells by transformation methods such as electroporation, liposome delivery, microinjection, and the like which are well known in the art. The endogenous rodent gene is replaced by the disrupted disease gene through homologous recombination and integration during cell division. Expression of the marker gene confers a selective advantage to the transformed cells when incubated with an otherwise toxic/lethal selecting agent. Transformed ES cells are selected, identified, and preferably microinjected into mouse cell blastocysts such as those from the C57BL/6 mouse strain. The blastocysts are surgically transferred to pseudopregnant dams and the resulting chimeric progeny are genotyped and bred to produce heterozygous or homozygous strains.

ES cells are also used to study the differentiation of various cell types and tissues in vitro, such as neural cells, hematopoietic lineages, and cardiomyocytes (Bain et al. (1995) Dev. Biol. 168:342-357; Wiles and Keller (1991) Development 111:259-267; and Klug et al. (1996) J. Clin. Invest. 98:216-224). Recent developments demonstrate that ES cells derived from human blastocysts may also be manipulated in vitro to differentiate into eight separate cell lineages, including endoderm, mesoderm, and ectodermal cell types (Thomson et al. (1998) Science 282:1145-1147).

Knockout Analysis

In gene knockout analysis, a region of a human disease gene candidate is enzymatically modified to include a non-mammalian gene such as the neomycin phosphotransferase gene (*neo*; Capecchi (1989) Science 244:1288-1292). The inserted coding sequence disrupts transcription and translation of the targeted gene and prevents biochemical synthesis of the disease candidate protein. The modified gene is transformed into cultured embryonic stem cells (described above), the transformed cells are injected into rodent blastulae, and the blastulae are implanted into pseudopregnant dams. Transgenic progeny are crossbred to obtain homozygous inbred lines.

Knockin Analysis

Totipotent ES cells, present in the early stages of embryonic development, can be used to create knockin humanized animals (pigs) or transgenic animal models (mice or rats) of human diseases. With knockin technology, a region of a human gene is injected into animal ES cells, and the human sequence integrates into the animal cell genome by recombination. Totipotent ES cells which contain the integrated human gene are handled as described above. Inbred animals are studied and treated to obtain information on the analogous human condition. These methods have been used to model several human diseases. (See, e.g., Lee et al. (1998) Proc. Natl. Acad. Sci. 95:11371-11376; Baudoin et al. (1998) Genes Dev. 12:1202-1216; and Zhuang et al. (1998) Mol. Cell Biol. 18:3340-3349).

Non-Human Primate Model

The field of animal testing deals with data and methodology from basic sciences such as physiology, genetics, chemistry, pharmacology and statistics. These data are paramount in evaluating the

effects of therapeutic agents on non-human primates as they can be related to human health. Monkeys are used as human surrogates in vaccine and drug evaluations, and their responses are relevant to human exposures under similar conditions. Cynomolgus and Rhesus monkeys (Macaca fascicularis and Macaca mulatta, respectively) and Common Marmosets (Callithrix jacchus) are the most common non-human primates (NHPs) used in these investigations. Since great cost is associated with developing and maintaining a colony of NHPs, early research and toxicological studies are usually carried out in rodent models. In studies using behavioral measures such as drug addiction, NHPs are the first choice test animal. In addition, NHPs and individual humans exhibit differential sensitivities to many drugs and toxins and can be classified as a range of phenotypes from "extensive metabolizers" to "poor metabolizers" of these agents.

In additional embodiments, the nucleic acid molecules which encode the mammalian protein may be used in any molecular biology techniques that have yet to be developed, provided the new techniques rely on properties of nucleic acid molecules that are currently known, including, but not limited to, such properties as the triplet genetic code and specific base pair interactions.

Examples

It is understood that this invention is not limited to the particular methodology, protocols, and reagents described, as these may vary. It is also understood that the terminology used herein is for the purpose of describing particular embodiments only, and is not intended to limit the scope of the present invention which will be limited only by the appended claims. The examples below are provided to best describe the subject invention and its representative constituents.

I cDNA Library Construction

The RALINOT01 cDNA library was constructed from liver tissue removed from a pool of fifty 10- to 11-week-old Sprague-Dawley female rats (Pharmakon, Waverly PA). The animals were housed in standard laboratory caging and fed PMI-certified Rodent Diet #5002. The animals appeared to be in good health at the time tissue was harvested. The animals were anesthetized by CO₂ inhalation, and then cardiocentesis was performed.

Frozen tissue was homogenized and lysed in TRIZOL reagent (1 g tissue/10 ml TRIZOL; Life Technologies), a monophasic solution of phenol and guanidine isothiocyanate, using a POLYTRON homogenizer (PT-3000; Brinkmann Instruments, Westbury NY). After a brief incubation on ice, chloroform (1:5 v/v) was mixed with the reagent, and then centrifuged at 1,000 rpm. The upper aqueous layer was removed to a fresh tube, and the RNA precipitated with isopropanol, resuspended in DEPC-treated water, and treated with DNase I for 25 min at 37°C. The RNA was re-extracted once with

phenol-chloroform, pH 4.7, and precipitated using 0.3 M sodium acetate and 2.5 volumes ethanol. The mRNA was then isolated using an OLIGOTEX kit (QIAGEN, Chatsworth CA) and used to construct the cDNA library.

The mRNA was handled according to the recommended protocols in the SUPERScript plasmid system (Life Technologies). The cDNAs were fractionated on a SEPHAROSE CL-4B column (Amersham Pharmacia Biotech), and those cDNAs exceeding 400 bp were ligated into the pINCY1 plasmid vector (Incyte Pharmaceuticals). The plasmid pINCY1 was subsequently transformed into DH5 α or DH10B competent cells (Life Technologies).

The RAKINOT01 library was constructed using mRNA isolated from kidney tissue removed from a pool of fifty, 7- to 8-week-old male Sprague-Dawley rats, as described above.

The RAKINOT02 library was constructed using mRNA isolated from kidney tissue removed from a pool of fifty, 10- to 11-week-old female Sprague-Dawley rats, as described above.

II cDNA Library Normalization

In some cases, cDNA libraries were normalized in a single round according to the procedure of Soares *et al.* (1994, Proc. Natl. Acad. Sci. 91:9228-9232) with the following modifications. The primer to template ratio in the primer extension reaction was increased from 2:1 to 10:1. Reduction of each dNTP concentration in the reaction to 150 μ M allowed the generation of longer (400-1000 nucleotide (nt)) primer extension products. The reannealing hybridization was extended from 13 to 19 hours. The single stranded DNA circles of the normalized library were purified by hydroxyapatite chromatography, converted to partially double-stranded by random priming, and electroporated into DH10B competent bacteria (Life Technologies).

The Soares normalization procedure is designed to reduce the initial variation in individual cDNA frequencies and to achieve abundances within one order of magnitude while maintaining the overall sequence complexity of the library. In the normalization process, the prevalence of high-abundance cDNA clones decreases significantly, clones with mid-level abundance are relatively unaffected, and clones for rare transcripts are increased in abundance. In the modified Soares normalization procedure, significantly longer hybridization times are used to increase gene discovery rates by biasing the normalized libraries toward low-abundance cDNAs that are well represented in a standard transcript image.

The RALINON03, RALINON04, and RALINON07 normalized rat liver cDNA libraries were constructed with 2.0×10^6 , 4.6×10^5 , and 2.0×10^6 independent clones from the RALINOT01 cDNA library, respectively. The RALINOT01 cDNA library was normalized in one round using conditions adapted from Soares (*supra*) except that a significantly longer (48-hour) reannealing hybridization was

used.

III cDNA Library Prehybridization

The RALINOH01 cDNA library was constructed with clones from the RALINOT01 cDNA library. After preparation of the RALINOT01 cDNA library, 9,984 clones were spotted onto a nylon filter, lysed, and the plasmid DNA was bound to the filter. The filter was incubated with pre-warmed hybridization buffer and then hybridized at 42°C for 14-16 hours in 0.75 M NaCl, 0.1 M Na₂HPO₄/NaH₂PO₄, 0.15 M tris-HCl (pH 7.5), 5x Denhardt's Solution, 2% SDS, 100 µg/ml sheared salmon sperm DNA, 50% formamide, and [³²P]-labeled oligonucleotide molecules made from reverse transcribed rat liver mRNA from an untreated animal. The filter was rinsed with 2 x SSC (saline sodium citrate) at ambient temperature for 5 minutes followed by washing for 30 minutes at 68°C with pre-warmed washing solution (2 x SSC, 1% SDS). The wash was repeated with fresh washing solution for an additional 30 minutes at 68°C. Filters were then washed twice with pre-warmed washing solution (0.6 x SSC, 1% SDS) for 30 minutes at 68°C. Some 4,224 clones had very low hybridization signals and about 20% of the clones had no signals and two groups were isolated and sequenced.

IV Isolation and Sequencing of cDNA Clones

DNA was isolated using the following protocol. Single bacterial colonies were transferred into individual wells of 384-well plates (Genetix Ltd, Christchurch, United Kingdom) using sterile toothpicks. The wells contained 1 ml of sterile Terrific Broth (Life Technologies) with 25 mg/l carbenicillin and 0.4% glycerol (v/v). The plates were covered and placed in an incubator (Thermodyne, Newtown Square PA) at 37°C for 8-10 hours. Plasmid DNA was released from the cells and amplified using direct link PCR (Rao, V.B. (1994) Anal. Biochem. 216:1-14) as follows. The direct link PCR solution included 30 µl of NUCLEIX PLUS PCR nucleotide mix (Amersham Pharmacia Biotech, Piscataway NJ) and 300 µl of Taq DNA polymerase (Amersham Pharmacia Biotech). Five microlitres of the PCR solution were added to each of the 384 wells using the MICROLAB 2200 system (Hamilton, Reno NV); plates were centrifuged at 1000 rpm for 20 seconds and refrigerated until use. A 384 pin tool (V&P Scientific Inc, San Diego CA) was used to transfer bacterial cells from the incubation plate into the plate containing the PCR solution where 0.1% Tween 20 caused the cells to undergo lysis and release the plasmid DNA. After lysis, the plates were centrifuged up to 500 rpm, covered with a cycle sealer, and cycled using a 384-well DNA ENGINE thermal cycler (MJ Research, Watertown MA) using the program dPCR30 with the following parameters: Step 1) 95 °C, 1 minute; Step 2) 94 °C, 30 seconds; Step 3) 55 °C, 30 seconds; Step 4) 72 °C, 2 minutes; Step 5) steps 2, 3, and 4 repeated 29 times; Step 6) 72 °C, 10 minutes; and Step 7) storage at 4 °C.

The concentration of DNA in each well was determined by dispensing 100 µl PICO GREEN quantitation reagent (0.25% (v/v), Molecular Probes, Eugene OR) dissolved in 1x TE and 0.5 µl of undiluted PCR product into each well of an opaque fluorimeter plate (Corning Costar, Acton MA), allowing the DNA to bind to the quantitation reagent. The plate was scanned in a Fluoroscan II
5 (Labsystems Oy, Helsinki, Finland) to measure the fluorescence of the sample and to quantitate the concentration of DNA. Typical concentrations of each DNA sample were in the range of 100 to 500 ng/ml.

The cDNAs were prepared for sequencing using either a HYDRA microdispenser (Robbins Scientific, Sunnyvale CA) or MICROLAB 2200 system (Hamilton) in combination with the DNA
10 ENGINE thermal cyclers (MJ Research). The cDNAs were sequenced using the method of Sanger, F. and A.R. Coulson (J. Mol. Biol. (1975) 94:441-448) and the ABI 377 sequencing systems (PE Biosystems). Most of the isolates were sequenced according to standard ABI protocols using ABI kits (PE Biosystems). The solution volumes were used at 0.25x - 1.0x concentrations. Typically, 500 to 700 base pairs were sequenced in 3.5 to 4 hours. In the alternative, cDNAs may have been sequenced using
15 solutions and dyes from Amersham Pharmacia Biotech.

V Rat Liver and Kidney Gene Selection

As a first step, originator molecules from high throughput sequencing experiments were derived from clone inserts from RALINOT01, RAKINOT01, RAKINOT02, RALINOH01, RALINON03,
20 RALINON04 and RALINON07. cDNA library clones were obtained. There were 18,140 rat liver molecules and 5,779 rat kidney molecules.

Additionally, 1,500 rat molecules derived from clone inserts of any of 113 rat cDNA libraries were selected based on their homology to genes coding for polypeptides implicated in toxicological responses including peroxisome-associated genes, lysosome-associated genes, apoptosis-associated
25 genes, cytochrome P450 genes, detoxification genes such as sulfotransferases, glutathione S-transferases, and cysteine proteases, and the like.

Then, all the remaining molecules derived from all of the rat cDNA library clones were clustered based on the originator molecules described above. The clustering process involved identifying overlapping molecules that have a match quality indicated by a product score of 50 using BLAST.
30 6581 master clusters were identified.

After forming the clone clusters, a consensus sequence was generated based on the assembly of the clone molecules using PHRAP (Phil Green, University of Washington). The assembled molecules were then annotated by first screening the assembled molecules against GenBank using BLASTn and then by screening the assembled molecules against GenPept using FASTX. About two thirds of the

assembled molecules were annotated, about one third of the assembled molecules were not annotated. For example, for nucleic acid sequence analysis, the program BLASTN 1.4.9MP-WashU was used with default parameters; ctxfactor=2.00; E=10; MatID, 0; Matrix name, +5,-4. In another example, for amino acid sequence analysis, the program NCBI-BLASTX 2.0.4 was used with default parameters; matrix, BLOSUM62; gap penalties, existence 11, extension 1; frameshift window, decay constant 50, 0.1.

VI Substrate and Array Element/Probe Preparation

Clones nominated in the process described in Example V were used to generate array elements. Each array element was amplified from bacterial cells. PCR amplification used primers complementary to the vector sequences flanking the cDNA insert. Array elements were amplified in thirty cycles of PCR from an initial quantity of 1-2 ng to a final quantity greater than 5 µg. Amplified array elements were then purified using SEPHACRYL-400 (Amersham Pharmacia Biotech).

Purified array elements were immobilized on polymer-coated glass slides. Glass microscope slides (Corning, Corning NY) cleaned by ultrasound in 0.1% SDS and acetone, with extensive distilled water washes between and after treatments. Glass slides were etched in 4% hydrofluoric acid (VWR, West Chester PA), washed extensively in distilled water, and coated with 0.05% aminopropyl silane (Sigma-Aldrich, St. Louis MO) in 95% ethanol. Coated slides were cured in a 110°C oven.

Array elements were applied to the coated glass substrate using a procedure described in US Patent No. 5,807,522 and incorporated herein by reference. In brief, 1 µl of the array element DNA, at an average concentration of 0.5 µg/ml in 3 x SSC, was loaded into the open capillary printing element by a high-speed robotic apparatus. The apparatus then deposited about 5 nl of the array element sample per slide. A total of 7404 array elements representing rat liver and kidney genes and a variety of control elements, including 14 synthetic control molecules, human genomic DNA, and yeast genomic DNA, were arrayed in four identical quadrants within a 1.8 cm² area of the glass substrate.

Microarrays were UV-crosslinked using a STRATALINKER UV-crosslinker (Stratagene). Microarrays were washed at room temperature once in 0.2% SDS and three times in distilled water. Non-specific binding sites were blocked by incubation of microarrays in 0.2% casein in phosphate buffered saline (PBS; Tropix Inc., Bedford MA) for 30 minutes at 60°C followed by washes in 0.2% SDS and distilled water as before.

VII Target Preparation

Male Sprague-Dawley rats (6-8 wk old) were dosed intraperitoneally with one of the following: clofibrate (CLO; Acros, Geel, Belgium) at 250 mg/kg body weight (bw); acetaminophen (APAP; Acros) at 1000 mg/kg bw; benzo(a)pyrene (B(a)P; Acros) at 10 mg/kg bw; or dimethylsulfoxide vehicle (DMSO;

Acros) at less than 2 ml/kg bw, and the animals were later euthanized by CO₂ inhalation. Animals were monitored daily for physical condition and body weight. Three animals per group were sacrificed approximately 12 hours, 24 hours, 3d (d), 7d, 14d, and 28d following the single dose. Prior to sacrifice a blood sample from each animal was drawn and assayed for serum alanine transferase (ALT) and serum aspartate aminotransferase (AST) levels using a diagnostic kit (Sigma-Aldrich). Observed gross pathology and liver weights were recorded at time of necropsy. Liver, kidney, brain, spleen and pancreas from each rat were harvested, flash frozen in liquid nitrogen, and stored at -80°C.

In the alternative, male Han-Wistar rats (8-9 wk old) were dosed by oral gavage with one of the following: fenofibrate (FEN; Sigma-Aldrich) at 250 mg/kg bw; carbon tetrachloride (CCL₄; Sigma-Aldrich) at 3160 mg/kg bw, hydrazine (HYDR; Sigma-Aldrich) at 120 mg/kg bw; α -naphthylisothiocyanate (ANIT; Sigma-Aldrich) at 200 mg/kg bw; 4-acetylaminofluorene (4-AFF; Lancaster Synthesis, Morecambe, Lancashire, UK) at 1000 mg/kg bw; corn oil vehicle, or sterile water vehicle, at 10 ml/kg bw. The animals were checked twice daily for clinical signs of distress. Blood was collected six days prior to the dose and at sacrifice. Three animals per group were sacrificed approximately six hours and 24 hours following the single dose. The animals were euthanized by exsanguination under isoflurane anaesthesia. Observed gross pathology and liver weights were recorded at time of necropsy. Livers from each rat were harvested, dissected into approximate 100 mg pieces, flash frozen in liquid nitrogen, and stored at -70°C.

For each target preparation, frozen liver was homogenized and lysed in TRIZOL reagent (Life Technologies, Gaithersburg MD) following the modifications for liver RNA isolation. Messenger RNA was isolated using an OLIGOTEX kit (QIAGEN) and labeled with either Cy3- or Cy5-labeled primers (Operon Technologies, Alameda CA) using the GEMBRIGHT labeling kit (Incyte Pharmaceuticals). Messenger RNA isolated from tissues of rats treated with clofibrate, acetaminophen, or benzo(a)pyrene was labeled with Cy5 and mRNA isolated from tissues of rats treated with DMSO was labeled with Cy3. Quantitative and differential expression pattern control cDNAs were added to each labeling reaction. Labeled cDNA was treated with 0.5 M sodium bicarbonate (pH 9.2) for 20 min at 85°C to degrade the RNA and purified using two successive CHROMA SPIN 30 gel filtration spin columns (Clontech, Palo Alto CA). Cy3-labeled control sample and Cy5-labeled experimental sample were combined and precipitated in glycogen, sodium acetate, and ethanol.

Targets are also prepared from tissue needle biopsy samples. Samples are used to identify changes within the tissue following exposure to, for example, a toxic compound, a potential toxic compound, a compound with unknown metabolic responses, and a pharmacological compound.

VIII Hybridization

Hybridizations were carried out using the methods described by Shalon (supra).

IX Detection

5 The microscope used to detect the reporter-labeled hybridization complexes was equipped with an Innova 70 mixed gas 10 W laser (Coherent Lasers, Santa Clara CA) capable of generating spectral lines at 488 nm for excitation of Cy3, and 632 nm for excitation of Cy5. The excitation laser light was focused on the array using a 20x microscope objective (Nikon, Melville NY). The slide containing the array was placed on a computer-controlled X-Y stage on the microscope and raster-scanned past the
10 objective. The 1.8 cm x 1.8 cm array used in the present example was scanned with a resolution of 20 micrometers.

 In two separate scans, a mixed gas multiline laser excited the two fluorophores sequentially. Emitted light was split, based on wavelength, into two photomultiplier tube detectors (PMT R1477, Hamamatsu Photonics, San Jose CA) corresponding to the two fluorophores. Appropriate filters
15 positioned between the array and the photomultiplier tubes were used to filter the signals. The emission maxima of the fluorophores used were 565 nm for Cy3 and 650 nm for Cy5. Each array was typically scanned twice, one scan per fluorophore using the appropriate filters at the laser source, although the apparatus was capable of recording the spectra from both fluorophores simultaneously.

 The sensitivity of the scans was typically calibrated using the signal intensity generated by a
20 cDNA control species added to the probe mix at a known concentration. A specific location on the array contained a complementary DNA sequence, allowing the intensity of the signal at that location to be correlated with a weight ratio of hybridizing species of 1:100,000. When two probes from different sources (e.g., representing test and control cells), each labeled with a different fluorophore, are hybridized to a single array for the purpose of identifying genes that are differentially expressed, the
25 calibration was done by labeling samples of the calibrating cDNA with the two fluorophores and adding identical amounts of each to the hybridization mixture.

 The output of the photomultiplier tube was digitized using a 12-bit RTI-835H analog-to-digital (A/D) conversion board (Analog Devices, Norwood MA) installed in an IBM-compatible PC computer. The digitized data were displayed as an image where the signal intensity was mapped using a linear 20-
30 color transformation to a pseudocolor scale ranging from blue (low signal) to red (high signal). The data was also analyzed quantitatively. Where two different fluorophores were excited and measured simultaneously, the data were first corrected for optical crosstalk (due to overlapping emission spectra) between the fluorophores using each fluorophore's emission spectrum.

 A grid was superimposed over the fluorescence signal image such that the signal from each spot

was centered in each element of the grid. The fluorescence signal within each element was then integrated to obtain a numerical value corresponding to the average intensity of the signal. The software used for signal analysis was the GEMTOOLS gene expression analysis program (Incyte Pharmaceuticals).

5

X Results

The expression patterns of eight cytochrome P450 isozymes known to be induced in a toxicological response were monitored during the 28 day time course. The results using clofibrate, acetaminophen, and benzo(a)pyrene are shown in Table 1, Table 2, and Table 3, respectively. Each of the known genes was upregulated or downregulated greater than 2-fold at least once during the time course.

TABLE 1 Gene expression patterns (x-fold change) of known genes in clofibrate-treated rat liver

15

Gene	12 hours	24 hours	3 days	7 days	28 days
P450 LA-omega 4A3	14.8	26.6	1.1	0.5	0.47
P450 4A	7.0	16.6	1.4	0.5	1.3
P450 3A2	0.14	1.2	0.63	0.50	0.45

TABLE 2 Gene expression patterns (x-fold change) of known genes in acetaminophen-treated rat liver

20

Gene	12 hours	24 hours	3 days	7 days	14 days	28 days
P450 4A	1.0	4.5	2.1	2.0	4.4	4.8
P450f 2C7	0.21	0.43	0.47	0.5	1.2	1.3
P450 14DM	0.31	0.20	2.0	1.1	1.4	0.42

TABLE 3 Gene expression patterns (x-fold change) of known genes in benzo(a)pyrene-treated rat liver

25

Gene	12 hours	24 hours	3 days	7 days	14 days	28 days
P450 LA-omega 4A3	1.2	2.3	2.4	1.4	6.8	1.2
P450 MCA-inducible 1A2	7.3	9.2	5.7	2.5	2.5	0.5

30

In addition, results from two samples that had been treated identically were compared to determine the range of normal variation of gene expression patterns between the samples. In one analysis, where two different samples were prepared from identically treated tissues, gene expression patterns of cDNAs which were upregulated or downregulated not more than 1.7-fold were within the 95% confidence limits of a Poisson normal distribution. In a separate analysis, gene expression patterns of cDNAs which were upregulated or downregulated more than 2-fold accounted for not more than 5% of

the total hybridizable sample nucleic acid molecules in two identically-treated tissue samples.

We have discovered novel nucleotide molecules that are up-regulated or down-regulated at least 2-fold at least once during the time course. These molecules are SEQ ID NOs:1-16 provided in the Sequence Listing. These polynucleotide molecules can be used for screening compounds or therapeutics for a toxicologic effect and applications including detecting metabolic and toxicological responses, and in monitoring drug mechanism of action.

Table 4 shows the gene expression pattern of selected molecules that were upregulated at least 2-fold at least once during the time course following treatment with clofibrate (CLO). Table 5 shows the gene expression pattern of selected molecules that were downregulated at least 2-fold at least once during the time course following treatment with CLO.

TABLE 4 Gene expression patterns (x-fold change) of CLO-upregulated nucleic acid molecules

SEQ ID NO:	12 hours	24 hours	3 days	7 days	28 days
2	2.6	1.4	0.5	1.1	1.2
3	1.3	2	1.3	1.5	1.5
4	2	0.36	0.47	0.26	0.30
5	1.7	2.9	1.6	1.5	1.2
8	2.6	1.7	1.3	1.3	1.4

TABLE 5 Gene expression patterns (x-fold change) of CLO-downregulated nucleic acid molecules

SEQ ID NO:	12 hours	24 hours	3 days	7 days	28 days
1	n.d.	0.26	0.45	0.26	1.1
4	2.0	0.36	0.47	0.26	0.30
7	0.24	0.42	0.37	1.1	1.5

(n.d. = not detected)

Table 6 shows the gene expression pattern of selected molecules that were upregulated at least 2-fold at least once during the time course following treatment with acetaminophen (APAP). Table 7 shows the gene expression pattern of selected molecules that were downregulated at least 2-fold at least once during the time course following treatment with APAP.

TABLE 6 Gene expression patterns (x-fold change) of APAP-upregulated nucleic acid molecules

SEQ ID NO:	12 hours	24 hours	3 days	7 days	14 days	28 days
2	1.3	2.2	1.1	0.5	1.2	1.3
3	1.2	2.1	0.47	0.46	1.8	1.5
4	3.3	0.47	0.47	0.23	0.35	0.36
5	1.1	2.1	1.1	1.2	1.3	1.4
6	1.8	5	2.5	1.1	1.4	1.3
8	1.1	2.5	1.1	1	1.7	1.4

TABLE 7 Gene expression patterns (x-fold change) of APAP-downregulated nucleic acid molecules

SEQ ID NO:	12 hours	24 hours	3 days	7 days	14 days	28 days
1	0.36	0.19	0.46	0.25	0.5	1.4
4	3.3	0.48	0.47	0.23	0.35	0.36
7	0.33	0.21	1.7	n.d.	1	0.39

(n.d. = not detected)

Table 8 shows the gene expression pattern of selected molecules that were upregulated at least 2-fold at least once during the time course following treatment with benzo(a)pyrene (B(a)P). Table 9 shows the gene expression pattern of selected molecules that were downregulated at least 2-fold at least once during the time course following treatment with B(a)P.

TABLE 8 Gene expression patterns (x-fold change) of B(a)P-upregulated nucleic acid molecules

SEQ ID NO:	12 hours	1 day	3 days	7 days	14 days	28 days
2	0.5	0.47	1.2	1.1	2.6	0.47
3	1.4	2.1	1.2	1.5	2.7	1.6
5	1.5	1.4	1.2	0.47	2	0.46
6	2.2	1.4	1.4	1.2	2.2	n.d.
7	1.2	2.2	1.4	0.5	0.42	1.1
8	1.6	1.7	1.3	1.3	2	1.1

(n.d. = not detected)

TABLE 9 Gene expression patterns (x-fold change) of B(a)P-downregulated nucleic acid molecules

SEQ ID NO:	12 hours	1 day	3 days	7 days	14 days	28
1	0.37	0.39	0.35	1.4	0.33	1.5
4	0.5	0.26	0.31	0.36	0.47	n.d.

(n.d. = not detected)

Table 10 shows the library abundance of selected molecules that were up- or down-regulated at least once following treatment with various agents. Library abundance of each SEQ ID NO is presented as relative to that library which included the least abundant levels of nucleic acid molecule (SEQ ID NO) present.

TABLE 10 Library abundance (least abundant = 1) patterns of nucleic acid molecules

SEQ ID NO:	Untreated	CLO	FEN	APAP	BaP	CCl ₄	HYDR	ANIT	4-AAF
8	4	7	6	3	9	4	1	1	3
9	13	5	6	4	15	5	6	6	2
10	n.d.	1	8	3	n.d.	n.d.	n.d.	1	n.d.
11	5	2	4	8	20	7	10	n.d.	2

(n.d. = not detected)

XI Identification and Analyses of Homologous Molecule in other Organisms

The rat sequences (SEQ ID NOs:1-16) were used to identify additional sequences in the ZOOSEQ and LIFESEQ databases (Incyte Pharmaceuticals) related to rat nucleic acid molecules regulated during toxicological response (SEQ ID NOs:18-47).

The first pass cDNAs, SEQ ID NOs:5, and 60 through 134, were assembled using PHRAP (Phil Green, supra), using the following default parameters, to produce the contiguous sequence SEQ ID NO:135. Mismatch penalty = -2; gap initiation penalty <0; gap extension penalty <0; minimum length of matching word = 14; minimum SWAT score = 30; bandwidth = 14; use raw SW scores, "No"; index word size = 10; maximum gap size = 30; number of initial bases to be converted to 'N', 0; vector segment length = 60; Mismatch penalty for scoring degenerate end sequence = -2; Min. score for converting degenerate end sequence to 'N', 20; Minimum size of confirming segment = 8; Amount by which confirming segments are trimmed = 1; Penalty for confirming matches = -5; Min. SWAT score for confirming matches = 30; LLR cutoff for displaying discrepancies = 20; Minimum segment size = 8; Spacing between nodes = 4; Split/reassemble initial 'greedy' assembly, "No".

Translation of SEQ ID NO:135 using MACDNASIS PRO software (version 1.0, Hitachi Software Engineering) using default parameters of the program elucidated the putative protein coding region, SEQ ID NO:136. The nucleic acid and amino acid sequences were queried against databases such as the LIFESEQ (Incyte), GenBank, and SwissProt databases using BLAST. Motifs, HMM algorithms, and alignments with BLOCKS, PRINTS, Prosite, and PFAM databases were used to perform functional analyses; the antigenic index (Jameson-Wolf analysis) was determined using LASERGENE software (version 1.62d1, DNASTAR). BLAST2 analysis of SEQ ID NOs:135 and 136 using the human EST LIFESEQ database (Incyte) identified Incyte Clone Numbers 746355H1 (SEQ ID NO:137) and 1294663H1 (SEQ ID NO:138) which were assembled with their respective clustered clones to produce SEQ ID NOs:37 and 38 which encoded SEQ ID NOs:51 and 52, respectively.

Functional analysis of SEQ ID NO:136 using BLOCKS, PRINTS, Prosite, PFAM, Motifs, and HMM algorithms identified a potential protein kinase C phosphorylation site at residue S84 (Motifs); a potential signal peptide from residue M1 through residue A33 (SPScan); a potential transmembrane domain from residue P37 through residue L56 (HMM TM), a sodium/neurotransmitter symporter signature from residue G34 through A53, a sodium/alanine symporter signature from G34 through A53, and an asparaginase/glutaminase family signature from residue W64 through residue G75 (BLOCKS and PRINTS).

Functional analysis of SEQ ID NO:51 using BLOCKS, PRINTS, Prosite, PFAM, Motifs, and HMM algorithms identified a potential protein kinase C phosphorylation site at residue S83 (Motifs); a potential signal peptide sequence from residue M1 through residue A52 (SPScan); a sodium/alanine symporter signature from residue G33 through residue A52, an asparaginase/glutaminase family signature from residue W63 through residue G74, and a channel-forming colicin domain from residue K31 through residue G49 (BLOCKS and PRINTS). Functional analysis of SEQ ID NO:52 using BLOCKS, PRINTS, Prosite, PFAM, Motifs, and HMM algorithms identified a potential signal peptide sequence from residue M1 through A53 (SPScan); a sodium/alanine symporter signature from residue G34 through residue A53, a 6-phosphogluconate dehydrogenase family signature from residue G15 through residue A40, an FAD-dependant glycerol-3-phosphate dehydrogenase family signature from residue Y18 through residue Y30, and a vacuolar ATP synthetase 16 kDa subunit signature from residue L39 through residue G65 (BLOCKS and PRINTS).

CLAIMS

What is claimed is:

1. A method for detecting or diagnosing the effect of a toxic compound or molecule associated with increased or decreased levels of nucleic acid molecules in a mammalian subject comprising:
 - 5 a) treating a mammalian subject with a toxic compound or molecule;
 - b) obtaining a sample containing nucleic acids from the mammalian subject treated with the toxic compound or molecule;
 - c) contacting the sample with a microarray comprising a plurality of nucleic acid molecules of SEQ ID NOs:1-47, or a fragment thereof under conditions for the formation of one or
10 more hybridization complexes; and
 - d) detecting the hybridization complexes, wherein the presence, absence or change in amount of the hybridization complex, as compared with the hybridization complexes formed from nucleic acid molecules from an untreated mammalian subject, is indicative of a metabolic response to the toxic compound or molecule.
- 15 2. The method of claim 1 wherein:
 - a) the sample is a tissue chosen from liver, kidney, brain, spleen, pancreas, and lung;
 - b) the sample is liver tissue;
 - c) the toxic compound or molecule which elicits the metabolic response induces at least a 2-fold change in the amount of at least one of the nucleic acid molecules of the sample;
 - 20 d) the toxic compound is a peroxisome proliferator;
 - e) the toxic compound is a hypolipidemic drug; and
 - f) the toxic compound is clofibrate or one of its corresponding metabolites.
3. The method of claim 1 wherein:
 - a) the sample is a tissue chosen from liver, kidney, brain, spleen, pancreas, and lung;
 - 25 b) the sample is liver tissue;
 - c) the toxic compound or molecule which elicits the metabolic response induces at least a 2-fold change in the amount of at least one of the nucleic acid molecules of the sample;
 - d) the toxic compound is acetaminophen or one of its corresponding metabolites.
4. The method of claim 1 wherein:
 - 30 a) the sample is a tissue chosen from liver, kidney, brain, spleen, pancreas, and lung;
 - b) the sample is liver tissue;
 - c) the toxic compound or molecule which elicits the metabolic response induces at least a 2-fold change in the amount of at least one of the nucleic acid molecules of the sample;
 - d) the toxic compound is a polycyclic aromatic hydrocarbon;

- e) the toxic compound is a diol epoxide; and
 - f) the toxic compound is benzo(a)pyrene, or one of its corresponding metabolites.
5. A method for detecting or diagnosing a toxicological response to a test compound or molecule in a mammalian subject, the method comprising:
- 5 a) treating a mammalian subject with a test compound or molecule;
 - b) obtaining a sample containing nucleic acids from the mammalian subject treated with the test compound or molecule;
 - c) contacting the sample with a microarray comprising a plurality of nucleic acid molecules of SEQ ID NOs:1-47, or a fragment thereof, under conditions for the formation of one or more
 - 10 hybridization complexes;
 - d) detecting the hybridization complexes, wherein the presence, absence or change in amount of the hybridization complex, as compared with the hybridization complexes formed from nucleic acid molecules from a normal or untreated mammalian subject, is indicative of a toxic response to the test compound or molecule.
- 15 6. The method of claim 5 wherein the test compound which elicits the metabolic response is a compound with a previously known metabolic response.
7. The method of claim 5 wherein the test compound which elicits the metabolic response is a compound with a previously unknown metabolic response.
8. An isolated and purified nucleic acid molecule selected from SEQ ID NOs:1-11, 17-33, 36, 39,
- 20 and 41, or a fragment thereof, wherein said fragments are at least 60 contiguous nucleotides in length.
9. A method of using a molecule selected from SEQ ID NOs:1-59 or a fragment thereof to screen a library of molecules or compounds to identify at least one molecule or compound which specifically binds the selected molecule, the method comprising:
- 25 a) combining the selected molecule with a library of molecules or compounds under conditions to allow specific binding; and
 - b) detecting specific binding, thereby identifying a molecule or compound which specifically binds the selected molecule.
10. The method of claim 9 wherein the library is selected from DNA molecules, RNA molecules, peptide nucleic acids, artificial chromosome constructions, peptides, proteins, and drugs.
- 30 11. An isolated and purified antibody identified using the method of claim 9.
12. An isolated and purified nucleic acid molecule variant having at least 70% nucleic acid sequence identity to the nucleic acid molecule of claim 8.
13. An isolated and purified nucleic acid molecule having a sequence which is complementary to the nucleic acid molecule of claim 8.

14. An isolated and purified agonist identified using the method of claim 9.
15. An isolated and purified antagonist identified using the method of claim 9.
16. An expression vector comprising at least a fragment of the nucleic acid molecule of claim 8.
17. A host cell comprising the expression vector of claim 16.
- 5 18. A method for producing a polypeptide, the method comprising the steps of:
 - a) culturing the host cell of claim 17 under conditions suitable for the expression of the polypeptide; and
 - b) recovering the polypeptide from the host cell culture.
19. An isolated and purified protein molecule encoded by the nucleic acid molecule selected from
- 10 SEQ ID NOs:1-11, 17-33, 36, 39, and 41, an isolated and purified protein molecule of SEQ ID NOs:50 and 53, or a portion thereof, wherein said portions encode at least 20 contiguous amino acids in length.
20. A pharmaceutical composition comprising the protein molecule of claim 19 in conjunction with a suitable pharmaceutical carrier.

SEQUENCE LISTING

<110> INCYTE GENOMICS, INC.
 CUNNINGHAM, Mary Jane
 ZWEIGER, Gary
 KASER, Matthew R.
 PANZER, Scott
 SEILHAMMER, Jeffrey J.
 YUE, Henry
 BAUGHN, Mariah
 AZIMZAI, Yalda
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ttctgagagt gtggtgttca ctgacacaaa ttcaatcctg cgctacttgg c 291

```

<210> 3

<211> 293

<212> DNA

<213> Rattus norvegicus

<220>

<221> misc_feature

<223> Incyte ID No.: 700510669H1

<400> 3

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aggacctgtc cttacatatt gtggcctgaa gggacaaaat atgaggagtt naatannagg 60
acaattccac tggtttatatt ccttggtgct aaattaaaga atcaagccct tggtcgagcc 120
tttgaaattt tggcctactt tatttcagac actcaaaata caaatgccaa caaatggtn 180
tgatatattt gagagtggga aggaatctct gatgtttaaa ttctactgtt gatctttcaa 240
aatggactag gcttaggatt acaatgaacc tttgtcctt tgtcagtgtt tcg 293
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<210> 4

<211> 260

<212> DNA

<213> Rattus norvegicus

<220>

<221> misc_feature

<223> Incyte ID No.: 700525676H1

<400> 4

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gcagctcgga ctagtcagag gnetctggcg aggggtggcat cgggatgccg tccgaagtca 60
cccacagtga cggangcccg ggtgcgaggg tctgcgcgca acgtcaggta cttagctccc 120
tgtggtatac tgatgaacag aacccttgca ccgtgggcct cagttttgcc taaagagatc 180
tgtgcaagaa ccttcttcag aatcactaca ccattagtaa ataagcgana ggagtattca 240
gagaggagaa ttatagggtg 260
```

<210> 5

<211> 290

<212> DNA

<213> Rattus norvegicus

<220>

<221> misc_feature

<223> Incyte ID No.: 700535332H1

<400> 5

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aagggccagt tgcattccgca ccagtgctt gtaccttgaa ctcatttctt cctgactgct 60
agaggcctgt gtgttcttaa ctgctccgac ctctcctcca caggtgcagg cctgggtgtg 120
tctccaaagt gactgaacaa tgcagaagga cagtggccca ctggttcctt tacattatna 180
tggtttcggc tatgcggccc tgggtggctac tgggtgggatt attggctatg caaaagcagg 240
tatgtgacct ccctggctgc tggatcttct ttgggggcct ggcaggctgg 290
```

<210> 6

<211> 287

<212> DNA

<213> Rattus norvegicus

<220>

<221> misc_feature

<223> Incyte ID No.: 700536004H1

<400> 6

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attatgtaaa taatgagcaa gatcaaatta acaaagacta gttaccagc attccgcac 60
tagtcagttt tgatcatggg cagttcaagc tgccacctga gaacatcact aggtctctcag 120
ggttcttggc accactcacc caagttacat ccaccagatt attttcagtc ttcacaagta 180
tcaccatgca tagtgggatt ttcagccatg aataaagggc gtgcgttttg ccataatcag 240
ctctaaaata acctttgcta atcaatgcag tgagttgcta aggttta 287
```

<210> 7
 <211> 264
 <212> DNA
 <213> Rattus norvegicus

<220>
 <221> misc_feature
 <223> Incyte ID No.: 700640924H1

<400> 7

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gtgatgaaat gaggtatctc aaatccactg acagataaga aaacagggtt agaggggaaag 60
tcacctctgt cacgtagagg cagaatatat gaacttaact ctagtttcca tgtctgtctt 120
tattaccttc atctttctac ttcttgcca caggcatttc acttaattga gcctaattgc 180
agtatctgtg tgtgtcaatg tcgttaccac attctgatga agctaaaaaa taaaatttnn 240
tttgggcca aaaaaaaaaa aagg 264
```

<210> 8
 <211> 238
 <212> DNA
 <213> Rattus norvegicus

<220>
 <221> misc_feature
 <223> Incyte ID No.: 700775760H1

<400> 8

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ganaccgaca ttttaatggt tcttangagg accaccacta gagtcaagg ganaatggga 60
tgacgcgtgt tgcngtcctg ctgattctga caagagctgn tcactatgac agacagatgg 120
actgaatgga ctagaattat gtgaatctgt attatttaca gttggtancc aagagcatcg 180
atactcttta gagaggcagg ttaaataaag gattaagtat ttaggatntg aaatttat 238
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<210> 9
 <211> 112
 <212> DNA
 <213> Rattus norvegicus

<220>
 <221> misc_feature
 <223> Incyte ID No.: 700132084H1

<400> 9

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ctatgcccaa ggaaaaggct ccagaacaca ttccccttct cttcattgcc ttcccatcaa 60
gcaaggatcc aacctgggag gaccgattcc cagnnncggg ncannaagnn gg 112
```

<210> 10
 <211> 238
 <212> DNA
 <213> Rattus norvegicus

<220>
 <221> misc_feature
 <223> Incyte ID No.: 700176719H1

<400> 10

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tcttggtccc ttcacctgac ctccggtgct ccaacgggcg gcagaatgga agaaggtgag 60
gaccaggaa gtctgattaa agtgatccac ttgctgggtct tgtctggtgt ctggggcatg 120
cagatgtggg tgacctttgc ctcaggcttc ctgcttttcc ggagcctccc gaggcacacg 180
tttggaactg tgcagagcaa gctcttccca gtctattttc acgtctcctt gggtttgtg 238
```


<210> 11
 <211> 247
 <212> DNA
 <213> Rattus norvegicus

<220>
 <221> misc_feature
 <223> Incyte ID No.: 701195696H1

<400> 11

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ggatctttct gggcgagcaa cccgcaaaac gttgtgcatt gcgttgaaaa ggtgcatctg 60
gttcccgatt ctactcccca cccgcgaccg cacacagcaa acatgaccca gcagccgcct 120
gacgttgagg aggatgactg tctttctgaa taccaccacc tcttctgccc ggaccttctc 180
caggacaaag tggcttttat cactggtggt ggttctggga ttggcttcg gatcgccgag 240
attttca. 247

```

<210> 12
 <211> 256
 <212> DNA
 <213> Rattus norvegicus

<220>
 <221> misc_feature
 <223> 700483259H1

<400> 12

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gtgacgtaca tggaaaacaa agcctacggg gacaggctca agccgcagac agcagcaagt 60
aaagcgctn cggcctgaa gcatggcagc tatcccttcc agcggctcgc tcgtggctac 120
ccatgactac tatcggcgta agtagccct cgccagcccc gcccgaggct ggcccagggc 180
tctgtggctg acccgctcc ccttcccagg acgtctgggc tctcgtcca gcaacagctc 240
cggcggaagt gcagag 256

```

<210> 13
 <211> 285
 <212> DNA
 <213> Rattus norvegicus

<220>
 <221> misc_feature
 <223> Incyte ID No.: 700607235H1

<400> 13

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ctgaagaccc accatgtctc tgctgactac tgtactactt ctctgggggt tcattctggg 60
cccagcaact gacacagcct gtatattcaa ggaagcctcg gaaaacagtc ccttgcccag 120
gccttggett tctgccaate cagtgccttg gatcacacct ggctgagga cattcctgct 180
gtgccagggg acagtgcggg atgtagtctt catgctgagg cgggaaggag atgatgggtt 240
cctggcgata gtccaacaga tgtttttctg gagggagctg gacct 285

```

<210> 14
 <211> 293
 <212> DNA
 <213> Rattus norvegicus

<220>
 <221> misc_feature
 <223> Incyte ID No.: 700609074H1

<400> 14

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ggcgtggagt tggagnagag cgtcaggcgc ctccgggaga agtttcatgg aaaagtgtcc 60
cccaagaagg caggggctct tatgaggaag tttggcagcg accacactgg agttgggcgc 120
tctatcgtgt acgggctcaa gcagaaagat ggacaggagc tgagcaacga tttggacgnc 180

```

caggacccac cagaggacat gaagcaggac caagatatcc aggcagtagc cacctctctg 240
ttgcccctga cgcaagccaa tcttcgaatg ttccaaagag cccaagatga cct 293

<210> 15
<211> 268
<212> DNA
<213> Rattus norvegicus

<220>
<221> misc_feature
<223> Incyte ID No.: 700627890H1

<400> 15
gtacaangag ngccggggct tgggtctagt tggaggggan gcagtggcca gtncagggt 60
cagatgagag agttagccga gttaggggca gctactagga tgggggcagg aggagaagcg 120
gggctaacta taaagaagac tagatttcgn cacagtgggt atgtggaagg cagctttcaa 180
accgcccttg tcaaacaaca cagggccagc agccttcaag accaggctat ccctgccgtc 240
tgctggcatg ggggcacttg taccgtcc 268

<210> 16
<211> 265
<212> DNA
<213> Rattus norvegicus

<220>
<221> misc_feature
<223> Incyte ID No.: 700629293H1

<400> 16
atgaccttta acttttctaa aaatgtgaag ttttgtactt atatatatca gctaaagtat 60
tntagcattc tttagtgtac ttagtttgat gccacttta gtgtttttgt tgcttttgtc 120
tgatttttat gaatgttcac tttaagactc cttgttgaaa tgggacagtt tcgttctttg 180
ataagccccga gaagaggatt cccttgggtg ttgacctcct ctgcatgatg tgcccaagca 240
tctgaactgc aaccaaggcc tttn 265

<210> 17
<211> 267
<212> DNA
<213> Mus musculus

<220>
<221> misc_feature
<223> Incyte ID No.: 701322438H1

<400> 17
acctgccctt acatattgtg gcctgaagng acaaaatatg agaagttcaa tgaaaagata 60
attccccctt tcaggaaaga tgttctctta ttttacttgg cgctaaatca aagaatcaag 120
cctttgttca agcctttgca attttggcct attttatttc agagagcaaa tggatgggtat 180
atatttggga gtgggaaggn tctttgattt tttaaatttca ctgntgagct ttcaaataga 240
ctaggcctta ggattacaat gaacaac 267

<210> 18
<211> 239
<212> DNA
<213> Mus musculus

<220>
<221> misc_feature
<223> Incyte ID No.: 701082352H2

<400> 18

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atttcttagt ggggcaagga cctgccctta catattgtgg cctgaaggga caaaatatga 60
gaagttcaat gaaaagataa ttcccccttt caggaaagat gttctcttat tttacttggn 120
gctaaatcaa agaattccagc cctttgttca agcctttgca attttggcct attttatttc 180
agagagcaaa tggttgttat atatttggga gtgggaagga atcttgattt ttaaatttc 239

```

```

<210> 19
<211> 244
<212> DNA
<213> Mus musculus

```

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<220>
<221> misc_feature
<223> Incyte ID No.: 701423834H1

```

```

<400> 19

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gtctcctgag tgcttaaatt acaggtgtgt accactaaac caaccctaag aatccatttt 60
aaaatgtcag tcactttaga tttcttagtg gggcaaggac ctgcccttac atattgtggc 120
ctgaagggac aaaatatgag aagttcaang aaaagataat tccccctttc aggaaagatg 180
ttctcttatt ntacttggtg ctaaatacaa gaatcaagcc tttgttcaag cctttgcaat 240
tntg 244

```

```

<210> 20
<211> 240
<212> DNA
<213> Mus musculus

```

```

<220>
<221> misc_feature
<223> Incyte ID No.: 701423842H1

```

```

<400> 20

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gtctcctgag tgcttaaatt acaggtgtgt accactaaac caaccctaag aatccatttt 60
aaaatgtcag tcactttana tttcttagng gggcaaggac ctgcccttac atattggggc 120
ctgaagggac aaaatatgag aagttcaatg nanagntnan tccccctttc aggaaagatg 180
gtctcttatt ttacttgng ctaaatacaa gaatcaagcc tttgntcaag cctttgcaat 240

```

```

<210> 21
<211> 224
<212> DNA
<213> Mus musculus

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```

<220>
<221> misc_feature
<223> Incyte ID No.: 701090430H1

```

```

<400> 21

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ggcagctcgg accagtcaga gggccctggc gaggggtggca tcggggtgcc atccgaagtc 60
gaccaccgtg acggaagccc cggcgcgggg gtctgcgcgc gacgtcagac acttagctgc 120
ctgtggtgta ctgataaaca gaacccttcc accgtgtgct gcagttttgc ctaaagagat 180
ctgtgcgaga actttcttca gantctctgc gccactagta aata 224

```

```

<210> 22
<211> 249
<212> DNA
<213> Mus musculus

```

```

<220>
<221> misc_feature
<223> Incyte ID No.: 700966369H1

```

```

<400> 22

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gcttttatgt ancccaatca gagcancgac cagnaaaatt gcaagtnttg agaggcacac 60
agcagaagan ctgcagantt ctgcttgatt ggcatctatc gttcctcctg agcagcaacc 120
cacagtagat aggaaaaagg tgtttgacag gcctgggctaa gctcttgagg agccactggc 180
atcagatggc gaagggactt gctgccaggt tgctgtctgt tggacagaag ctengatgag 240
gtgtgctgg 249

```

```

<210> 23
<211> 260
<212> DNA
<213> Mus musculus

```

```

<220>
<221> misc_feature
<223> Incyte ID No.: 700828522H1

```

```

<400> 23

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caggcctggt gtggtctcca aagcgactga acaatgcaga aagacagtgg ccattgatg 60
cctttacatt attttggttt cggctatgca gccctggttg ctaccgggtg gattattggc 120
tatgccaaag caggtagtgt gccgtccctg gctgctggac tcttcttcgg gggcctggca 180
ggcctggggg cctaccagct gtctcaggat ccaggaatg tgtggggttt cctagctaca 240
tctgggacct tggccggaat 260

```

```

<210> 24
<211> 246
<212> DNA
<213> Mus musculus

```

```

<220>
<221> misc_feature
<223> Incyte ID No.: 701250723H1

```

```

<400> 24

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ctcggcttct cgctgtctgc tcgcgccttc gtctctacagc acaggcctcc cggctccggc 60
tccggctcca gtgttggttg ggtgcaggcc tgggtgtggtc tccaaagcga ctgaacaatg 120
cagaaagaca gtggcccatt gatgccttta cattattttg gtttcggcta tgcagccctg 180
gttgctaccg gtgggattat tggctatgcc aaagcaggta gtgtgccgtc cctgggtgct 240
ggactc 246

```

```

<210> 25
<211> 252
<212> DNA
<213> Mus musculus

```

```

<220>
<221> misc_feature
<223> Incyte ID No.: 701254093H1

```

```

<400> 25

```

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acctcggctt ctcgctgtct gctcgcgccc tcgtcttaca gcacaggcct cccggctccg 60
gcttcgggct ccagtgttg ttgggatgcc ttacattat tttggtttcg gctatgcagc 120
cctggttgct accggtggga ttattggcta tgccaaagca ggtagtgtgc cgtccctggc 180
tgetggactc ttcttcgggg gcctggcagg cctggggggt accagctgtc tcaggatccc 240
aggaatgtgt gg 252

```

```

<210> 26
<211> 237
<212> DNA
<213> Mus musculus

```

```

<220>
<221> misc_feature

```

<223> Incyte ID No.: 701423901H1

<400> 26

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atTTtggTtt cggctatgca gccctggttg ctaccggtgg gattattggc tatgccaaag 60
caggtagtgt gccgccctgg ctgctggact cttcttcggg ggccctggcag gcctggggcc 120
taccagctgt ctcaggatcc caggaatgtg tgggttttcc tagctacatc tgggaccttg 180
ccggaattat ggggatgaga ttctacaact cggggaaatt tatnctgcag gntaatc 237
```

<210> 27

<211> 274

<212> DNA

<213> Mus musculus

<220>

<221> misc_feature

<223> Incyte ID No.: 701251161H1

<400> 27

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ggtgtttcgt gggttatctt tgcaaatggg ctccgcggcc tagcgccctg gtggcctaa 60
aacgaagcct gcaaggaagg ggttctccgc cgagcgccctc ggtcctgaag catggcagcc 120
atcccttcca gcggctcgct cgtggctacc catgactact atcggcgtaa gtagccctc 180
gccagccccc cccagggctg gcccagggcc ctgtggctga cccgcctccc ctcccagga 240
cgctgggct cctcgccag cagcagctcc ggcg 274
```

<210> 28

<211> 141

<212> DNA

<213> Mus musculus

<220>

<221> misc_feature

<223> Incyte ID No.: 701085115H2

<400> 28

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aaagtgtccc ccaanaaggc aggggctctt atgaggaagt ttggcagcna ccacaccgga 60
gttggngct ctatcgtgta tgggtgcaag cagnaagacg gacangagct gatgcaacga 120
cctggacgct caggaccac c 141
```

<210> 29

<211> 274

<212> DNA

<213> Mus musculus

<220>

<221> misc_feature

<223> Incyte ID No.: 701387375H1

<400> 29

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ggagggctcg ctcttggggc tagtggtggg gaggcagtgg ccagttcagg gctcagatga 60
gagaggtggc agaattagag gcagccacta ggatgggggt gcnaggagaa gcgggggctaa 120
gtataaagga nactagattt tgggacagtg gacgtgtgga aggcagcttc caaagcgct 180
ttaacaatcc acaaagaacc agnngctttc aagaccaggc tatccctgct gnetgctgna 240
cttggacgtn caggangcac angtttcaca ggcg 274
```

<210> 30

<211> 257

<212> DNA

<213> Mus musculus

<220>
 <221> misc_feature
 <223> Incyte ID No.: 701389479H1

<400> 30

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agggtctgct cttggggcta gtggtgggga ggcagtggcc agntcagggc tnagatgaga 60
gangtggcag aattagaggc agccactagg atgggggtgc gaggagaagc ggggctaagt 120
ataaaggaga ctanattttg ggacagtgga cgtgtggaag gcagnttnca aagcgcttt 180
aacaatccac anagaaccag cagctttcaa gaccangcta tccctgctgc tgctgcactt 240
gacgtcagga ngnacaa 257
```

<210> 31
 <211> 246
 <212> DNA
 <213> Mus musculus

<220>
 <221> misc_feature
 <223> Incyte ID No.: 701389530H1

<400> 31

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caaggagggc tcgctcttgg ggctagtggg gngagggcag nggccagttc agggctcaga 60
tganagaggc ggcanaatta gaggcagcca ctaggatggg ggtgccgagg agaagcgggg 120
ctaagtataa aggagactag attttgggac agtggacgtg tggaaggcag cttccaaagc 180
gcctttaaca atccacaaag aaccagcagc tttcaagacc angctatccc tgctgctgct 240
gcactt 246
```

<210> 32
 <211> 258
 <212> DNA
 <213> Mus musculus

<220>
 <221> misc_feature
 <223> Incyte ID No.: 701388372H1

<400> 32

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gagggctcgc tcttggnggc taagnngtgg ggagtcagtg gccacgttca gggctcanat 60
gagagaggtg gcagaattag aggcagccac taggatggg gngccaggag aagcnggcta 120
agtataaagg agactagatt ttgggacagt ggacgtgngg aaggcagctt ccaaagcgcc 180
tttaacaatc cacanagaac cagnagcttt caaagaccag gctatccctc tgctgctggc 240
acttgacgtc cagaaggc 258
```

<210> 33
 <211> 257
 <212> DNA
 <213> Mus musculus

<220>
 <221> misc_feature
 <223> Incyte ID No.: 701270715H1

<400> 33

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gtttttctcat gaattgtttt tgcattgttg ataaagctag tatacccttt ggccttagcc 60
tataaatttt aaatatataa acaaaatatt aaagatgtag ttaattttta atgaccttta 120
acttttctaa aaatgtgaag ttttgtactt acatatcatc taaagtatta tagcatTTTT 180
aagtgtactt agtttgatgc cacttttagt gttttgttgc ttttgtctga tttttgtgaa 240
tgttcatnta agactcc 257
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<210> 34

<211> 4850

<212> DNA

<213> Homo sapiens

<220>

<221> misc_feature

<223> Incyte ID No.: 2302721

<400> 34

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cgcacacggt gcatcttctt cctttcggcg ggtcctccgt agttctggca cgagccaggc 60
gtactgacag gtggaccagc ggactggtgg agatggcgac gctctctctg accgtgaatt 120
caggagaccc tccgctagga gctttgctgg cagtagaaca cgtgaaagac gatgtcagca 180
tttccggtga agaagggaaa gagaatattc ttcattgttc tgaaaatgtg atattcacag 240
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ctaactctgat ggaacatact gagattgata actgggttga gttcagtgct acaaaaattat 360
cttcatgtga ttcccttact tctacaatta atgaactcaa tcattgcctg tctctgagaa 420
catacttagt tggaaactcc ttgagtttag cagatttatg tggttggggc accctaaaag 480
gaaatgctgc ctggcaagaa cagttagaac agaagaaagc tccagttcat gtaaaacggt 540
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caacaaccaa agctcgagtg gcacctgaga aaaagcaaga tgttgggaaa tttgttgagc 660
ttccaggttg ggagatggga aaggttaccg tcagatttcc tccagaggcc agtgggtact 720
tacacgaattg ctatgcacaaa gactgctctt tgaaccagca ctaccagggt aactttaaag 780
ggaaactgat catgagattt gatgacacaa atcctgaaaa agaaaaggaa gattttgaga 840
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cggatcattt tgaactata atgaagtatg cagagaagct aattcaagaa gggaagggtt 960
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gccagtttgg tcagtcctgt tgtttgagag caaaaattga catgagtagt aacaatggat 1140
gcatgagaga tccaacctt tatcgtgca aaattcaacc acatccaaga actggaaata 1200
aatacaatgt ttatccaaca tatgattttg cctgccccat agttgacagc atcgaagggt 1260
ttacacatgc cctgagaaca acagaatacc atgacagaga tgagcagttt tactggatta 1320
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gactgaaaca gtttattgct gctcagggtt cctcacgttc agtcgtgaac atggagtggt 1560
acaaaatctg ggcgtttaac aaaaagggtt ttgacccagt ggctccacga tatgttgcat 1620
tactgaagaa agaagtgat ccagtgaatg tactgaagc tcaggaggag atgaaagaag 1680
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attggggcaa cctcaacatt acaaaaatac acaaaaatgc agatggaaaa atcatatctc 1860
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<213> Homo sapiens

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<221> misc_feature

<223> Incyte ID No.: 2742442

<400> 35

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<400> 36

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<211> 1263
<212> DNA
<213> Homo sapiens

<220>

<221> misc_feature

<223> Incyte ID No.: 1968009

<400> 37

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<210> 38

<211> 978

<212> DNA

<213> Homo sapiens

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<223> Incyte ID No.: 1923127

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<210> 39

<211> 851

<212> DNA

<213> Homo sapiens

<220>

<221> misc_feature

<223> Incyte ID No.: 3123954

<400> 39

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<210> 40

<211> 1907

<212> DNA

<213> Homo sapiens

<220>

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<223> Incyte ID No.; 1321844

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<212> DNA

<213> Homo sapiens

<220>

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<223> Incyte ID No.: 375724.9

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<211> 408
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<213> Homo sapiens

<220>
<221> misc_feature
<223> Incyte ID No.: 375724.3

<400> 42

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<211> 3136
<212> DNA
<213> Homo sapiens

<220>
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<400> 43

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<210> 44
 <211> 1376
 <212> DNA
 <213> Homo sapiens

<220>
 <221> misc_feature
 <223> Incyte ID No.: 1461451

<400> 44

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<210> 45
 <211> 649
 <212> DNA
 <213> Homo sapiens

<220>
 <221> misc_feature
 <223> Incyte ID No.: 2345712

<400> 45

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<210> 46

<211> 1554

<212> DNA

<213> Homo sapiens

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<221> misc_feature

<223> Incyte ID No.: 1810320

<400> 46

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<210> 47

<211> 1083

<212> DNA

<213> Homo sapiens

<220>

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<223> Incyte ID No.: 964996

<400> 47

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<210> 48

<211> 1512

<212> PRT

<213> Homo sapiens

<220>

<221> misc_feature

<223> Incyte ID No.: 2302721

<400> 48

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  35      40      45
Val Ile Phe Thr Asp Val Asn Ser Ile Leu Arg Tyr Leu Ala Arg
  50      55      60
Val Ala Thr Thr Ala Gly Leu Tyr Gly Ser Asn Leu Met Glu His
  65      70      75
Thr Glu Ile Asp His Trp Leu Glu Phe Ser Ala Thr Lys Leu Ser
  80      85      90
Ser Cys Asp Ser Phe Thr Ser Thr Ile Asn Glu Leu Asn His Cys
  95     100     105
Leu Ser Leu Arg Thr Tyr Leu Val Gly Asn Ser Leu Ser Leu Ala
 110     115     120
Asp Leu Cys Val Trp Ala Thr Leu Lys Gly Asn Ala Ala Trp Gln
 125     130     135
Glu Gln Leu Lys Gln Lys Lys Ala Pro Val His Val Lys Arg Trp
 140     145     150
Phe Gly Phe Leu Glu Ala Gln Gln Ala Phe Gln Ser Val Gly Thr
 155     160     165
Lys Trp Asp Val Ser Thr Thr Lys Ala Arg Val Ala Pro Glu Lys
 170     175     180
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Gly Lys Val Thr Val Arg Phe Pro Pro Glu Ala Ser Gly Tyr Leu
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His Ile Gly His Ala Lys Ala Ala Leu Leu Asn Gln His Tyr Gln
 215     220     225
Val Asn Phe Lys Gly Lys Leu Ile Met Arg Phe Asp Asp Thr Asn
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Pro Glu Lys Glu Lys Glu Asp Phe Glu Lys Val Ile Leu Glu Asp
 245     250     255
Val Ala Met Leu His Ile Lys Pro Asp Gln Phe Thr Tyr Thr Ser
 260     265     270
Asp His Phe Glu Thr Ile Met Lys Tyr Ala Glu Lys Leu Ile Gln
 275     280     285
Glu Gly Lys Ala Tyr Val Asp Asp Thr Pro Ala Glu Gln Met Lys
 290     295     300
Ala Glu Arg Glu Gln Arg Ile Glu Ser Lys His Arg Lys Asn Pro

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Ile Glu Lys Asn	305	Leu Gln Met Trp Glu	310	Met Lys Lys Gly	315
Gln Phe Gly Gln	320	Ser Cys Cys Leu Arg	325	Lys Ile Asp Met	330
Ser Asn Asn Gly	335	Cys Met Arg Asp Pro	340	Leu Tyr Arg Cys	345
Ile Gln Pro His	350	Pro Arg Thr Gly Asn	355	Lys Tyr Asn Val Tyr	360
Thr Tyr Asp Phe	365	Ala Cys Pro Ile Val	370	Ser Ile Glu Gly	375
Thr His Ala Leu	380	Arg Thr Thr Glu Tyr	385	His Asp Arg Asp Glu	390
Phe Tyr Trp Ile	395	Ile Glu Ala Leu Gly	400	Arg Lys Pro Tyr	405
Trp Glu Tyr Ser	410	Arg Leu Asn Leu Asn	415	Thr Val Leu Ser	420
Arg Lys Leu Thr	425	Phe Val Asn Glu Gly	430	Leu Val Asp Gly	435
Asp Asp Pro Arg	440	Phe Pro Thr Val Arg	445	Val Leu Arg Arg	450
Met Thr Val Glu	455	Gly Leu Lys Gln Phe	460	Ala Ala Gln Gly	465
Ser Arg Ser Val	470	Val Asn Met Glu Trp	475	Lys Ile Trp Ala	480
Asn Lys Lys Val	485	Ile Asp Pro Val Ala	490	Pro Arg Tyr Val Ala	495
Leu Lys Lys Glu	500	Val Ile Pro Val Asn	505	Pro Glu Ala Gln	510
Glu Met Lys Glu	515	Val Ala Lys His Pro	520	Lys Asn Pro Glu Val	525
Leu Lys Pro Val	530	Trp Tyr Ser Pro Lys	535	Val Phe Ile Glu Gly	540
Asp Ala Glu Thr	545	Phe Ser Glu Gly Glu	550	Met Val Thr Phe Ile	555
Trp Gly Asn Leu	560	Asn Ile Thr Lys Ile	565	His Lys Asn Ala Asp	570
Lys Ile Ile Ser	575	Leu Asp Ala Lys Leu	580	Asn Leu Glu Asn Lys	585
Tyr Lys Lys Thr	590	Thr Lys Val Thr Trp	595	Leu Ala Glu Thr Thr	600
Ala Leu Pro Ile	605	Pro Val Ile Cys Val	610	Thr Tyr Glu His Leu	615
Thr Lys Pro Val	620	Leu Gly Lys Asp Glu	625	Asp Phe Lys Gln Tyr	630
Asn Lys Asn Ser	635	Lys His Glu Glu Leu	640	Met Leu Gly Asp Pro	645
Leu Lys Asp Leu	650	Lys Lys Gly Asp Ile	655	Ile Gln Leu Gln Arg	660
Gly Phe Phe Ile	665	Cys Asp Gln Pro Tyr	670	Glu Pro Val Ser Pro	675
Ser Cys Lys Glu	680	Ala Pro Cys Val Leu	685	Ile Tyr Ile Pro Asp	690
His Thr Lys Glu	695	Met Pro Thr Ser Gly	700	Ser Lys Glu Lys Thr	705
Val Glu Ala Thr	710	Lys Asn Glu Thr Ser	715	Ala Pro Phe Lys Glu	720
Pro Thr Pro Ser	725	Leu Asn Asn Asn Cys	730	Thr Thr Ser Glu Asp	735
Leu Val Leu Tyr	740	Asn Arg Val Ala Val	745	Gln Gly Asp Val Val	750
Glu Leu Lys Ala	755	Lys Lys Ala Pro Lys	760	Glu Asp Val Asp Ala	765
Val Lys Gln Leu	770	Leu Ser Leu Lys Ala	775	Glu Tyr Lys Glu Lys	780
Gly Gln Glu Tyr	785	Lys Pro Gly Asn Pro	790	Pro Ala Glu Ile Gly	795
Asn Ile Ser Ser	800	Asn Ser Ser Ala Ser	805	Ile Leu Glu Ser Lys	810

Leu Tyr Asp Glu	815	820	825
Val Ala Ala Gln Gly	830	Val Val Arg Lys	Leu
Lys Ala Glu Lys	835	Ile Asn Glu Ala Val	840
845	850	855	
Cys Leu Leu Ser	860	Lys Lys Thr Gly	Lys
865	870		
Glu Tyr Ile Pro	875	Gln Ser Ser Asp	Ser
880	885		
Ser Pro Thr Arg	890	Leu Glu Thr Pro	Glu
895	900		
Ala Lys Val Leu	905	Gln Gly Glu Val	Val
910	915		
Arg Lys Leu Lys	920	Asp Gln Val Asp	Ile
925	930		
Ala Val Gln Glu	935	Leu Leu Gln Leu Lys	Leu
940	945		
Ile Gly Val Glu	950	Ala Thr Gly Ala Glu	Asp
955	960		
Lys Asp Lys Lys	965	Lys Lys Glu Lys Glu	Asn
970	975	Lys Ser Glu Lys	Gln
Asn Lys Pro Gln	980	Gln Asn Asp Gly	Arg
985	990	Lys Asp Pro	Ser
Lys Asn Gln Gly	995	Gly Leu Ser Ser	Gly
1000	1005	Gly Ala Gly Glu	Gly
Gln Gly Pro Lys	1010	Gln Thr Arg Leu Gly	Leu
1015	1020	Leu Glu Ala Lys	Lys
Glu Glu Asn Leu	1025	Ala Asp Trp Tyr Ser	Gln
1030	1035	Val Ile Thr Lys	Ser
Glu Met Ile Glu	1040	Tyr His Asp Ile Ser	Gly
1045	1050	Cys Tyr Ile Leu	Arg
Pro Trp Ala Tyr	1055	Ala Ile Trp Glu Ala	Ile
1060	1065	Lys Asp Phe Phe	Asp
Ala Glu Ile Lys	1070	Lys Leu Gly Val Glu	Asn
1075	1080	Cys Tyr Phe Pro	Met
Phe Val Ser Gln	1085	Ser Ala Leu Glu Lys	Glu
1090	1095	Lys Thr His Val	Ala
Asp Phe Ala Pro	1100	Glu Val Ala Trp Val	Thr
1105	1110	Arg Ser Gly Lys	Thr
Glu Leu Ala Glu	1115	Pro Ile Ala Ile Arg	Pro
1120	1125	Thr Ser Glu Thr	Val
Met Tyr Pro Ala	1130	Tyr Ala Lys Trp Val	Gln
1135	1140	Ser His Arg Asp	Leu
Pro Ile Lys Leu	1145	Asn Gln Trp Cys Asn	Val
1150	1155	Val Arg Trp Glu	Phe
Lys His Pro Gln	1160	Pro Phe Leu Arg Thr	Arg
1165	1170	Glu Phe Leu Trp	Gln
Glu Gly His Ser	1175	Ala Phe Ala Thr Met	Glu
1180	1185	Glu Glu Ala Ala Glu	Glu
Val Leu Gln Ile	1190	Leu Asp Leu Tyr Ala	Gln
1195	1200	Val Tyr Glu Glu	Leu
Leu Ala Ile Pro	1205	Val Lys Gly Arg Lys	Thr
1210	1215	Thr Glu Lys Glu	Lys
Phe Ala Gly Gly	1220	Asp Tyr Thr Thr Thr	Ile
1225	1230	Glu Ala Phe Ile	Ser
Ala Ser Gly Arg	1235	Ala Ile Gln Gly Gly	Thr
1240	1245	Ser His His Leu	Gly
Gln Asn Phe Ser	1250	Lys Met Phe Glu Ile	Val
1255	1260	Phe Glu Asp Pro	Lys
Ile Pro Gly Glu	1265	Lys Gln Phe Ala Tyr	Gln
1270	1275	Asn Ser Trp Gly	Leu
Thr Thr Arg Thr	1280	Ile Gly Val Met Thr	Met
1285	1290	Val His Gly Asp	Asn
Met Gly Leu Val	1295	Leu Pro Pro Arg Val	Ala
1300	1305	Cys Val Gln Val	Val
Ile Ile Pro Cys	1310	Gly Ile Thr Asn Ala	Leu
1315	1320	Ser Glu Glu Asp	Lys
Glu Ala Leu Ile	1325	Ala Lys Cys Asn Asp	Tyr
		Arg Arg Arg Leu	Leu

	1325		1330		1335
Ser Val Asn Ile Arg	Val Arg Ala Asp Leu	Arg Asp Asn Tyr Ser			
	1340		1345		1350
Pro Gly Trp Lys Phe	Asn His Trp Glu Leu	Lys Gly Val Pro Ile			
	1355		1360		1365
Arg Leu Glu Val Gly	Pro Arg Asp Met Lys	Ser Cys Gln Phe Val			
	1370		1375		1380
Ala Val Arg Arg Asp	Thr Gly Glu Lys Leu	Thr Val Ala Glu Asn			
	1385		1390		1395
Glu Ala Glu Thr Lys	Leu Gln Ala Ile Leu	Glu Asp Ile Gln Val			
	1400		1405		1410
Thr Leu Phe Thr Arg	Ala Ser Glu Asp Leu	Lys Thr His Met Val			
	1415		1420		1425
Val Ala Asn Thr Met	Glu Asp Phe Gln Lys	Ile Leu Asp Ser Gly			
	1430		1435		1440
Lys Ile Val Gln Ile	Pro Phe Cys Gly Glu	Ile Asp Cys Glu Asp			
	1445		1450		1455
Trp Ile Lys Lys Thr	Thr Ala Arg Asp Gln	Asp Leu Glu Pro Gly			
	1460		1465		1470
Ala Pro Ser Met Gly	Ala Lys Ser Leu Cys	Ile Pro Phe Lys Pro			
	1475		1480		1485
Leu Cys Glu Leu Gln	Pro Gly Ala Lys Cys	Val Cys Gly Lys Asn			
	1490		1495		1500
Pro Ala Lys Tyr Tyr	Thr Leu Phe Gly Arg	Ser Tyr			
	1505		1510		

<210> 49
 <211> 238
 <212> PRT
 <213> Homo sapiens

<220>
 <221> misc_feature
 <223> Incyte ID No.: 2742442

<400> 49

Met Ala Ala Arg Thr	Gly His Thr Ala Leu	Arg Arg Val Val Ser
1	5	10
Gly Cys Arg Pro Lys	Ser Ala Thr Ala Ala	Gly Ala Gln Ala Pro
	20	25
Val Arg Asn Gly Arg	Tyr Leu Ala Ser Cys	Gly Ile Leu Met Ser
	35	40
Arg Thr Leu Pro Leu	His Thr Ser Ile Leu	Pro Lys Glu Ile Cys
	50	55
Ala Arg Thr Phe Phe	Lys Ile Thr Ala Pro	Leu Ile Asn Lys Arg
	65	70
Lys Glu Tyr Ser Glu	Arg Arg Ile Leu Gly	Tyr Ser Met Gln Glu
	80	85
Met Tyr Asp Val Val	Ser Gly Val Glu Asp	Tyr Lys His Phe Val
	95	100
Pro Trp Cys Lys Lys	Ser Asp Val Ile Ser	Lys Arg Ser Gly Tyr
	110	115
Cys Lys Thr Arg Leu	Glu Ile Gly Phe Pro	Pro Val Leu Glu Arg
	125	130
Tyr Thr Ser Val Val	Thr Leu Val Lys Pro	His Leu Val Lys Ala
	140	145
Ser Cys Thr Asp Gly	Arg Leu Phe Asn His	Leu Glu Thr Ile Trp
	155	160
Cys Phe Ser Pro Gly	Leu Pro Gly Tyr Pro	Arg Thr Cys Thr Leu
	170	175
Asp Phe Ser Ile Ser	Phe Glu Phe Arg Ser	Leu Leu His Ser Gln
	185	190
Leu Ala Thr Leu Phe	Phe Asp Glu Val Val	Lys Gln Met Val Ala
	200	205

Ala	Phe	Glu	Arg	Arg	Ala	Cys	Lys	Leu	Tyr	Gly	Pro	Glu	Thr	Asn
				215					220					225
Ile	Pro	Arg	Glu	Leu	Met	Leu	His	Glu	Val	His	His	Thr		
				230					235					

<210> 50
 <211> 653
 <212> PRT
 <213> Homo sapiens

<220>
 <221> misc_feature
 <223> Incyte ID No.: 3511087

<400> 50

Met	Pro	Phe	Ser	Ala	Ser	Leu	Leu	Gly	Thr	Leu	Pro	Ile	Gly	Ala
1				5					10					15
Arg	Tyr	Ala	Pro	Pro	Pro	Ser	Phe	Ser	Glu	Phe	Tyr	Pro	Pro	Leu
				20					25					30
Thr	Ser	Ser	Leu	Glu	Asp	Phe	Cys	Ser	Ser	Leu	Asn	Ser	Phe	Ser
				35					40					45
Met	Ser	Glu	Ser	Lys	Arg	Asp	Leu	Ser	Thr	Ser	Thr	Ser	Arg	Glu
				50					55					60
Gly	Thr	Pro	Leu	Asn	Asn	Ser	Asn	Ser	Ser	Leu	Leu	Leu	Met	Asn
				65					70					75
Gly	Pro	Gly	Ser	Leu	Phe	Ala	Ser	Glu	Asn	Phe	Leu	Gly	Ile	Ser
				80					85					90
Ser	Gln	Pro	Arg	Asn	Asp	Phe	Gly	Asn	Phe	Phe	Gly	Ser	Ala	Val
				95					100					105
Thr	Lys	Pro	Ser	Ser	Ser	Val	Thr	Pro	Arg	His	Pro	Leu	Glu	Gly
				110					115					120
Thr	His	Glu	Leu	Arg	Gln	Ala	Cys	Gln	Ile	Cys	Phe	Val	Lys	Ser
				125					130					135
Gly	Pro	Lys	Leu	Met	Asp	Phe	Thr	Tyr	His	Ala	Asn	Ile	Asp	His
				140					145					150
Lys	Cys	Lys	Lys	Asp	Ile	Leu	Ile	Gly	Arg	Ile	Lys	Asn	Val	Glu
				155					160					165
Asp	Lys	Ser	Trp	Lys	Lys	Ile	Arg	Pro	Arg	Pro	Thr	Lys	Thr	Asn
				170					175					180
Tyr	Glu	Gly	Pro	Tyr	Tyr	Ile	Cys	Lys	Asp	Val	Ala	Ala	Glu	Glu
				185					190					195
Glu	Cys	Arg	Tyr	Ser	Gly	His	Cys	Thr	Phe	Ala	Tyr	Cys	Gln	Glu
				200					205					210
Glu	Ile	Asp	Val	Trp	Thr	Leu	Glu	Arg	Lys	Gly	Ala	Phe	Ser	Arg
				215					220					225
Glu	Ala	Phe	Phe	Gly	Gly	Asn	Gly	Lys	Ile	Asn	Leu	Thr	Val	Phe
				230					235					240
Lys	Leu	Leu	Gln	Glu	His	Leu	Gly	Glu	Phe	Ile	Phe	Leu	Cys	Glu
				245					250					255
Lys	Cys	Phe	Asp	His	Lys	Pro	Arg	Met	Ile	Ser	Lys	Arg	Asn	Lys
				260					265					270
Asp	Asn	Ser	Thr	Ala	Cys	Ser	His	Pro	Val	Thr	Lys	His	Glu	Phe
				275					280					285
Glu	Asp	Asn	Lys	Cys	Leu	Val	His	Ile	Leu	Arg	Glu	Thr	Thr	Val
				290					295					300
Lys	Tyr	Ser	Lys	Ile	Arg	Ser	Phe	His	Gly	Gln	Cys	Gln	Leu	Asp
				305					310					315
Leu	Cys	Arg	His	Glu	Val	Arg	Tyr	Gly	Cys	Leu	Arg	Glu	Asp	Glu
				320					325					330
Cys	Phe	Tyr	Ala	His	Ser	Leu	Val	Glu	Leu	Lys	Val	Trp	Ile	Met
				335					340					345
Gln	Asn	Glu	Thr	Gly	Ile	Ser	His	Asp	Ala	Ile	Ala	Gln	Glu	Ser
				350					355					360
Lys	Arg	Tyr	Trp	Gln	Asn	Leu	Glu	Ala	Asn	Val	Pro	Gly	Ala	Gln
				365					370					375
Val	Leu	Gly	Asn	Gln	Ile	Met	Pro	Gly	Phe	Leu	Asn	Met	Lys	Ile

	380		385		390
Lys Phe Val Cys	Ala Gln Cys Leu Arg	Asn Gly Gln Val Ile	Glu		
	395		400		405
Pro Asp Lys Asn	Arg Lys Tyr Cys Ser	Ala Lys Ala Arg His	Ser		
	410		415		420
Trp Thr Lys Asp	Arg Arg Ala Met Arg	Val Met Ser Ile Glu	Arg		
	425		430		435
Lys Lys Trp Met	Asn Ile Arg Pro Leu	Pro Thr Lys Lys Gln	Met		
	440		445		450
Pro Leu Gln Phe	Asp Leu Cys Asn His	Ile Ala Ser Gly Lys	Lys		
	455		460		465
Cys Gln Tyr Val	Gly Asn Cys Ser Phe	Ala His Ser Pro Glu	Glu		
	470		475		480
Arg Glu Val Trp	Thr Tyr Met Lys Glu	Asn Gly Ile Gln Asp	Met		
	485		490		495
Glu Gln Phe Tyr	Glu Leu Trp Leu Lys	Ser Gln Lys Asn Glu	Lys		
	500		505		510
Ser Glu Asp Ile	Ala Ser Gln Ser Asn	Lys Glu Asn Gly Lys	Gln		
	515		520		525
Ile His Met Pro	Thr Asp Tyr Ala Glu	Val Thr Val Asp Phe	His		
	530		535		540
Cys Trp Met Cys	Gly Lys Asn Cys Asn	Ser Glu Lys Gln Trp	Gln		
	545		550		555
Gly His Ile Ser	Ser Glu Lys His Lys	Glu Lys Val Phe His	Thr		
	560		565		570
Glu Asp Asp Gln	Tyr Cys Trp Gln His	Arg Phe Pro Thr Gly	Tyr		
	575		580		585
Phe Ser Ile Cys	Asp Arg Tyr Met Asn	Gly Thr Cys Pro Glu	Gly		
	590		595		600
Asn Ser Cys Lys	Phe Ala His Gly Asn	Ala Glu Leu His Glu	Trp		
	605		610		615
Glu Glu Arg Arg	Asp Ala Leu Lys Met	Lys Leu Asn Lys Ala	Arg		
	620		625		630
Lys Asp His Leu	Ile Gly Pro Asn Asp	Asn Asp Phe Gly Lys	Tyr		
	635		640		645
Ser Phe Leu Phe	Lys Asp Leu Asn				
	650				

<210> 51
 <211> 112
 <212> PRT
 <213> Homo sapiens

<220>
 <221> misc_feature
 <223> Incyte ID No.: 1968009

<400> 51

Met Gln Asp Thr Gly	Ser Val Val Pro Leu	His Trp Phe Gly Phe	
1	5	10	15
Gly Tyr Ala Ala Leu	Val Ala Ser Gly Gly	Ile Ile Gly Tyr Val	
	20	25	30
Lys Ala Gly Ser Val	Pro Ser Leu Ala Ala	Gly Leu Leu Phe Gly	
	35	40	45
Ser Leu Ala Gly Leu	Gly Ala Tyr Gln Leu	Ser Gln Asp Pro Arg	
	50	55	60
Asn Val Trp Val Phe	Leu Ala Thr Ser Gly	Thr Leu Ala Gly Ile	
	65	70	75
Met Gly Met Arg Phe	Tyr His Ser Gly Lys	Phe Met Pro Ala Gly	
	80	85	90
Leu Ile Ala Gly Ala	Ser Leu Leu Met Val	Ala Lys Val Gly Val	
	95	100	105
Ser Met Phe Asn Arg	Pro His		
	110		

<210> 52
 <211> 114
 <212> PRT
 <213> Homo sapiens
 <220>
 <221> misc_feature
 <223> Incyte ID No.: 1923127

<400> 52

Met	Glu	Lys	Pro	Leu	Phe	Pro	Leu	Val	Pro	Leu	His	Trp	Phe	Gly	
1				5					10					15	
Phe	Gly	Tyr	Thr	Ala	Leu	Val	Val	Ser	Gly	Gly	Ile	Val	Gly	Tyr	
				20					25					30	
Val	Lys	Thr	Gly	Ser	Val	Pro	Ser	Leu	Ala	Ala	Gly	Leu	Leu	Phe	
				35					40					45	
Gly	Ser	Leu	Ala	Gly	Leu	Gly	Ala	Tyr	Gln	Leu	Tyr	Gln	Asp	Pro	
				50					55					60	
Arg	Asn	Val	Trp	Gly	Phe	Leu	Ala	Ala	Thr	Ser	Val	Thr	Phe	Val	
				65					70					75	
Gly	Val	Met	Gly	Met	Arg	Ser	Tyr	Tyr	Tyr	Gly	Lys	Phe	Met	Pro	
				80					85					90	
Val	Gly	Leu	Ile	Ala	Gly	Ala	Ser	Leu	Leu	Met	Ala	Ala	Lys	Val	
				95					100					105	
Gly	Val	Arg	Met	Leu	Met	Thr	Ser	Asp							
				110											

<210> 53
 <211> 114
 <212> PRT
 <213> Homo sapiens
 <220>
 <221> misc_feature
 <223> Incyte ID No.: 3123954

<400> 53

Met	Ala	Ala	Ile	Pro	Ser	Ser	Gly	Ser	Leu	Val	Ala	Thr	His	Asp	
1				5					10					15	
Tyr	Tyr	Arg	Arg	Arg	Leu	Gly	Ser	Thr	Ser	Ser	Asn	Ser	Ser	Cys	
				20					25					30	
Ser	Ser	Thr	Glu	Cys	Pro	Gly	Glu	Ala	Ile	Pro	His	Pro	Pro	Gly	
				35					40					45	
Leu	Pro	Lys	Ala	Asp	Pro	Gly	His	Trp	Trp	Ala	Ser	Phe	Phe	Phe	
				50					55					60	
Gly	Lys	Ser	Thr	Leu	Pro	Phe	Met	Ala	Thr	Val	Leu	Glu	Ser	Ala	
				65					70					75	
Glu	His	Ser	Glu	Pro	Pro	Gln	Ala	Ser	Ser	Ser	Met	Thr	Ala	Cys	
				80					85					90	
Gly	Leu	Ala	Arg	Asp	Ala	Pro	Arg	Lys	Gln	Pro	Gly	Gly	Gln	Ser	
				95					100					105	
Ser	Thr	Ala	Ser	Ala	Gly	Pro	Pro	Ser							
				110											

<210> 54
 <211> 291
 <212> PRT
 <213> Homo sapiens

<220>
 <221>
 <223>

<400> 54

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Met Ser Gln Glu Gly Val Glu Leu Glu Lys Ser Val Arg Gly Leu
 1      5      10      15
Arg Glu Lys Phe His Gly Lys Val Ser Ser Lys Lys Ala Gly Ala
 20      25      30
Leu Met Arg Lys Phe Gly Ser Asp His Thr Gly Val Gly Arg Ser
 35      40      45
Ile Val Tyr Gly Val Lys Gln Lys Asp Gly Gln Glu Leu Ser Asn
 50      55      60
Asp Leu Asp Ala Gln Asp Pro Pro Glu Asp Met Lys Gln Asp Arg
 65      70      75
Asp Ile Gln Ala Val Ala Thr Ser Leu Leu Pro Leu Thr Glu Ala
 80      85      90
Asn Leu Arg Met Phe Gln Arg Ala Gln Asp Asp Leu Ile Pro Ala
 95      100     105
Val Asp Arg Gln Phe Ala Cys Ser Ser Cys Asp His Val Trp Trp
110     115     120
Arg Arg Val Pro Gln Arg Lys Glu Val Ser Arg Cys Arg Lys Cys
125     130     135
Arg Lys Arg Tyr Glu Pro Val Pro Ala Asp Lys Met Trp Gly Leu
140     145     150
Ala Glu Phe His Cys Pro Lys Cys Arg His Asn Phe Arg Gly Trp
155     160     165
Ala Gln Met Gly Ser Pro Ser Pro Cys Tyr Gly Cys Gly Phe Pro
170     175     180
Val Tyr Pro Thr Arg Ile Leu Pro Pro Arg Trp Asp Arg Asp Pro
185     190     195
Asp Arg Arg Ser Thr His Thr His Ser Cys Ser Ala Ala Asp Cys
200     205     210
Tyr Asn Arg Arg Glu Pro His Val Pro Gly Thr Ser Cys Ala His
215     220     225
Pro Lys Ser Arg Lys Gln Asn His Leu Pro Lys Val Leu His Pro
230     235     240
Ser Asn Pro His Ile Ser Ser Gly Ser Thr Val Ala Thr Cys Leu
245     250     255
Ser Gln Gly Gly Leu Leu Glu Asp Leu Asp Asn Leu Ile Leu Glu
260     265     270
Asp Leu Lys Glu Glu Glu Glu Glu Glu Glu Val Glu Asp Glu
275     280     285
Glu Gly Gly Pro Arg Glu
290

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<210> 55

<211> 610

<212> PRT

<213> Homo sapiens

<220>

<221> misc_feature

<223> Incyte ID No.: 1867333

<400> 55

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Met Trp Leu Pro Leu Val Leu Leu Leu Ala Val Leu Leu Leu Ala
 1      5      10      15
Val Leu Cys Lys Val Tyr Leu Gly Leu Phe Ser Gly Ser Ser Pro
 20      25      30
Asn Pro Phe Ser Glu Asp Val Lys Arg Pro Pro Ala Pro Leu Val
 35      40      45
Thr Asp Lys Glu Ala Arg Lys Lys Val Leu Lys Gln Ala Phe Ser
 50      55      60
Ala Asn Gln Val Pro Glu Lys Leu Asp Val Val Val Ile Gly Ser
 65      70      75
Gly Phe Gly Gly Leu Ala Ala Ala Ala Ile Leu Ala Lys Ala Gly
 80      85      90
Lys Arg Val Leu Val Leu Glu Gln His Thr Lys Ala Gly Gly Cys

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	95		100		105
Cys His Thr Phe	Gly Lys Asn Gly Leu	Glu Phe Asp Thr Gly	Ile		
	110		115		120
His Tyr Ile Gly	Arg Met Glu Glu Gly	Ser Ile Gly Arg Phe	Ile		
	125		130		135
Leu Asp Gln Ile	Thr Glu Gly Gln Leu	Asp Trp Ala Pro Leu	Ser		
	140		145		150
Ser Pro Phe Asp	Ile Met Val Leu Glu	Gly Pro Asn Gly Arg	Lys		
	155		160		165
Glu Tyr Pro Met	Tyr Ser Gly Glu Lys	Ala Tyr Ile Gln Gly	Leu		
	170		175		180
Lys Glu Lys Phe	Pro Gln Glu Glu Ala	Ile Ile Asp Lys Tyr	Ile		
	185		190		195
Lys Leu Val Lys	Val Val Ser Ser Gly	Ala Pro His Ala Ile	Leu		
	200		205		210
Leu Lys Phe Leu	Pro Leu Pro Val Val	Gln Leu Leu Asp Arg	Cys		
	215		220		225
Gly Leu Leu Thr	Arg Phe Ser Pro Phe	Leu Gln Ala Ser Thr	Gln		
	230		235		240
Ser Leu Ala Glu	Val Leu Gln Gln Leu	Gly Ala Ser Ser Glu	Leu		
	245		250		255
Gln Ala Val Leu	Ser Tyr Ile Phe Pro	Thr Tyr Gly Val Thr	Pro		
	260		265		270
Asn His Ser Ala	Phe Ser Met His Ala	Leu Leu Val Asn His	Tyr		
	275		280		285
Met Lys Gly Gly	Phe Tyr Pro Arg Gly	Gly Ser Ser Glu Ile	Ala		
	290		295		300
Phe His Thr Ile	Pro Val Ile Gln Arg	Ala Gly Gly Ala Val	Leu		
	305		310		315
Thr Lys Ala Thr	Val Gln Ser Val Leu	Leu Asp Ser Ala Gly	Lys		
	320		325		330
Ala Cys Gly Val	Ser Val Lys Lys Gly	His Glu Leu Val Asn	Ile		
	335		340		345
Tyr Cys Pro Ile	Val Val Ser Asn Ala	Gly Leu Phe Asn Thr	Tyr		
	350		355		360
Glu His Leu Leu	Pro Gly Asn Ala Arg	Cys Leu Pro Gly Val	Lys		
	365		370		375
Gln Gln Leu Gly	Thr Val Arg Pro Gly	Leu Gly Met Thr Ser	Val		
	380		385		390
Phe Ile Cys Leu	Arg Gly Thr Lys Glu	Asp Leu His Leu Pro	Ser		
	395		400		405
Thr Asn Tyr Tyr	Val Tyr Tyr Asp Thr	Asp Met Asp Gln Ala	Met		
	410		415		420
Glu Arg Tyr Val	Ser Met Pro Arg Glu	Glu Ala Ala Glu His	Ile		
	425		430		435
Pro Leu Leu Phe	Phe Ala Phe Pro Ser	Ala Lys Asp Pro Thr	Trp		
	440		445		450
Glu Asp Arg Phe	Pro Gly Arg Ser Thr	Met Ile Met Leu Ile	Pro		
	455		460		465
Thr Ala Tyr Glu	Trp Phe Glu Glu Trp	Gln Ala Glu Leu Lys	Gly		
	470		475		480
Lys Arg Gly Ser	Asp Tyr Glu Thr Phe	Lys Asn Ser Phe Val	Glu		
	485		490		495
Ala Ser Met Ser	Val Val Leu Lys Leu	Phe Pro Gln Leu Glu	Gly		
	500		505		510
Lys Val Glu Ser	Val Thr Ala Gly Ser	Pro Leu Thr Asn Gln	Phe		
	515		520		525
Tyr Leu Ala Ala	Pro Arg Gly Ala Cys	Tyr Gly Ala Asp His	Asp		
	530		535		540
Leu Gly Arg Leu	His Pro Cys Val Met	Ala Ser Leu Arg Ala	Gln		
	545		550		555
Ser Pro Ile Pro	Asn Leu Tyr Leu Thr	Gly Gln Asp Ile Phe	Thr		
	560		565		570
Cys Gly Leu Val	Gly Ala Leu Gln Gly	Ala Leu Leu Cys Ser	Ser		
	575		580		585
Ala Ile Leu Lys	Arg Asn Leu Tyr Ser	Asp Leu Lys Asn Leu	Asp		
	590		595		600
Ser Arg Ile Arg	Ala Gln Lys Lys Lys	Asn			

605

610

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<210> 56
<211> 352
<212> PRT
<213> Homo sapiens

<220>
<221> misc_feature
<223> Incyte ID No.: 1461451
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<400> 56

Pro	Arg	Val	Arg	Gly	Arg	Trp	Val	Ala	His	Ala	Ser	Ala	His	Ala
1				5					10					15
Ser	Ala	His	Ala	Ser	Asp	Glu	Ile	Pro	Ala	Ser	Gly	Trp	Thr	Gln
				20					25					30
Trp	Phe	Cys	Thr	Glu	Ala	Leu	Val	Met	Val	Ala	Pro	Val	Trp	Tyr
				35					40					45
Leu	Val	Ala	Ala	Ala	Leu	Leu	Val	Gly	Phe	Ile	Leu	Phe	Leu	Thr
				50					55					60
Arg	Ser	Arg	Gly	Arg	Ala	Ala	Ser	Ala	Gly	Gln	Glu	Pro	Leu	His
				65					70					75
Asn	Glu	Glu	Leu	Ala	Gly	Ala	Gly	Arg	Val	Ala	Gln	Pro	Gly	Pro
				80					85					90
Leu	Glu	Pro	Glu	Glu	Pro	Arg	Ala	Gly	Gly	Arg	Pro	Arg	Arg	Arg
				95					100					105
Arg	Asp	Leu	Gly	Ser	Arg	Leu	Gln	Ala	Gln	Arg	Arg	Ala	Gln	Arg
				110					115					120
Val	Ala	Trp	Ala	Glu	Ala	Asp	Glu	Asn	Glu	Glu	Glu	Ala	Val	Ile
				125					130					135
Leu	Ala	Gln	Glu	Glu	Glu	Gly	Val	Glu	Lys	Pro	Ala	Glu	Thr	His
				140					145					150
Leu	Ser	Gly	Lys	Ile	Gly	Ala	Lys	Lys	Leu	Arg	Lys	Leu	Glu	Glu
				155					160					165
Lys	Gln	Ala	Arg	Lys	Ala	Gln	Arg	Glu	Ala	Glu	Glu	Ala	Glu	Arg
				170					175					180
Glu	Glu	Arg	Lys	Arg	Leu	Glu	Ser	Gln	Arg	Glu	Ala	Glu	Trp	Lys
				185					190					195
Lys	Glu	Glu	Glu	Arg	Leu	Arg	Leu	Glu	Glu	Glu	Gln	Lys	Glu	Glu
				200					205					210
Glu	Glu	Arg	Lys	Ala	Arg	Glu	Glu	Gln	Ala	Gln	Arg	Glu	His	Glu
				215					220					225
Glu	Tyr	Leu	Lys	Leu	Lys	Glu	Ala	Phe	Val	Val	Glu	Glu	Glu	Gly
				230					235					240
Val	Gly	Glu	Thr	Met	Thr	Glu	Glu	Gln	Ser	Gln	Ser	Phe	Leu	Thr
				245					250					255
Glu	Phe	Ile	Asn	Tyr	Ile	Lys	Gln	Ser	Lys	Val	Val	Leu	Leu	Glu
				260					265					270
Asp	Leu	Ala	Ser	Gln	Val	Gly	Leu	Arg	Thr	Gln	Asp	Thr	Ile	Asn
				275					280					285
Arg	Ile	Gln	Asp	Leu	Leu	Ala	Glu	Gly	Thr	Ile	Thr	Gly	Val	Ile
				290					295					300
Asp	Asp	Arg	Gly	Lys	Phe	Ile	Tyr	Ile	Thr	Pro	Glu	Glu	Leu	Ala
				305					310					315
Ala	Val	Ala	Asn	Phe	Ile	Arg	Gln	Arg	Gly	Arg	Val	Ser	Ile	Ala
				320					325					330
Glu	Leu	Ala	Gln	Ala	Ser	Asn	Ser	Leu	Ile	Ala	Trp	Gly	Arg	Glu
				335					340					345
Ser	Pro	Ala	Gln	Ala	Pro	Ala								
				350										

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<210> 57
<211> 216
<212> PRT
<213> Homo sapiens
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<220>
 <221> misc_feature
 <223> Incyte ID No.: 2345712

<400> 57

Tyr	Asp	Pro	Ile	Gly	Phe	Gly	Leu	Ser	Trp	Glu	Ala	Gly	Arg	Ile	1	5	10	15
Ile	Gly	Trp	Gly	Lys	Pro	Thr	Arg	Gly	Arg	Gly	Arg	Gly	Gly	Ser	20	25	30	35
Leu	Ser	Thr	Arg	Gly	Arg	Gly	Ser	Glu	Val	Pro	Asp	Ser	Ala	His	40	45	50	55
Leu	Ala	Pro	Thr	Pro	Leu	Phe	Ser	Glu	Ser	Gly	Cys	Cys	Gly	Leu	60	65	70	75
Arg	Ser	Arg	Phe	Leu	Thr	Asp	Cys	Lys	Met	Glu	Glu	Gly	Gly	Asn	80	85	90	95
Leu	Gly	Gly	Leu	Ile	Lys	Met	Val	His	Leu	Leu	Val	Leu	Ser	Gly	100	105	110	115
Ala	Trp	Gly	Met	Gln	Met	Trp	Val	Thr	Phe	Val	Ser	Gly	Phe	Leu	120	125	130	135
Leu	Phe	Arg	Ser	Leu	Pro	Arg	His	Thr	Phe	Gly	Leu	Val	Gln	Ser	140	145	150	155
Lys	Leu	Phe	Pro	Phe	Tyr	Phe	His	Ile	Ser	Met	Gly	Cys	Ala	Phe	160	165	170	175
Ile	Asn	Leu	Cys	Ile	Leu	Ala	Ser	Gln	His	Ala	Trp	Ala	Gln	Leu	180	185	190	195
Thr	Phe	Trp	Glu	Ala	Ser	Gln	Leu	Tyr	Leu	Leu	Phe	Leu	Ser	Leu	200	205	210	215
Thr	Leu	Ala	Thr	Val	Asn	Ala	Arg	Trp	Leu	Glu	Pro	Arg	Thr	Thr				
Ala	Ala	Met	Trp	Ala	Leu	Gln	Thr	Val	Glu	Lys	Glu	Arg	Gly	Leu				
Gly	Gly	Glu	Val	Pro	Gly	Ser	His	Gln	Gly	Ser	Asp	Pro	Tyr	Arg				
Gln	Leu	Arg	Glu	Lys	Asp													

<210> 58
 <211> 292
 <212> PRT
 <213> Homo sapiens

<220>
 <221> misc_feature
 <223> Incyte ID No.: 1810320

<400> 58

Met	Ala	Gln	Pro	Pro	Pro	Asp	Val	Glu	Gly	Asp	Asp	Cys	Leu	Pro	1	5	10	15
Ala	Tyr	Arg	His	Leu	Phe	Cys	Pro	Asp	Leu	Leu	Arg	Asp	Lys	Val	20	25	30	35
Ala	Phe	Ile	Thr	Gly	Gly	Gly	Ser	Gly	Ile	Gly	Phe	Arg	Ile	Ala	40	45	50	55
Glu	Ile	Phe	Met	Arg	His	Gly	Cys	His	Thr	Val	Ile	Ala	Ser	Arg	60	65	70	75
Ser	Leu	Pro	Arg	Val	Leu	Thr	Ala	Ala	Arg	Lys	Leu	Ala	Gly	Ala	80	85	90	95
Thr	Gly	Arg	Arg	Cys	Leu	Pro	Leu	Ser	Met	Asp	Val	Arg	Ala	Pro	100	105	110	115
Pro	Ala	Val	Met	Ala	Ala	Val	Asp	Gln	Ala	Leu	Lys	Glu	Phe	Gly	120	125	130	135
Arg	Ile	Asp	Ile	Leu	Ile	Asn	Cys	Ala	Ala	Gly	Asn	Phe	Leu	Cys				
Pro	Ala	Gly	Ala	Leu	Ser	Phe	Asn	Ala	Phe	Lys	Thr	Val	Met	Asp				
Ile	Asp	Thr	Ser	Gly	Thr	Phe	Asn	Val	Ser	Arg	Val	Leu	Tyr	Glu				

	140		145		150
Lys Phe Phe Arg	Asp His Gly Gly Val	Ile Val Asn Ile Thr	Ala		
	155		160		165
Thr Leu Gly Asn	Arg Gly Gln Ala Leu	Gln Val His Ala Gly	Ser		
	170		175		180
Ala Lys Ala Ala	Val Asp Ala Met Thr	Arg His Leu Ala Val	Glu		
	185		190		195
Trp Gly Pro Gln	Asn Ile Arg Val Asn	Ser Leu Ala Pro Gly	Pro		
	200		205		210
Ile Ser Gly Thr	Glu Gly Leu Arg Arg	Leu Gly Gly Pro Gln	Ala		
	215		220		225
Ser Leu Ser Thr	Lys Val Thr Ala Ser	Pro Leu Gln Arg Leu	Gly		
	230		235		240
Asn Lys Thr Glu	Ile Ala His Ser Val	Leu Tyr Leu Ala Ser	Pro		
	245		250		255
Leu Ala Ser Tyr	Val Thr Gly Ala Val	Leu Val Ala Asp Gly	Gly		
	260		265		270
Ala Trp Leu Thr	Phe Pro Asn Gly Val	Lys Gly Leu Pro Asp	Phe		
	275		280		285
Ala Ser Phe Ser	Ala Lys Leu				
	290				

<210> 59
 <211> 158
 <212> PRT
 <213> Homo sapiens

<220>
 <221> misc_feature
 <223> Incyte ID No.: 964996

<400> 59

Glu Gly Gly Pro	Ser Trp Thr Arg	Glu Arg Thr Leu Val Ala Val
1	5	10 15
Lys Pro Asp Gly	Val Gln Arg Arg	Leu Val Gly Asp Val Ile Gln
	20	25 30
Arg Phe Glu Arg	Arg Gly Phe Thr	Leu Val Gly Met Lys Met Leu
	35	40 45
Gln Ala Pro Glu	Ser Val Leu Ala	Glu His Tyr Gln Asp Leu Arg
	50	55 60
Arg Lys Pro Phe	Tyr Pro Ala Leu	Ile Arg Tyr Met Ser Ser Gly
	65	70 75
Pro Val Val Ala	Met Val Trp Glu	Gly Tyr Asn Val Val Arg Ala
	80	85 90
Ser Arg Ala Met	Ile Gly His Thr	Asp Ser Ala Glu Ala Ala Pro
	95	100 105
Gly Thr Ile Arg	Gly Tyr Phe Ser	Val His Ile Ser Arg Asn Val
	110	115 120
Ile His Ala Ser	Asp Ser Val Glu	Gly Ala Gln Arg Glu Ile Gln
	125	130 135
Leu Trp Phe Gln	Ser Ser Glu Leu	Val Ser Trp Ala Asp Gly Gly
	140	145 150
Gln His Ser Ser	Ile His Pro Ala	
	155	

<210> 60
 <211> 559
 <212> DNA
 <213> Rattus norvegicus

<220>
 <221> misc_feature
 <223> Incyte ID No.: 701884305H1

<400> 60

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ggaacaccta acgcgcgtgc gcttcttcca cgccacggaa accgtgcagg cctgggtgtgg 60
tctccaaagt gactgaacaa tgcagaagga cagtggccca ctggttcctt tacattatta 120
tggtttcggc tatgcggccc tgggtggctac tgggtgggatt attggctatg caaaagcagg 180
tagtgtgccg tccctggctg ctggactctt ctttgggggc ctggcaggcc tgggtgccta 240
ccagctgtct caggacccca ggaacgtgtg ggttttccta gctacgtctg ggactttggc 300
tggcattatg gggatgagat tctacaactc tgggaaattt atgcctgcag gtttgatcgc 360
gggagccagt ttgctgatgg ttgccaactc tggacttagt atgttgagtt caccatcc 420
gtagtagcca tagtctgcg tgggtcatg atgagttgac actctccagt cctccacatt 480
accacgtga agagataaga acagcaaaga cctacactga gcacatggag gcgaagacgt 540
ggttactata gtgaccgtc                                     559

```

<210> 61

<211> 326

<212> DNA

<213> Rattus norvegicus

<220>

<221> misc_feature

<223> Incyte ID No.: 701607951H1

<400> 61

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gtgttgggtg tgttcttact ttgcggattt taccaccctg gaattgttcc gtacgcgcag 60
gcgcgcgggc gctctccgtg gcaactctctg ctgagctagc ggactgcccg cctctctaaa 120
acgtcctgta actgcggttc cgggagtggg aacctaaacg cgctgctgct tcttccacgc 180
cagggaacc gtgcaggcct ggtgtggtct ccaaagtac tgaacaatgc agaaggacag 240
tggccactg gttcctttac attattatg tttcggctat gcggccctgg tggctactgg 300
tgggattatt ggctatgcaa aagcag                                     326

```

<210> 62

<211> 333

<212> DNA

<213> Rattus norvegicus

<220>

<221> misc_feature

<223> Incyte ID No.: 701644253H1

<400> 62

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aacgtcctgt aactgcggtt ccgggagtgg aaacctaaac gcgcgtgcgc tttcttccac 60
gccacggaaa accgtgcagg cctngtgtgg tctocanagt gactgaacaa tgcagaagga 120
cagtggccca ctgntcctt tacattatta tggtttcggc tatgcggccc tgggtggctac 180
tgggtgggatt attggctatg caaaagcagg tagtgtgccg tccctggctg ctggactctt 240
ctttgggggc ctggcaggcc tgggtgccta ccagctgtct caggacccca ggaacgtgtg 300
ggttttccta gctacgncgt ggactttggc tgg                                     333

```

<210> 63

<211> 318

<212> DNA

<213> Rattus norvegicus

<220>

<221> misc_feature

<223> Incyte ID No.: 701513151H1

<400> 63

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cttactttgc ggattttacc accctggaat tgttccgtac gcgcangnc gcggggctct 60
cccgctgact ctctgctgag ctacgggact gcccgctct ctaaaacgtc ctgtaactgc 120
ggttccggga gtggaaacct aaacgcgcgt gcgcttcttc cagccacgg aaaccgtgca 180
ggcctgggtg ggtctccaaa gtgactgaac aatgcagaag gacagtggcc cactggttcc 240
tttacattat tatggtttcg gctatgcggc cctgggtggc actggtggga ttattggcta 300

```

tgcaaaagca ggtagtgt

318

<210> 64

<211> 315

<212> DNA

<213> Rattus norvegicus

<220>

<221> misc_feature

<223> Incyte ID No.: 701652337H1

<400> 64

```
cagcncaggc ctccgggctc cagctccggt gttgggtnca ggcttggtgt ggtctccaaa 60
gtgactgaac aatgcagaag gacagtggcc cactggttcc tttacattat tatggtttcg 120
gctatgcggc cctggtggct actggtggga ttattggcta tgcaaaagca ggtagtgtgc 180
cgctccctggc tgctggactc ttctttgggg gcctggcagg cctgggtgcc taccagctgt 240
ctcaggacct caggaacgtg tgggttttcc tagctacgtc tgggactttg gctggcatat 300
ggggatgaga ttcta 315
```

<210> 65

<211> 313

<212> DNA

<213> Rattus norvegicus

<220>

<221> misc_feature

<223> Incyte ID No.: 701562183H1

<400> 65

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ggtctccaaa gtgactgaac aatgcagaag gacagtggcc cactggttcc tttacattat 60
tatggtttcg gctatgcggc cctggtggct actggtggga ttattggcta tgcaaaagca 120
ggtagtgtgc cgctccctggc tgctggactc ttctttgggg gcctggcagg cctgggtgcc 180
taccagctgt ctcaggacct caggaacgtg tgggttttcc tagctacgtc tgggactttg 240
gctggcatta tggggatgag attctacaac tctgggaaat ttatgcctgc aggtttgatc 300
gcgggancat ttt 313
```

<210> 66

<211> 304

<212> DNA

<213> Rattus norvegicus

<220> misc_feature

<223> Incyte ID No.: 700227356H1

<400> 66

```
cgccgtcgtc ctccagcgca ggctccggg ctccagctcc ggtgttggtt gcaggcctgg 60
tgtggtctcc aaagtgactg aacaatgcag aaggacagtg gccactggt tcctttacat 120
tattatggtt tcggctatgc ggccctggtg gctactggtg ggattattgg ctatgcaaaa 180
gcaggtagtg tgccgtccct ggctgctgga ctcttctttg ggggcctggc aggctgggt 240
gcctaccagc tgtctcagga cccaggaac gtgtgggttt tcctagctac gtctgggact 300
ttgg 304
```

<210> 67

<211> 327

<212> DNA

<213> Rattus norvegicus

<220>

<221> misc_feature

<223> Incyte ID No.: 701649802H1

<400> 67

```

ctccgggtgtt ggggtgcaggc ctggtgtggt ctccaaagtg actgaacaat gcagaaggac 60
agtggaccac tggttcctta cattattatg gtttcggcta tgcggccctg gtggctactg 120
gtgggattat tgnctttgca aaagcaggta gtgtgccgtc cctggctgtt ggactcttct 180
ttggggggcct ggcaggcctg ggtgccctacc agctgtctca ggaccccagg aacgtgtggg 240
ttttcctagc tacgtctggg actttggctg gcattatggg gatgagattc tacaactctg 300
ggaaatttat gcctgcagtt tgatcgc 327

```

<210> 68

<211> 305

<212> DNA

<213> Rattus norvegicus

<220>

<221> misc_feature

<223> Incyte ID No.: 700226414H1

<400> 68

```

gccgtcgtcc tccagcncag gcctccgggc tccagctccg gtgttgggtg caggcctggt 60
gtggtctcca aagtgactga acaatgcaga aggacagtgg cccactgggt cctttacatt 120
attatggttt cggctatgcg gccctgggtg ctactgggtg gattattggc tatgcaaaag 180
caggtagtgt gccgtccctg gctgctggac tcttcttttg gggcctggca ggccctgggtg 240
cctaccagct gtctcaggac cccaggaagt gtgggttttc ctactacgt ctgggacttg 300
gctgg 305

```

<210> 69

<211> 295

<212> DNA

<213> Rattus norvegicus

<220>

<221> misc_feature

<223> Incyte ID No.: 700275094H1

<400> 69

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tctccagcn caggentccg ggctccagct ccggtgttgg gtgcaggcct ggtgtggtct 60
ccaaagtga tgaacaatgc agaaggacag tggcccactg gttcctttac attattatgg 120
tttcggctat ggggccctgg tggctactgg tgggattatt ggctatgcaa aagcaggtag 180
tgtgccgtcc ctggctgctg gactcttctt tggggggcct ggcaggcctg ggtgcctacc 240
agctgtctca ggaccccagg aacgtgtggg ttttcctagc tacgtctggg atttg 295

```

<210> 70

<211> 301

<212> DNA

<213> Rattus norvegicus

<220>

<221> misc_feature

<223> Incyte ID No.: 700226425H1

<400> 70

```

cctgacctct gttcctgtgc tccgcccgtc gtcctccagc gcaggcctcc gggctccagc 60
tccggtgttg ggtgcaggcc tgggtgtggt tccaaagtga ctgaacaatg cagaaggaca 120
gtggcccact ggttccttta cattattatg gtttcggcta tgcggccctg gtggctactg 180
gtgggattat tggctatgca aaagcaggta gtgtgccgtc cctggctgct ggactcttct 240
ttgggggcct ggcaggcctg ggtgcctacc agctgtctca ggaccccagg aacgtgtggg 300
t 301

```

<210> 71

<211> 282

<212> DNA
<213> Rattus norvegicus

<220>
<221> misc_feature
<223> Incyte ID No.: 700275207H1

<400> 71

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tcctccagcg caggectccg ggctccagct cgggtgttgg gtgcaggcct ggtgtggtct 60
ccaaagtgc tgaacaatgc agaaggacag tggccactg gttcctttac attattatgg 120
tttcggctat gcggccctgg tggctactgg tgggattatt ggctatgcaa aagcaggtag 180
tgtgccgtcc ctggctctg gactcttctt tgggggctg gcaggcctgg gtgcctacca 240
gctgtctcag gacccaggga acgtgtgggt tttcctagct ac 282
```

<210> 72
<211> 282
<212> DNA
<213> Rattus norvegicus

<220>
<221> misc_feature
<223> Incyte ID No.: 701507568H1

<400> 72

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cgccgtcgct ctccagcgca ggcctccggg ctccagctcc ggtgttgggt gcaggcctgg 60
tgtggtctcc aaagtgactg aacaatgcag aaggacagtg gccactgggt tcctttacat 120
tattatgggt tcggctatgc ggccctgggt gctactgggt ggattattgg ctatgcaaaa 180
gcaggtagtg tgccgtccct ggctgctgga ctcttctttg ggggcctggc aggctgggt 240
gcctaccagc tgtctcagga cccagggaac gtgtgggttt tc 282
```

<210> 73
<211> 281
<212> DNA
<213> Rattus norvegicus

<220>
<221> misc_feature
<223> Incyte ID No.: 700300118H1

<400> 73

```
cgccgtcgct ctccagcgca ggcctccggg ctccagctcc ggtgttgggt gcaggcctgg 60
tgtggtctcc aaagtgactg aacaatgcag aaggacagtg gccactgggt tcctttacat 120
tattatgggt tcggctatgc ggccctgggt gctactgggt ggattattgg ctatgcaaaa 180
gcaggtagtg tgccgtccct ggctgctgga ctcttctttg ggggcctggc aggctgggt 240
gcctaccagc tgtctcagga cccagggaac gtgtgggttt t 281
```

<210> 74
<211> 292
<212> DNA
<213> Rattus norvegicus

<220>
<221> misc_feature
<223> Incyte ID No.: 700301710H1

<400> 74

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cctgnacctc tgttctgtg ctcccgccgt cgtcctccag cgcaggcctc cgggctccag 60
ctccggtggt ggggtgcaggc ctggtgtggt ctccaaagtg actgaacaat gcagaaggac 120
agtggccac tggttccttt acattattat ggtttcggct atgcggccct ggtggctact 180
ggtgggatta ttggctatgc aaaagcagggt agtgtgccgt ccctggctgc tggactcttc 240
tttgggggcc tggcaggcct ggggtgcctac cagctgtctc aggaccccag ga 292
```

<210> 75
 <211> 289
 <212> DNA
 <213> Rattus norvegicus

<220>
 <221> misc_feature
 <223> Incyte ID No.: 700064344H1

<400> 75

```
cagcgcaggc ctccgggctc cagctccggt gttgggtgtg ttcttacttt gcggatttta 60
ccaccctgga attgttccgt acgcgcaggc gcgcgggcgc tctcccggtc actctctgct 120
gagctagcgg actgcccgcc tctctaaaac gtcctgtaac tgcggttccg ggagtggaaa 180
cctaaacgcg cgtgcgcttc ttccacgcca cggaaaccgt gcaggcctgg tgtggtctcc 240
aaagtgatga acatgcagaa ggacantggc ccactgggtc ttanatatt 289
```

<210> 76
 <211> 276
 <212> DNA
 <213> Rattus norvegicus

<220>
 <221> misc_feature
 <223> Incyte ID No.: 701423273H1

<400> 76

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agcgcaggcc tcagggtccc agctccggtg ttgggtgcag gcctgggtgn gtctccaaag 60
tgactgaaca atgcagaagg acagtggccc actgggttcc ttacattatt atgggttccg 120
ctatgcggcc ctggtggcta ctggtgggat tattggctat gcaaaagcag gtagtgtgcc 180
gtccctggct gctggactct tctttggggg cctggcaggc ctgggtgcct accagctgtc 240
tcaggacccc aggaacgtgt gggtttccct agctac 276
```

<210> 77
 <211> 293
 <212> DNA
 <213> Rattus norvegicus

<220>
 <221> misc_feature
 <223> Incyte ID No.: 700225847H1

<400> 77

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ccgtcgtcct ccagcncagg cctccgggct ccagctccgg tgttgggtgc aggccctggtg 60
tggtctccaa agtgactgaa caatgcagaa ggacagtggc ccactgggtc ctttacatta 120
ttatgggttc ggctatgcgg ccctgggtgg tactgggtgg attattggct atgcaaaagc 180
aggtagtgtg ccgtccctgg ctgctggact ctctttgggg gcctggcang cctgggtgcc 240
taccagctgt ctcaggaccc cagaacgtgt gggtttccta gctacgtctg gga 293
```

<210> 78
 <211> 274
 <212> DNA
 <213> Rattus norvegicus

<220>
 <221> misc_feature
 <223> Incyte ID No.: 701462776H1

<400> 78

```
tgtctccgcc gtcgtcctcc agcgcaggcc tccgggctcc agctccggtg ttgggtgcag 60
gcctgggtgtg gtctccaaag tgactgaaca atgcagaagg acagtggcnc actggttcct 120
```


ttacattatt atggttttcgg ctatgcggcc ctggtggcta ctggtgggat tattggctat 180
 gcaaaagcag gtagtgtgcc gtccctggct gctggactct tctttggggg cctggcaggc 240
 ctgggtgcct accagctgtc tcaggacccc agga 274

<210> 79
 <211> 282
 <212> DNA
 <213> Rattus norvegicus

<220>
 <221> misc_feature
 <223> Incyte ID No.: 700916803H1

<400> 79

gtgctcccg cgtcgtcctc cagcgcaggc ctccgggctc cagctnccgg tgttgggtgt 60
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 tctcccggtg actctctgct gagctagcgg actgcccgc tctctaaaac gtctgtaac 180
 tgcggttccg ggagtggaaa cctaaacgcg cgtgcgcttc ttccacgcca cggaaccgt 240
 gcaggcctgg tgtggtctcc aaagtgactg aacaatgcag aa 282

<210> 80
 <211> 280
 <212> DNA
 <213> Rattus norvegicus

<220>
 <221> misc_feature
 <223> Incyte ID No.: 700478141H1

<400> 80

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 ttatgggttc ggctatgcgg ccctggtggc tactggtggg attattggct atgcaaaagc 180
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 ctaccagctg tctcaggacc ccaggaaact gtgggttttc 280

<210> 81
 <211> 299
 <212> DNA
 <213> Rattus norvegicus

<220>
 <221> misc_feature
 <223> Incyte ID No.: 701646690H1

<400> 81

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 aacaatgcan aangacagtn gccactgggt tcctttacnt tattatgggt tcnmntatgc 120
 ngccctgggt gctactgggt ggattattgg ctatgcaaaa ncaggtagtg tgccgtccct 180
 ggctgntgga ntcttctttg ggggcctggc aggcctgggt gcctaccagc tgtctcagga 240
 ccccaggaac gtgtgggttt tcttagctac gtctggnact ttgctggca tatggggat 299

<210> 82
 <211> 286
 <212> DNA
 <213> Rattus norvegicus

<220>
 <221> misc_feature
 <223> Incyte ID No.: 701624261H1

<400> 82

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tctcctccac aggtgcangc ctggtgtggt ctccaaagtg actgmncaat gcagaaggac 60
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ggtgggatta ttggctatgc aaaagcagggt agtgtgccgt ccttggtgc nngactcttc 180
tttgggggcc tggcaggcct ggtgcctac cagctgtctc aggaccccag gaacgtgtgg 240
gttttcctag ctacgtctgg gactttggct ggcattatgg ggatga 286

```

<210> 83

<211> 266

<212> DNA

<213> *Rattus norvegicus*

<220>

<221> misc_feature

<223> Incyte ID No.: 700912920H1

<400> 83

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gcagaaggac agtggcccac tggttccttt acattattat ggtttcggct atgcggccct 60
ggtgggtact ggtgggatta ttggctatgc aaaagcagggt agtgtgccgt ccttggtgc 120
tggactcttc tttgggggcc tggcaggcct ggtgcctac cagctgtctc aggaccccag 180
gaacgtgtgg gttttcctag ctacgtctgg gactttggct ggcattatgg ggatgagatt 240
ctacaactct gggaaattta tgctg 266

```

<210> 84

<211> 262

<212> DNA

<213> *Rattus norvegicus*

<220>

<221> misc_feature

<223> Incyte ID No.: 701482566H1

<400> 84

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ctggctgctg gactcttctt tgggggcctg gcaggcctgg gtgcctacca gctgtctcag 60
gaccccagga acgtgtgggt tttcctagct acgtctggga ctttggtctg cattatgggg 120
atgagattct acaactctgg gaaatttatg cctgcagggt tgatcgcggg agccagtttg 180
ctgatgggtg ccaaacttgg acttagtatg ttgagttcac cccatccgta gtagccatag 240
cctgcgtggt gctcatgatg ag 262

```

<210> 85

<211> 285

<212> DNA

<213> *Rattus norvegicus*

<220>

<221> misc_feature

<223> Incyte ID No.: 700270272H1

<400> 85

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ctgttctctgt gctcccgccg tcgtctctcca gncaggcct cggggtcca gctccggtgt 60
tgggtgcagg cntgntgtgg tctccaaagt gactgaacaa tgcagaagga cagtggccca 120
ctggttcctt tacattatta tggtttcggc tatgcggccc tgggtggctac tgggtgggatt 180
attggctatg caaaagcagg tagtgtgccg tccctggcct gctggactct tctttggggg 240
cctggcaggc ctgggtgcct accagctgtc tcaggacccc aggaa 285

```

<210> 86

<211> 268

<212> DNA

<213> *Rattus norvegicus*

<220>
 <221> misc_feature
 <223> Incyte ID No.: 700628520H1

<400> 86

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ctccagcnca ggcctccggg ctccagctcc ggtgttgggt gcaggcctgg tgtggtctcc 60
aaagtgactg aacaatgcag aaggacagtg gccactgggt tcctttacat tattatgggt 120
tcggctatgc ggccctgggt gctactgggt ggattattgg ctatgcaaaa gcaggtagtg 180
tgccgtccct ggctgctgga ctcttctttg ggggcctggc aggcctgggt gcctaccagc 240
tgtctcagga ccccggaac gtgtgggt 268
```

<210> 87
 <211> 269
 <212> DNA
 <213> Rattus norvegicus

<220>
 <221> misc_feature
 <223> Incyte ID No.: 700534975H1

<400> 87

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tgctccccgc gtcgtctctc agcgcaggcc tccgggctcc agctccgggt ttgggtgcag 60
gcctgggtgtg gtctccaaag tgactgaaca atgcagaagg acagtggctc actgggtcct 120
ttacattant atggtttcgg ctatgcggcc ctgggtggcta ctgggtggat tattggctat 180
gcaaaagcag gtagtgtgcc gtccctggct gctggactct tctttggggg cctggcaggc 240
ctgggtgcct accagctgtc tcaggacc 269
```

<210> 88
 <211> 262
 <212> DNA
 <213> Rattus norvegicus

<220>
 <221> misc_feature
 <223> Incyte ID No.: 700176004H1

<400> 88

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tatgcngccc tgggtggctac tgggtgggatt attggctatg canaagcagg tagtgtgccg 60
tccttggtctg ctggactctt ctttgggggc ctggcaggcc tgggtgccta ccagctgtct 120
caggacccca ggaacgtgtg ggttttctta gctacgtctg ggactttggc tggcattatg 180
gggatgagat tctacaactc tgggaaattt atgctgacag gtttgatcgc gggagccagt 240
ttgctgatgg ttgccaaact tg 262
```

<210> 89
 <211> 349
 <212> DNA
 <213> Rattus norvegicus

<220>
 <221> misc_feature
 <223> Incyte ID No.: 701609236H1

<400> 89

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cgtacgcgca ggcgcgcggt gctctcccggt gcaactctctg gctgagcnng cggactgccc 60
gcctctctaa aacgtcctgt aactgcggtt ccgggagtgg aaacctaaac gcgcgtgcgc 120
ttcttccacg ccacggaaac cgtgcaggcc tgggtgtggc tccaaagtga ctgaacaatg 180
cagaaggaca gtggccactt ggttccttta cattattatg gtttcggcta tgccggccctg 240
gtggctactg gtgggatatt ggctatgcaa aagcagtatg tgccgtccct ggctgctgga 300
ctctcttggg ggctngcagc ctgggtgctaa caactgtctc aganccag 349
```

<210> 90
 <211> 263
 <212> DNA
 <213> Rattus norvegicus

<220>
 <221> misc_feature
 <223> Incyte ID No.: 701473437H1

<400> 90

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tgactgaaca atgcagaagg acagtggccc actgggttcct ttacattatt atgggtttcgg 120
ctatgcggcc ctgggtggcta ctgggtgggat tattggctat gcaaaagcag gtagtggtgcc 180
gtccctgggt gctggactct tctttggggg cctggcaggc ctgggtgcct accagctgtc 240
tcaggacccc aggaacgtgt ggg                                     263
```

<210> 91
 <211> 303
 <212> DNA
 <213> Rattus norvegicus

<220>
 <221> misc_feature
 <223> Incyte ID No.: 701606089H1

<400> 91

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gcgcaggcct ccggggctcc agctccggtg ttgggtgcag gcctgggtgng gtctccaaag 60
tgactgaaca atgcagaagg acgttngccc actggntcct ttacattatt atgggtttcgg 120
ctatgcggcc ctgggtggcta ctgggtgggan tattggctat gcaaaagcag gtagtggtgcc 180
gtccctngct gctggactct tcttngggg cctgncangc ctgggtgcct accagctgtc 240
tcangacccc aggaacgtgt gggttttccn agctacgtct gggatttgnc tggcatatng 300
gga                                     303
```

<210> 92
 <211> 273
 <212> DNA
 <213> Rattus norvegicus

<220>
 <221> misc_feature
 <223> Incyte ID No.: 701736525H2

<400> 92

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taactgtctc gacctctcct ccacaggtgc aggcctggtg tgggtctcaa agtgactgaa 60
caatgcagaa ggacagtggc ccaactggttc ctttacatta ttatggtttc ggctatgcgg 120
ccctggtggc tactggtggg attattggct atgcaaaagc aggtagtgtg ccgtccctgg 180
ctgctggact cttctttggg ggcctggcag gcctgggtgc ctaccagctg tctcaggacc 240
ccaggaacgt gtgggttttc ctagctacgt ctg                                     273
```

<210> 93
 <211> 262
 <212> DNA
 <213> Rattus norvegicus

<220>
 <221> misc_feature
 <223> Incyte ID No.: 701532848H1

<400> 93

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cngccgtent cctccagcgc angcntccgg gctccagctc cgggtgttggg tgcaggcctg 60
gtgtggtctc caaagtgact gaacaatgca gaaggacagt ggcncactgg ttcctttaca 120
```

ttattatggt ttccgctatg cggccctggt ggctactggt gggattattg gctatgcaaa 180
agcaggtagt gtgccgtccc tggctgctgg actcttcttt gggggcctgg caggcctggg 240
tgcctaccag ctgtctcagg ac 262

<210> 94
<211> 247
<212> DNA
<213> Rattus norvegicus

<220>
<221> misc_feature
<223> Incyte ID No.: 700181220H1

<400> 94

aaaacgtcct gtaactgcgg ttccgggagt ggaaacctaa acgcgcgtgc gcttcttcca 60
cgccacggaa accgtgcagg cctggtgtgg tctccaaagt gactgaacaa tgcagaagga 120
cagtggccca ctggttcctt tacattatta tggtttcggc tatgcccggc tgggtggctac 180
tgggtgggatt attggctatg caaaagcagg tagtgtgccg tccctggctg ctggactctt 240
ctttggg 247

<210> 95
<211> 284
<212> DNA
<213> Rattus norvegicus

<220>
<221> misc_feature
<223> Incyte ID No.: 701462707H1

<400> 95

tacacacccg gctcctgacc tctgttctctg tgetccccgc gtcgtcctcc agcgcaggcc 60
tccgggctcc agctccggtg ttgggtgcag gcctgggtgtg gtctccaaag tgactgaaca 120
atgcagaagg acagtggccc actggttctt ttacattatt atggtttcgg ctatgcccgc 180
ctggtggcta ctggtgggat tattggctat gcaaaagcag gtagtgtgcc gtccctggct 240
gctggactct tctttggggg cctggcaggc ctgggtgcct acca 284

<210> 96
<211> 282
<212> DNA
<213> Rattus norvegicus

<220>
<221> misc_feature
<223> Incyte ID No.: 701462863H1

<400> 96

tacacacccg gctcctgacc tctgttctctg tgetccccgc gtcgtcctcc agcgcaggcc 60
tccgggctcc agctccggtg ttgggtgcag gcctgggtgtg gtctccaaag tgactgaaca 120
atgcagaagg acagtggccc actggttctt ttacattatt atggtttcgg ctatgcccgc 180
ctggtggcta ctggtgggat tattggctat gcaaaagcag gtagtgtgcc gtccctggct 240
gctggactct tctttggggg cctggcaggc ctgggtgcct ac 282

<210> 97
<211> 281
<212> DNA
<213> Rattus norvegicus

<220>
<221> misc_feature
<223> Incyte ID No.: 701481465H1

<400> 97

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ttcttaactg ctccgacctc tctccacag gtgcaggcct ggtgtggtct ccaaagtgc 60
tgaacaatgc agaaggacag tggccactg gttcctttac attattatgg ttccggctat 120
gcggcccttg tggctactgg tgggattatt ggctatgcaa aagcaggtag tgtgccgtcc 180
ctgggctgct ggactcttct ttgggggcct ggcaggcctg ggtgcctacc agctgtctca 240
ggaccccagg aacgtgtggg tttcctagc tacgtctggg a 281

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<210> 98

<211> 265

<212> DNA

<213> Rattus norvegicus

<220>

<221> misc_feature

<223> Incyte ID No.: 701308467H1

<400> 98

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tgttctgtg ctcccgcctg cgtcctccag cgcaggcctc cgggctccag ctccgngtt 60
gggtgcaggc ctggtgtggt ctccaaagtg actgaacaat gcagaaggac agtggcccac 120
tggttccttt acattattat ggtttcggt atgcggcctt ggtggctact ggtgggatta 180
ttggctatgc aaaagcagg agtgtgccgt ccctggctgc tggactcttc tttgggggcc 240
tgnagnctgg gtgcctacca gctgt 265

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<210> 99

<211> 291

<212> DNA

<213> Rattus norvegicus

<220>

<221> misc_feature

<223> Incyte ID No.: 701564368H1

<400> 99

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gggggccttg caggcctggg tgccctaccag ctgtctcagg accccaggaa cgtgtggggt 60
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aaatttatgc ctgcaggttt gatcgcgga gccagtttgc tgatggttgc caaacttgga 180
cttagtatgt tgagttcacc ccatccgtag tagccatagt cctgcgtggg ctcgatga 240
gttgacactc tccagtcctc cacattacca cgtgaagag ataagaacag c 291

```

<210> 100

<211> 271

<212> DNA

<213> Rattus norvegicus

<220>

<221> misc_feature

<223> Incyte ID No.: 700533180H1

<400> 100

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caggtagtgt gccgtccctg gctgctggac tttcttttg gggcctggca ggctgggtg 60
gectaccagc tgctctcagg aacccaggga acgtgtgggt tttcctagct acgtctggga 120
ctttggctgg cattatgggg atgagattct acaactctgg gaaatttatg cctgcagggt 180
tgatcgcgga agccagtttg ctgatggttg ccaaacttgg acttagtatg ttgagttcac 240
cccatccgta gtagccatag tctgcgtgg g 271

```

<210> 101

<211> 255

<212> DNA

<213> Rattus norvegicus

<220>
 <221> misc_feature
 <223> Incyte ID No.: 700124647H1

<400> 101

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ccgtcgtcct ccagcncagg cctccgggct ccagctccgg tgttgggtgc aggcctgggtg 60
tgggtctccaa agtgactgaa caatgcagaa ngacagtggc ccactgggtc ctttacatta 120
ttatgggttc ggctatgcgg ccctgggtggc tactgggtggg attattggct atgcaaaagc 180
aggtagtggt ccgtccctgg ctgctggatc ttctttgggg gcctggcagg cctgggtgcc 240
tannagctgt ctcaa 255
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<210> 102
 <211> 297
 <212> DNA
 <213> Rattus norvegicus

<220>
 <221> misc_feature
 <223> Incyte ID No.: 700537020H1

<400> 102

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gccctgggtgg ctactgggtgg gattattggc tatgcaaaaag caggtagtgt gccgtccctg 60
gctgcnggac tcttctttgg gggcctggca ggctgggtg cctacnagct aggcctcagga 120
ccccaggaac gtgtgggttt tctagctac tctggacct nggctggcat tatggggatg 180
agattctaca actctgggaa atttatgcct gcaggtttga tcgcggggagc cagtttgctg 240
atggttgcca aacttggact tagtatgttg agttcaccac atccgtagta gccatag 297
```

<210> 103
 <211> 261
 <212> DNA
 <213> Rattus norvegicus

<220>
 <221> misc_feature
 <223> Incyte ID No.: 700765205H1

<400> 103

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gacctctgtt cctgtgctcc cgcctgctc ctccagcgca ggccctccggg ctccagctcc 60
ggtgttgggt gcaggcctgg tgtgtctcc aaagtgactg aacaatgcag aaggacagtg 120
gccactgggt tctttacat tattatggtt tcggctatgc ggccctgggt gctactgggt 180
ggattattgg ctatgcaaaa gcaggtagtg tgccgtccct ggctgctgga ctcttctttg 240
ggggcctggc aggcctgggtg c 261
```

<210> 104
 <211> 312
 <212> DNA
 <213> Rattus norvegicus

<220>
 <221> misc_feature
 <223> Incyte ID No.: 701942992H1

<400> 104

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cgacgtctac ncaccgggct cctgacctct gttcctgtgc tcccggcgtc gtccctccagc 60
gcaggcctcc gggctccagc tccggtgttg ggtgcaggcc tgggtgggtc tccaaagtna 120
ctgaacaatg cagaaggaca gtggcccaact ggttccttta cattattatg gtttcggcta 180
tcgggccctg gtggctactg gtgggattat tggctatgca aaagcaggta gtgtgccgtc 240
cctggctgct ggactcttct ttgggggcct ggcagcctgg ggctacaag tttntcagg 300
nccaggnan nt 312
```

<210> 105
 <211> 241
 <212> DNA
 <213> Rattus norvegicus

<220>
 <221> misc_feature
 <223> Incyte ID No.: 701197694H1

<400> 105

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tgcctccgcc gtcgtcctcc agcgcaggcc tccgggctcc agctccgggtg ttgggtgcag 60
gcctgggtgtg gtctccaaag tgactgaaca atgcagaagg acagtggccc actgggtcct 120
ttacattatt atggtttcgg ctatgcggcc ctggtggcta ctggtgggat tattggctat 180
gcaaaagcag gaacgtgtgg gttttcctag ctacgtctgg gactttggct ggnattatgg 240
g                                                    241

```

<210> 106
 <211> 268
 <212> DNA
 <213> Rattus norvegicus

<220>
 <221> misc_feature
 <223> Incyte ID No.: 701024952H1

<400> 106

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cccggctcct gacctctgtt cctgtgctcc cgcgctcgtc ctccagcgca ggcctccggg 60
ctccagctcc ggtgttgggt gcaggcctgg tgtggtctcc aaagtgactg nacaatgcag 120
aaggncagtg gccactggt tcctttacat tattatggtt tcggctatgc ggcctgggtg 180
gctactgggtg ggattattgg ctatgcaaaa gcaggtagtg tgccgtccct ggctgctgga 240
ctctnctttn ggggcctggc aggcttag                                                    268

```

<210> 107
 <211> 318
 <212> DNA
 <213> Rattus norvegicus

<220>
 <221> misc_feature
 <223> Incyte ID No.: 701582676H1

<400> 107

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gcctaccagc tgtctcagga cccaggaac gtgtggggtt tcctagctac gtctgggact 60
ttggctggca ttatggggat gagattctac aactctggga aatttatgcc tgcaggtttg 120
atcgcgggag ccagtttgct gatggttgcc aaacntggac ttagtatggt gaggtcaccc 180
catccgtagt agccatagtc ctgcgtgggc tcatgatgag ttgacactct ccagtcctcc 240
acattaccac gctgaagaga taagaacagc aaagacctac actgagcaca tggaggcgaa 300
gacgtgggta ctatagtg                                                    318

```

<210> 108
 <211> 255
 <212> DNA
 <213> Rattus norvegicus

<220>
 <221> misc_feature
 <223> Incyte ID No.: 701293154H1

<400> 108

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ggattattgg ctattgcaaa agcaggtaag tgtgccgtcc ctggtgctg gactcttctt 60
tgggggcctg gcaggcctgg gtgcctacca gctgtctcag gacccagga acgtgtgggt 120

```


tttcctagct acgtctggga ctttggcttg cattatgggg atgagattct acaactctgg 180
 gaaatttatg cctgcaggtt tgatcgcggg agccagtttg ctgatgggtg ccaaacttgg 240
 attagtatgt tgagg 255

<210> 109
 <211> 254
 <212> DNA
 <213> Rattus norvegicus

<220>
 <221> misc_feature
 <223> Incyte ID No.: 701298824H1

<400> 109

catgcgcagg cctccgggct ccatgctcgg gtgttgggtg catggcctgg tngggtctcc 60
 aaagngactg aacaatgcag aaggacagtg gccactgggt tcctttacat tattatggnt 120
 tcggctatgc ggccctgggt gctactgggt ggattattgg ctatgcaaaa gcnngtagtg 180
 tgccgccttg gctgctggac tcttctttgg gggcctgcag nctgggtgcc taccagctgt 240
 ctcaggaccc agga 254

<210> 110
 <211> 294
 <212> DNA
 <213> Rattus norvegicus

<220>
 <221> misc_feature
 <223> Incyte ID No.: 700524204H1

<400> 110

tcaggacccc aggaacgtgt gggttttcct agctacgtct gggactttgg ctggcattat 60
 ggggatgaga ttctacaact ctgggaaatt tatgcctgca ggtttgatcg cgggagccag 120
 ttgtctgatg gttgccaaac ttggacttag tatgttgagt tcaccccatc cgtagtagcc 180
 atagccctgc gtgggctcat gatgagttga cactctccag tcctctacat taccacgctg 240
 aagagataag aacagcaaag acctacactg agcacatgga ggcgaagagt gggt 294

<210> 111
 <211> 289
 <212> DNA
 <213> Rattus norvegicus

<220>
 <221> misc_feature
 <223> Incyte ID No.: 700067537H1

<400> 111

gacgtctaca caccgggtc ctgacctctg ttctgtgct cccgccgtcg tcctccagcg 60
 caggcctcgg ggctccagct ccgctgttgg gtgcaggcct ggtgtggtct ccaaagtgc 120
 tgaacaatgc agaaggacag tggccactg gtccctttac attattatgg ttccggctat 180
 gcggccctgg tggctactgg tgggattatt ggctatgcaa aagcagtagt gtgccgtccc 240
 tggctgctgg atcttctttg ggggctggca ggctgggtgc ctacaactg 289

<210> 112
 <211> 276
 <212> DNA
 <213> Rattus norvegicus

<220>
 <221> misc_feature
 <223> Incyte ID No.: 701258019H1

<400> 112

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tggttcctgtg cteccgcggt cgtccctccag cgcaggcctc cgggctccag ctccgggtgtt 60
gggtgcaggc ctggtgtggt ctccaaagtg actgaacaat gcatgaagga cagttggccc 120
actggttcct ttacattatt atggnntccg gctatgcggc cctgggtggct actggtgnga 180
ttattggcta tgcaaaagca ggtagtgtgc cgccctggct gctggactct tctttggggg 240
cctgcagnct ggtgcctacc agctgctctg cgtngg 276

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<210> 113

<211> 254

<212> DNA

<213> Rattus norvegicus

<220>

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<223> Incyte ID No.: 700532493H1

<400> 113

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tttgctgatg gttgccaaac ttggacttag tatgttgagt tcaccccatc cytagtagcc 180
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gagatanaac agca 254

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<210> 114

<211> 282

<212> DNA

<213> Rattus norvegicus

<220>

<221> misc_feature

<223> Incyte ID No.: 700523302H1

<400> 114

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gggtanangn ncccgnnnng nnaagggggg atnttgnnt acgnaagagc ngntagtgtg 180
cggtccctgg ctgctggact cttctttggg ggcctggcag gcctgggtgc ctaccagctg 240
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<210> 115

<211> 256

<212> DNA

<213> Rattus norvegicus

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gcagaaggac agtggccac tgggtccttt acattattat ggtttcggct atgcggccct 180
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<210> 116

<211> 244

<212> DNA

<213> Rattus norvegicus

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tgctaccag ctgctcagga ccccaggaac gtgtgggttt tcctagctac gtctgggact 180
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<400> 117

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gaacaatgca gaaggacagt ggcccaactgg ttccctttaca ttattatggt ttcggctatc 180
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<210> 118
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 <212> DNA
 <213> Rattus norvegicus

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 <223> Incyte ID No.: 700493358H1

<400> 118

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tatgcaaaag caggtagtgt gccgtccctg gctgctggac tcttctttgg ggncttggca 180
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<210> 119
 <211> 265
 <212> DNA
 <213> Rattus norvegicus

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tgcccaactg gttcctttac attattatgg ttctggctat gcggccctgg tggctactgg 180
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gggggcctgg caggcctggg tgcct                                         265
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 <212> DNA
 <213> Rattus norvegicus

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<400> 120

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gtgactgaac aatgcagaag gacagtggcc cactggttcc ttacattat tatgggttcg 180
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gctccct 247
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<210> 121
 <211> 263
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 <213> Rattus norvegicus

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<400> 121

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tgaacaatgc agaaggacag tggccactg gtccctttac attattatgg ttccggctat 180
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<220>
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<400> 122

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tgntgggatt attggtatn caaaagcagg tagtgtncg tccctggctg ctggactctt 180
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<210> 123
 <211> 343
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 <213> Rattus norvegicus

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 <221> misc_feature
 <223> Incyte ID No.: 701582848H1

<400> 123

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gcgggagcca nttgctgata gttgccaact tngacttagt atgttgagtn caccatcc 180
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gtagtagcat ancctgcgtg ggctcagatg agtnacactc tccaggcctc cacatttacc 240
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<210> 124
 <211> 241
 <212> DNA
 <213> Rattus norvegicus

<220>
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 <223> Incyte ID No.: 701305531H1

<400> 124

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 tgaacaatgc agaaggacag tggcccactg gttcctttac attattatgg attcggctat 180
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 g 241

<210> 125
 <211> 155
 <212> DNA
 <213> Rattus norvegicus

<220>
 <221> misc_feature
 <223> Incyte ID No.: 700916103H1

<400> 125

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 ttacattat tatggtttcg gctatgcggc cctgg 155

<210> 126
 <211> 185
 <212> DNA
 <213> Rattus norvegicus

<220>
 <221> misc_feature
 <223> Incyte ID No.: 701294764H1

<400> 126

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 agcgg 185

<210> 127
 <211> 125
 <212> DNA
 <213> Rattus norvegicus

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 <221> misc_feature
 <223> Incyte ID No.: 700066710H1

<400> 127

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aactc 125

<210> 128
<211> 266
<212> DNA
<213> Rattus norvegicus

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<221> misc_feature
<223> Incyte ID No.: 701471559H1

<400> 128

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gacactctcc agtcctctac attaccacgc tgaagagata agaacagcaa agacctacac 180
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<211> 208
<212> DNA
<213> Rattus norvegicus

<220>
<221> misc_feature
<223> Incyte ID No.: 700325006H1

<400> 129

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<210> 130
<211> 263
<212> DNA
<213> Rattus norvegicus

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<221> misc_feature
<223> Incyte ID No.: 701258479H1

<400> 130

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<210> 131
<211> 258
<212> DNA
<213> Rattus norvegicus

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<400> 131

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ttgacactct ccagtcctcc acattaccac gctgaagaga taagaacagc aaagacctac 180
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<210> 132

<211> 272

<212> DNA

<213> Rattus norvegicus

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<223> Incyte ID No.: 701246066H1

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<210> 133

<211> 253

<212> DNA

<213> Rattus norvegicus

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<221> misc_feature

<223> Incyte ID No.: 700594190H1

<400> 133

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acgtgggttac tatagtgacc gttcagagac ggcgagtgtc tgacctcaga gctcacactg 180
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<220>

<221> misc_feature

<223> Incyte ID No.: 700627108H1

<400> 134

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tgnnttgaac actctccagt cagtccagat naacgncgct gntagagatn aagaccagcn 180
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acggcgngtg tntggatcan agatcca 267
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<210> 135

<211> 650

<212> DNA

<213> Rattus norvegicus

<220>

<223> RnAUG.conN

<400> 135

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<210> 136

<211> 114

<212> PRT

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<220>

<223> RnAUG.conP

<400> 136

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 35          40          45
Gly Gly Leu Ala Gly Leu Gly Ala Tyr Gln Leu Ser Gln Asp Pro
 50          55          60
Arg Asn Val Trp Val Phe Leu Ala Thr Ser Gly Thr Leu Ala Gly
 65          70          75
Ile Met Gly Met Arg Phe Tyr Asn Ser Gly Lys Phe Met Pro Ala
 80          85          90
Gly Leu Ile Ala Gly Ala Ser Leu Leu Met Val Ala Lys Leu Gly
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Leu Ser Met Leu Ser Ser Pro His Pro
110

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<210> 137

<211> 223

<212> DNA

<213> Homo sapiens

<220>

<221> misc_feature

<223> Incyte ID No.: 746355H1

<400> 137

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gggaatgagg ttctaccact ctggaaaatt catgcctgca ggt 223

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<210> 138

<211> 243

<212> DNA

<213> Homo sapiens

<220>

<221> misc_feature

<223> Incyte ID No.: 1294663H1

<400> 138

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aggattaaaa atctgcatct tccactatct tcaatgtatt aagagaaata agtgcagcat 180
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aat 0 243
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(19) World Intellectual Property Organization
International Bureau



(43) International Publication Date
25 May 2001 (25.05.2001)

PCT

(10) International Publication Number
WO 01/36684 A3

(51) International Patent Classification⁷: **C12Q 1/68**

(21) International Application Number: PCT/US00/31743

(22) International Filing Date:
16 November 2000 (16.11.2000)

(25) Filing Language: English

(26) Publication Language: English

(30) Priority Data:
09/443,184 19 November 1999 (19.11.1999) US

(63) Related by continuation (CON) or continuation-in-part (CIP) to earlier application:
US 09/443,184 (CIP)
Filed on 19 November 1999 (19.11.1999)

(71) Applicant (for all designated States except US): **INCYTE GENOMICS, INC.** [US/US]; 3160 Porter Drive, Palo Alto, CA 94304 (US).

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Lois Avenue, Sunnyvale, CA 94087 (US). **BAUGHN, Mariah, R.** [US/US]; 14244 Santiago Road, San Leandro, CA 94577 (US). **AZIMZAI, Yalda** [US/US]; 5518 Boulder Canyon Drive, Castro Valley, CA 94552 (US). **LAL, Preeti** [IN/US]; 2382 Lass Drive, Santa Clara, CA 95054 (US).

(74) Agents: **TURNER, Christopher** et al.; 3160 Porter Drive, Palo Alto, CA 94304 (US).

(81) Designated States (*national*): AE, AG, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, BZ, CA, CH, CN, CR, CU, CZ, DE, DK, DM, DZ, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MA, MD, MG, MK, MN, MW, MX, MZ, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, TZ, UA, UG, US, UZ, VN, YU, ZA, ZW.

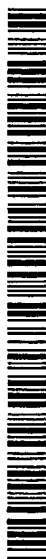
(84) Designated States (*regional*): ARIPO patent (GH, GM, KE, LS, MW, MZ, SD, SL, SZ, TZ, UG, ZW). Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM). European patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE, TR). OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, GW, ML, MR, NE, SN, TD, TG).

Published:

— with international search report

(88) Date of publication of the international search report:
14 March 2002

For two-letter codes and other abbreviations, refer to the "Guidance Notes on Codes and Abbreviations" appearing at the beginning of each regular issue of the PCT Gazette.



WO 01/36684 A3

(54) Title: MAMMALIAN TOXICOLOGICAL RESPONSE MARKERS

(57) Abstract: The present invention relates to mammalian nucleic acid and protein molecules comprising a plurality of nucleic acid and protein molecules. The mammalian nucleic acid molecules can be used as hybridizable array elements in a microarray in diagnostic and therapeutic applications including detecting metabolic and toxicological responses, and in monitoring drug mechanism of action. The protein molecules can be used in a pharmaceutical composition. The present invention also relates to methods for screening compounds and therapeutics for metabolic responses indicative of a toxic compound or molecule.

INTERNATIONAL SEARCH REPORT

International Application No
PCT/US 00/31743

A. CLASSIFICATION OF SUBJECT MATTER

IPC 7 C12Q1/68

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC 7 C12Q

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

EPO-Internal, SEQUENCE SEARCH, WPI Data, CHEM ABS Data, MEDLINE, BIOSIS, EMBASE

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	<p>DATABASE EMBEST [Online] EMBL, Heidelberg; Accession Number: RS2274, XP002172701 abstract</p> <p>---</p> <p>-/--</p>	12

☒ Further documents are listed in the continuation of box C.

☒ Patent family members are listed in annex.

* Special categories of cited documents :

- "A" document defining the general state of the art which is not considered to be of particular relevance
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- "L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)
- "O" document referring to an oral disclosure, use, exhibition or other means
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- "Z" document member of the same patent family

Date of the actual completion of the international search

24 July 2001

Date of mailing of the international search report

19. 8. 01

Name and mailing address of the ISA

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INTERNATIONAL SEARCH REPORT

International Application No
PCT/US 00/31743

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT		
Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	LEE N H ET AL: "COMPARATIVE EXPRESSED-SEQUENCE-TAG ANALYSIS OF DIFFERENTIAL GENE EXPRESSION PROFILES IN PC-12 CELLS BEFORE AND AFTER NERVE GROWTH FACTOR TREATMENT" PROCEEDINGS OF THE NATIONAL ACADEMY OF SCIENCES OF USA, NATIONAL ACADEMY OF SCIENCE. WASHINGTON, US, vol. 92, no. 18, 29 August 1995 (1995-08-29), pages 8303-8307, A, B, XP002033656 ISSN: 0027-8424 the whole document	12
Y	--- WO 97 13877 A (LYNX THERAPEUTICS INC ; MARTIN DAVID W (US)) 17 April 1997 (1997-04-17) the whole document	1-10, 13, 16-20
Y	--- WO 99 23254 A (AFFYMETRIX INC ; NAIR ARCHANA (US); LOCKHART DAVID J (US); WARRINGT) 14 May 1999 (1999-05-14) the whole document -----	1-10, 13, 16-20

INTERNATIONAL SEARCH REPORT

International application No.
PCT/US 00/31743

Box I Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)

This International Search Report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

1. ☐ Claims Nos.:
because they relate to subject matter not required to be searched by this Authority, namely:
2. ☒ Claims Nos.: 11 (partially), 14 and 15 (both completely)
because they relate to parts of the International Application that do not comply with the prescribed requirements to such an extent that no meaningful International Search can be carried out, specifically:
see FURTHER INFORMATION sheet PCT/ISA/210
3. ☐ Claims Nos.:
because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

Box II Observations where unity of invention is lacking (Continuation of item 2 of first sheet)

This International Searching Authority found multiple inventions in this international application, as follows:

see additional sheet

1. ☐ As all required additional search fees were timely paid by the applicant, this International Search Report covers all searchable claims.
2. ☐ As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
3. ☐ As only some of the required additional search fees were timely paid by the applicant, this International Search Report covers only those claims for which fees were paid, specifically claims Nos.:
4. ☒ No required additional search fees were timely paid by the applicant. Consequently, this International Search Report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:
1-13, 16-20 all partially

Remark on Protest

- ☐ The additional search fees were accompanied by the applicant's protest.
☐ No protest accompanied the payment of additional search fees.

FURTHER INFORMATION CONTINUED FROM PCT/ISA/ 210

This International Searching Authority found multiple (groups of) inventions in this international application, as follows:

1. Claims: 1-13,16-20 (all partially)

A method for detecting or diagnosing the effect of a toxic compound comprising the use of a microarray having immobilised thereon a plurality of probes, one being SEQ ID 1, a nucleic acid molecule having the sequence SEQ ID 1 or fragments and variants thereof, protein encoded thereby.

2. Claims: 1-13,16-20 (all partially)

A method for detecting or diagnosing the effect of a toxic compound comprising the use of a microarray having immobilised thereon a plurality of probes, one being SEQ ID 2, a nucleic acid molecule having the sequence SEQ ID 2 or fragments and variants thereof, protein encoded thereby.

3.-30. ...As above, for SEQ ID 4-11, 17-33, 36, 39 and 41

FURTHER INFORMATION CONTINUED FROM PCT/ISA/ 210

Continuation of Box 1.2

Claims Nos.: 11 (partially), 14 and 15 (both completely)

1

The applicant's attention is drawn to the fact that claims, or parts of claims, relating to inventions in respect of which no international search report has been established need not be the subject of an international preliminary examination (Rule 66.1(e) PCT). The applicant is advised that the EPO policy when acting as an International Preliminary Examining Authority is normally not to carry out a preliminary examination on matter which has not been searched. This is the case irrespective of whether or not the claims are amended following receipt of the search report or during any Chapter II procedure.

INTERNATIONAL SEARCH REPORT

Information on patent family members

International Application No

PCT/US 00/31743

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